

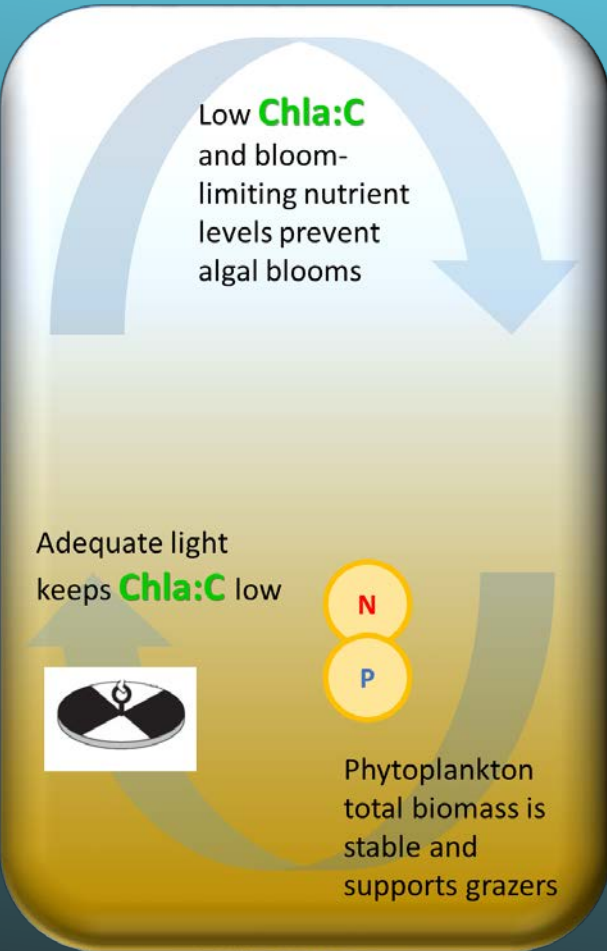
A Phytoplankton Conceptual Diagram and Some Phytoplankton-Water Clarity Relationships

Claire Buchanan (ICPRB)

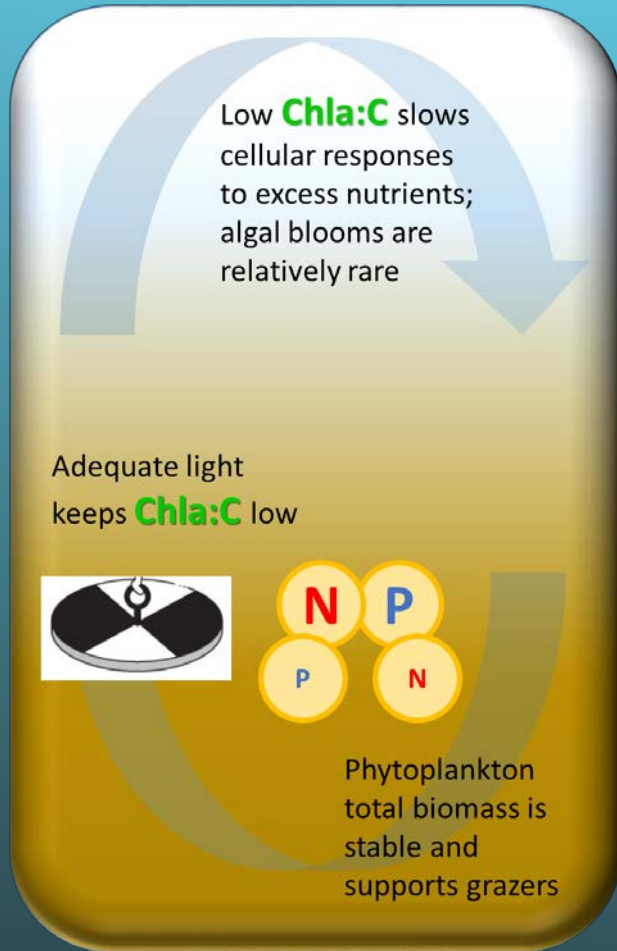
May 2-3, 2017

Water Clarity Workshop II

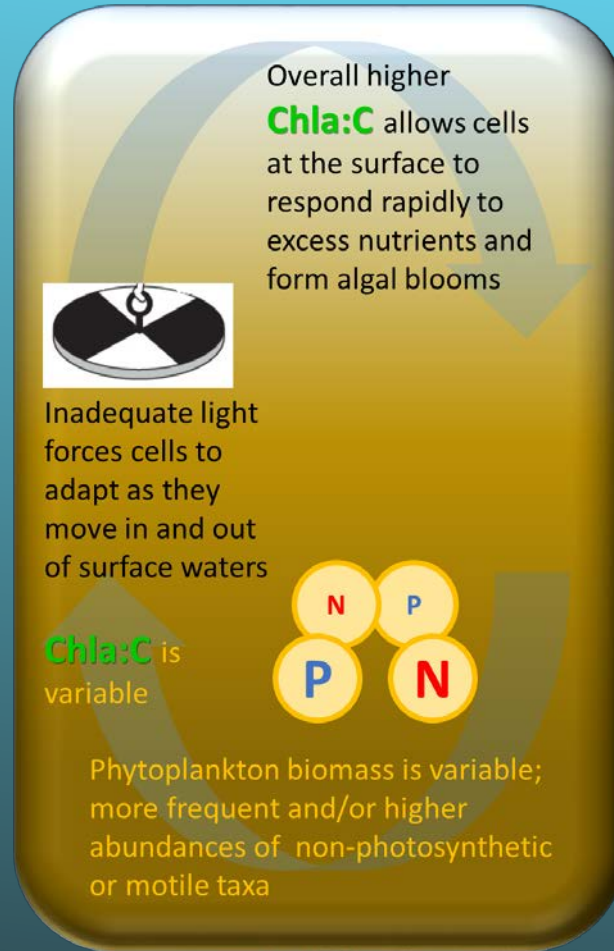
Phytoplankton Conceptual Diagram



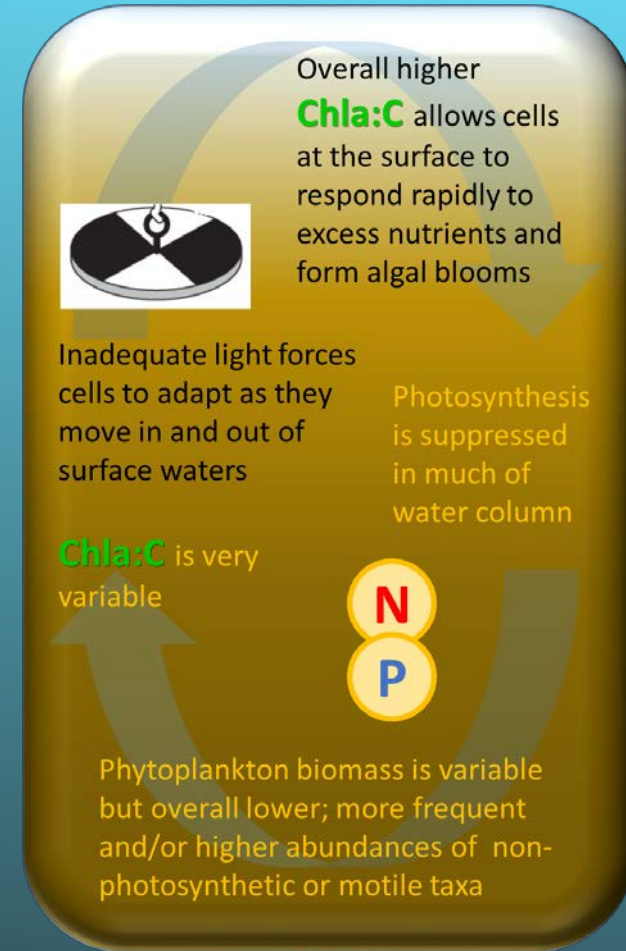
Reference
Adequate clarity *and* bloom-limiting DIN *and* bloom-limiting PO₄



Mixed Better Light
Adequate clarity but DIN *and/or* PO₄ is excess



Mixed Poor Light
Inadequate clarity but DIN *and/or* PO₄ is bloom-limiting

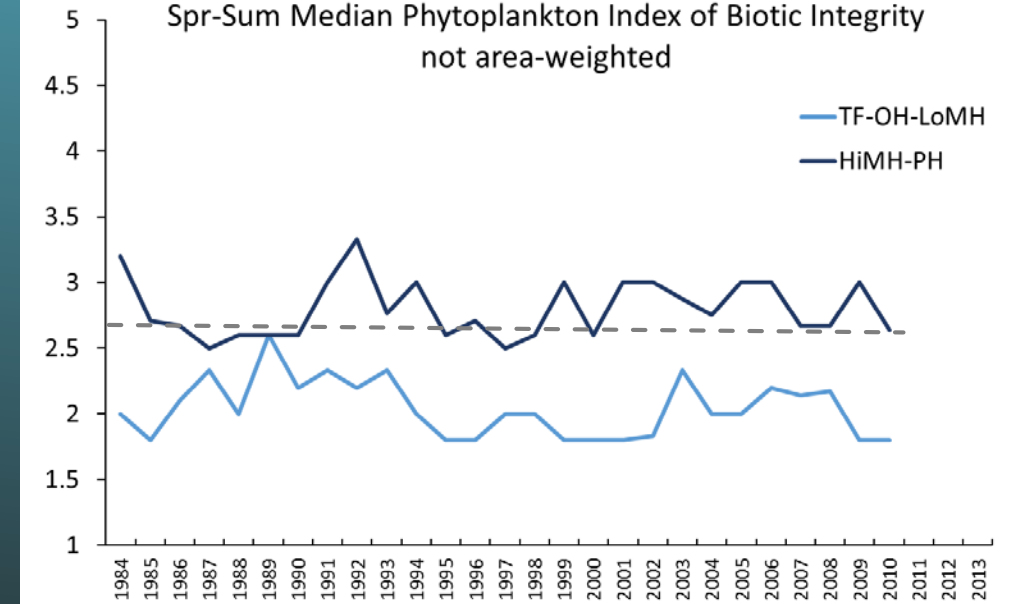
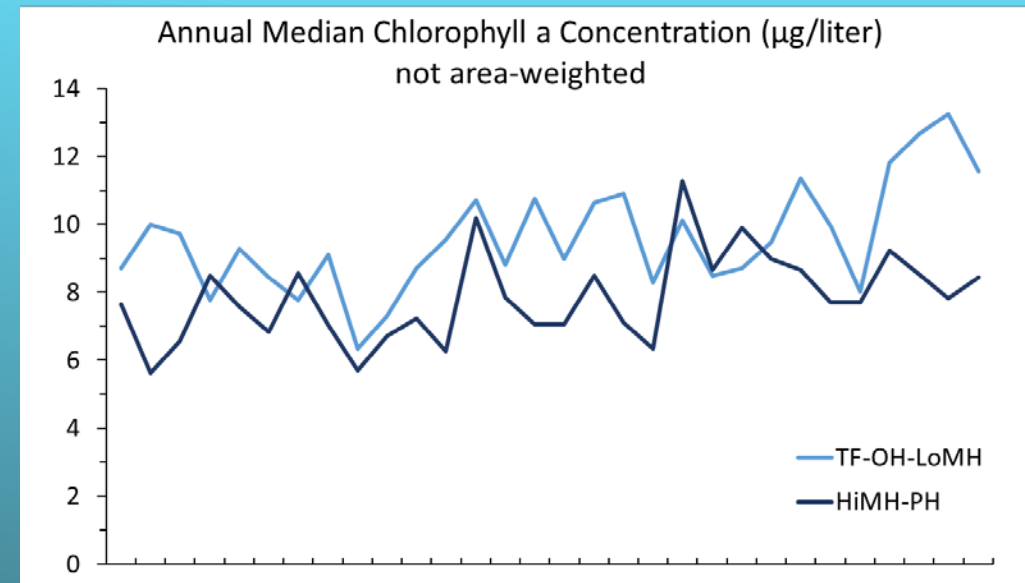
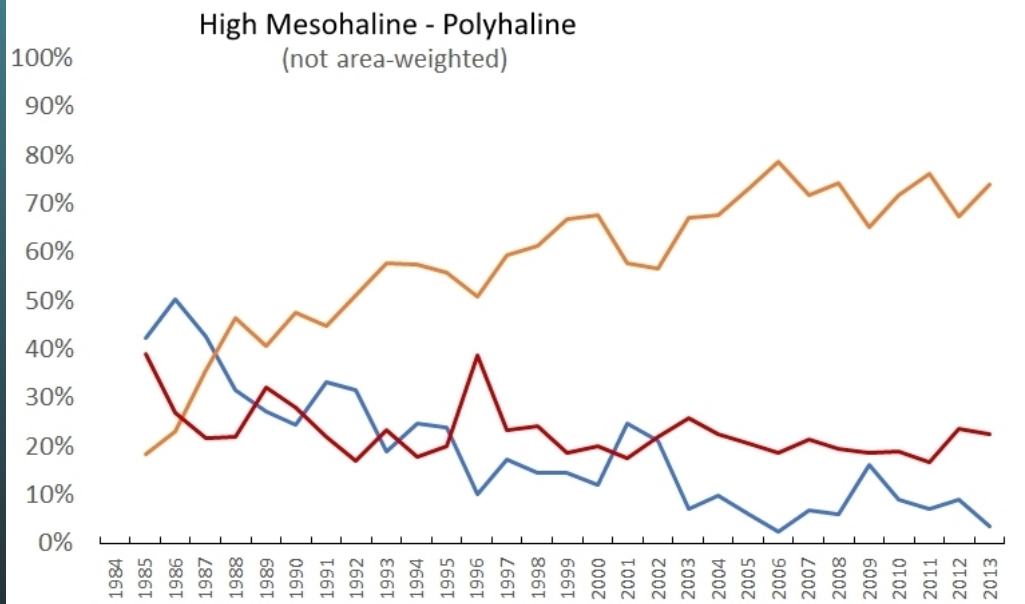
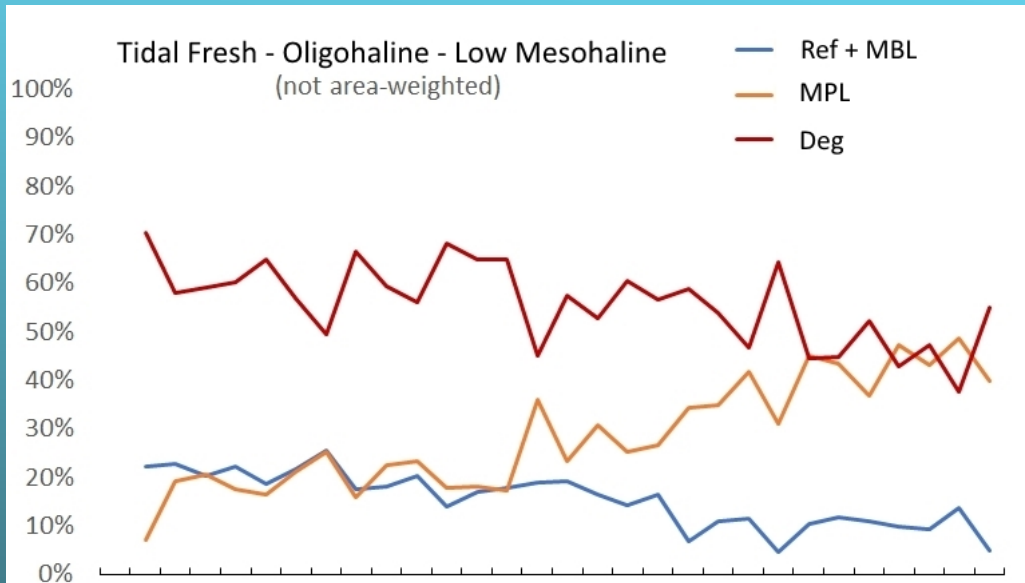


Degraded
Inadequate clarity *and* excess DIN *and* excess PO₄

Phytoplankton Conceptual Diagram

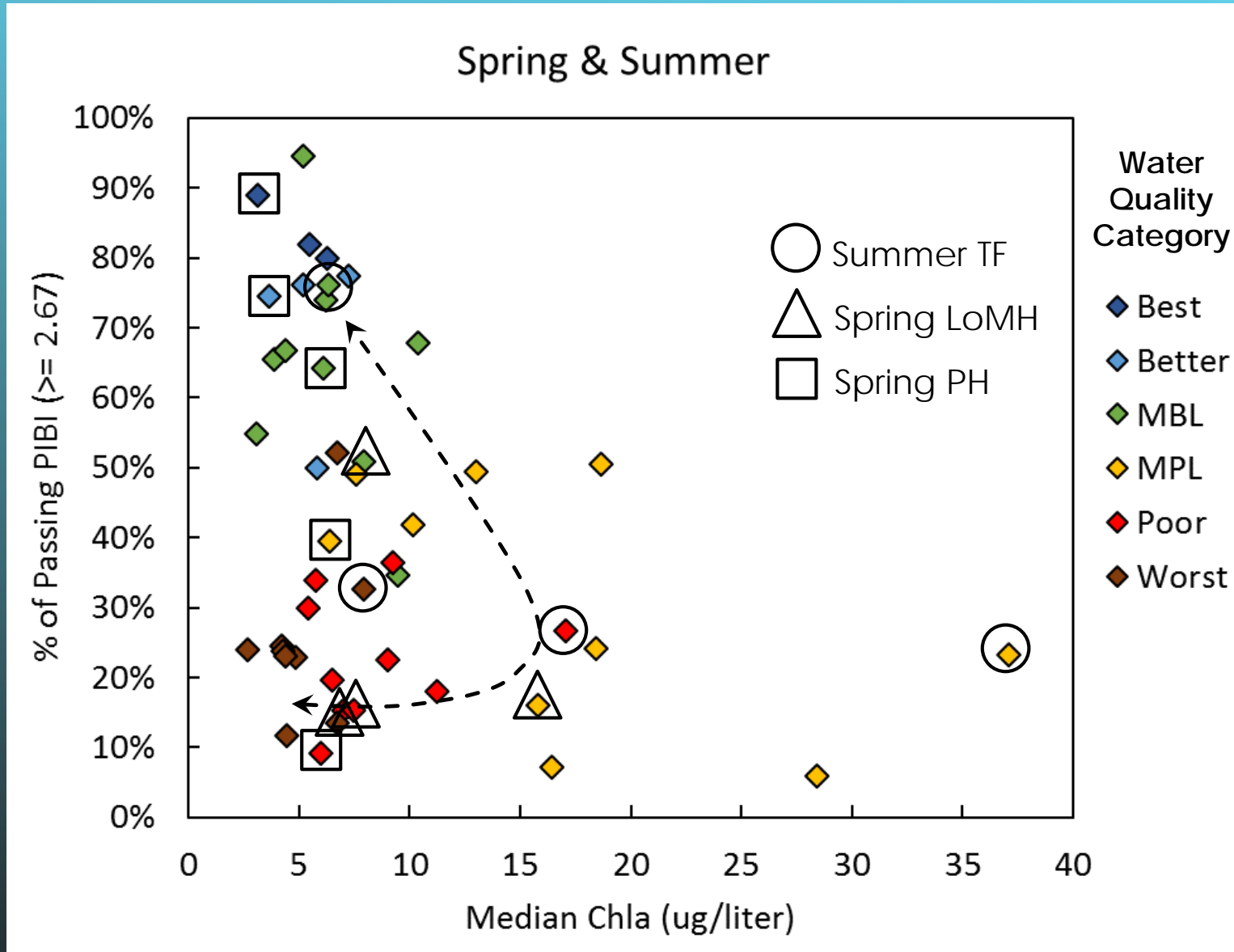
Light Requirements (Secchi Depth, m)

Salinity Zone	Phytoplankton (seasonal)	SAV (1m & 2m application depth)	
Tidal Fresh	0.7 – 0.9	0.7	1.4
Oligohaline	0.7 – 0.9	0.7	1.4
Low Mesohaline	1.2 – 1.4	1.0	1.9
High Mesohaline	1.4 – 2.0	1.0	1.9
Polyhaline	1.6 – 2.1	1.0	1.9



PIBI correctly ID's REF and DEG 70% - 84%

Phytoplankton Recovery (Degradation) Trajectory?



Water Clarity \leftrightarrow Phytoplankton

Photosynthesis
requires photons

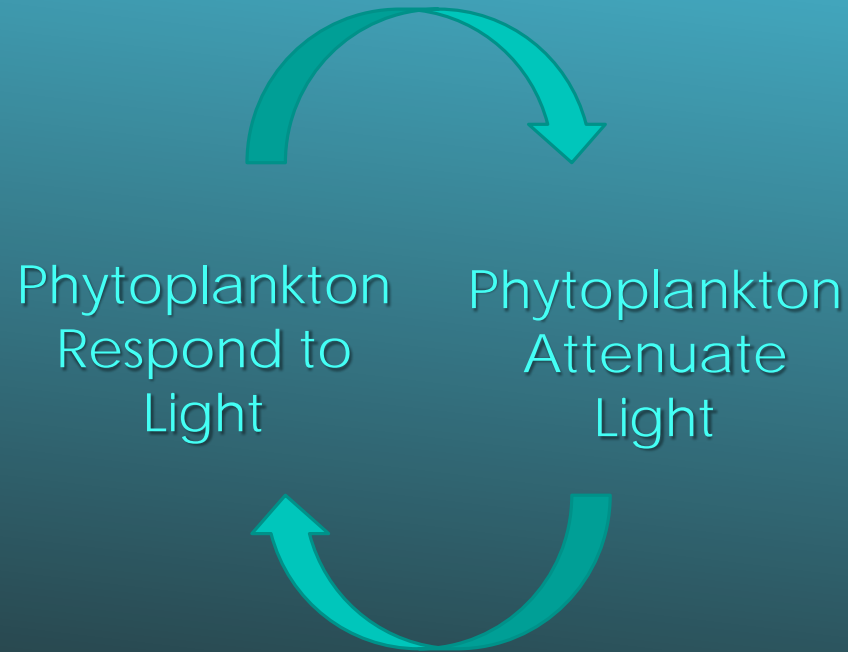


Phytoplankton cells
adjust photopigment
levels to capture
sufficient photons

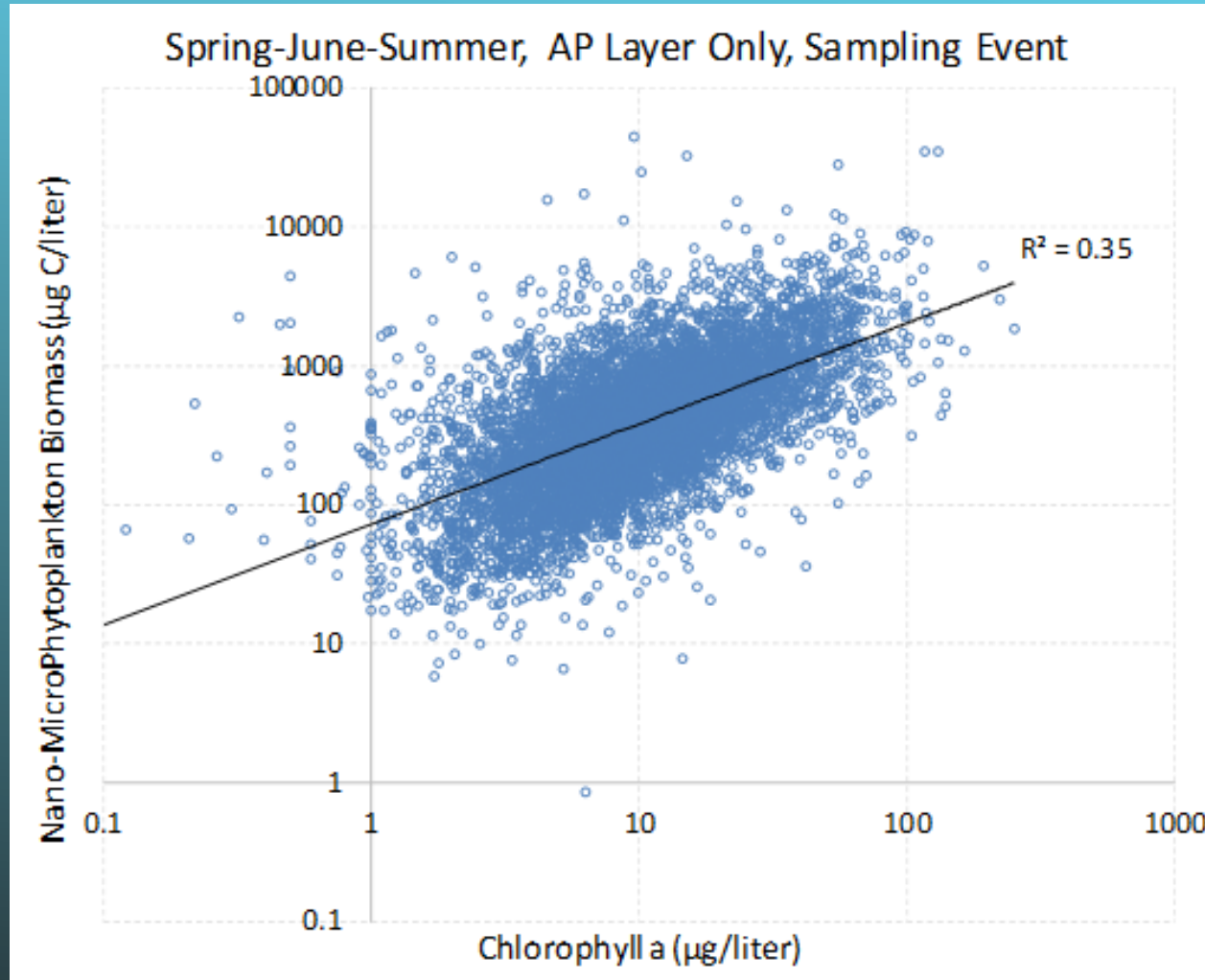


Cells adapt or die in
poor light environments

- Mixotrophy
- Buoyancy/motility



Phytoplankton Chlorophyll as a Surrogate for Biomass

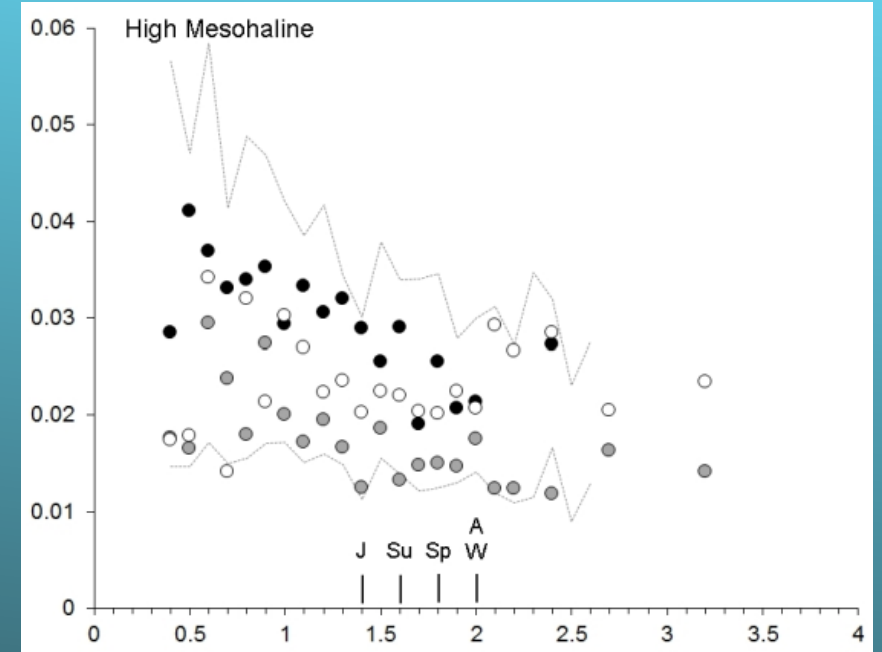
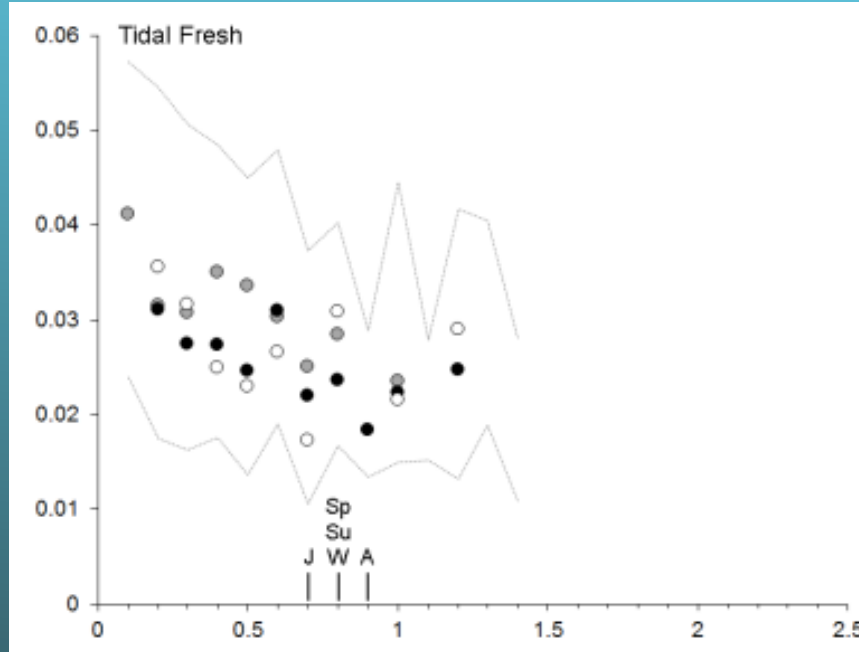


...not so good!

Cell Adaptation in Low Light

Chla : PhytoC

- Spring
- Summer
- Autumn-Winter
- 25th – 75th %ile



Secchi Depth (m) →

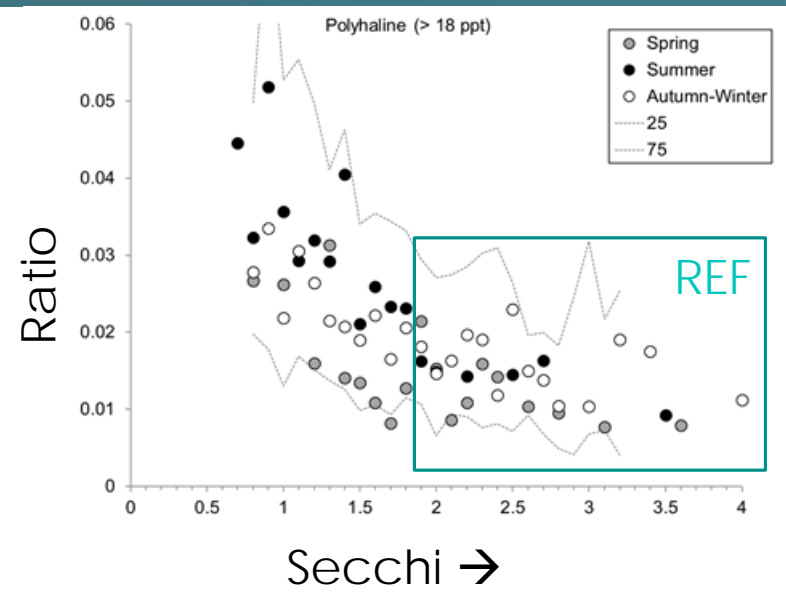
$$\text{Chla : PhytoC} = \frac{\text{Chlorophyll } a \text{ (}\mu\text{g/liter)}}{\text{Total Nano-Micro-Phytoplankton Biomass (}\mu\text{g C/liter)}}$$

$$\text{Pheo : PhytoC} = \frac{\text{Pheophytin (}\mu\text{g/liter)}}{\text{Total Nano-Micro-Phytoplankton Biomass (}\mu\text{g C/liter)}}$$

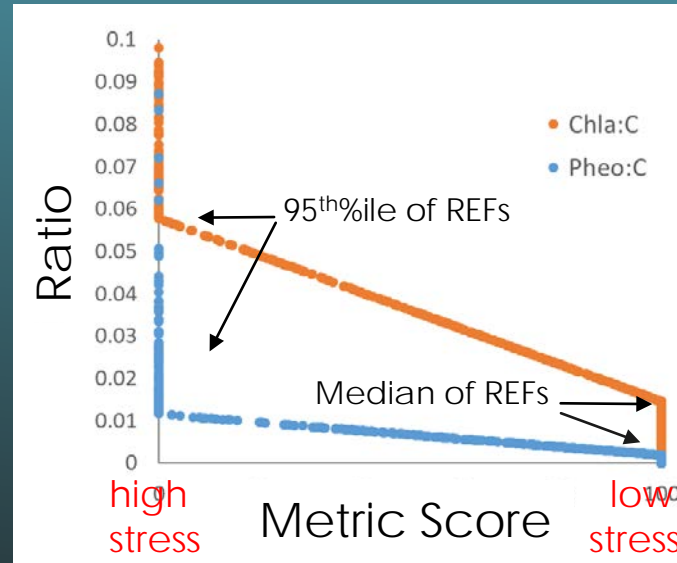
Cell Adaptation to Low Light

Indicator of low-light stress

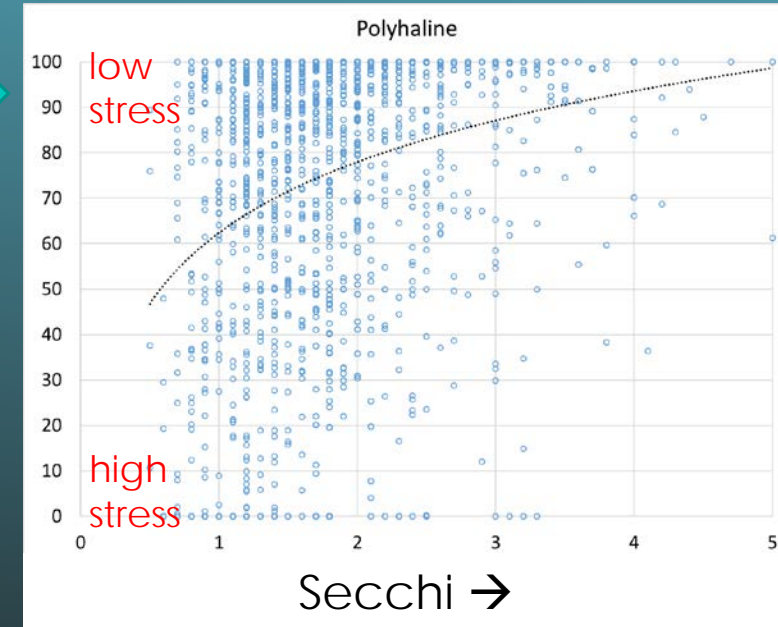
Ratios vs
Secchi depth



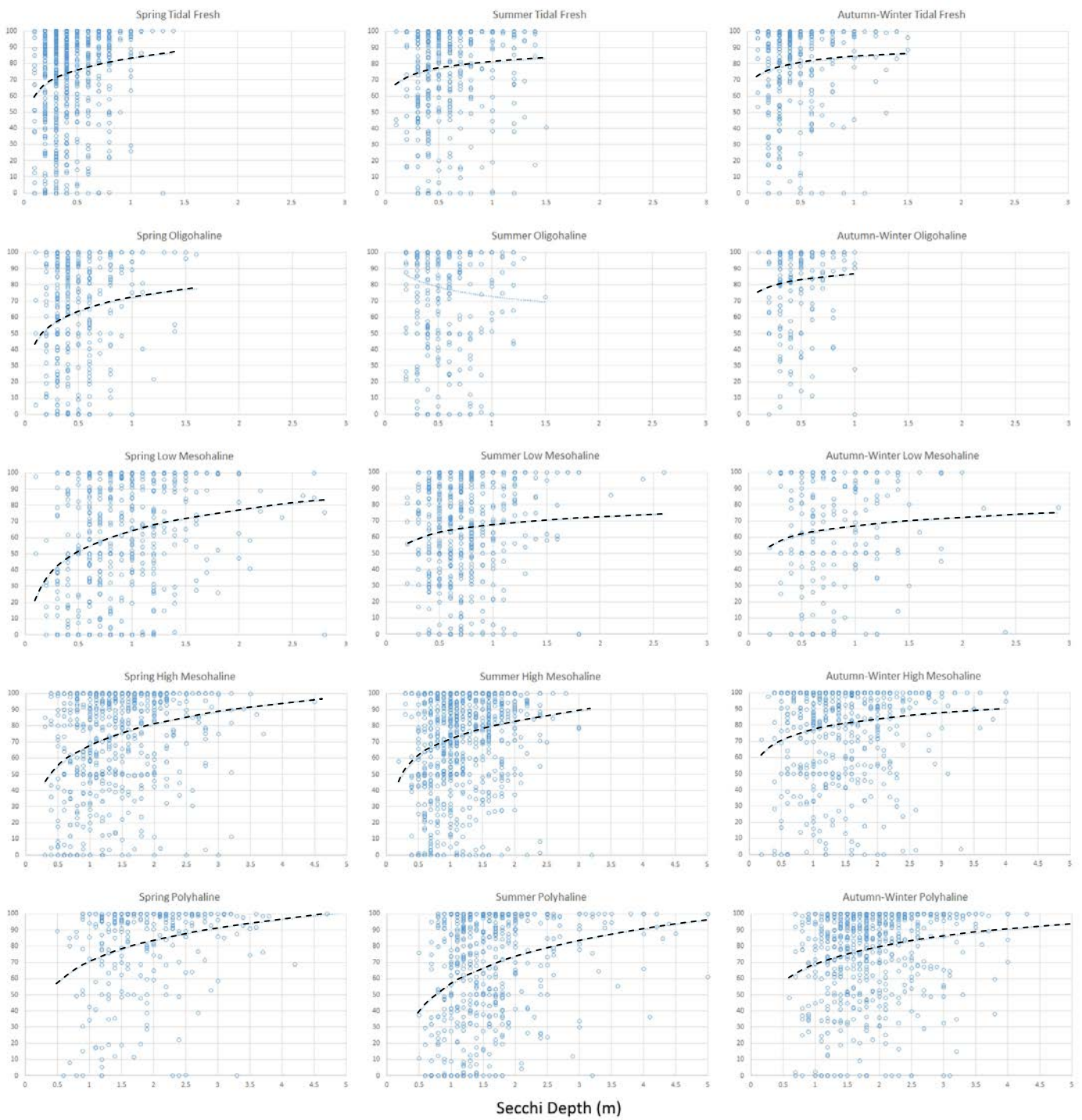
Ratios converted to unitless
scores and averaged



Indicator score vs
Secchi depth

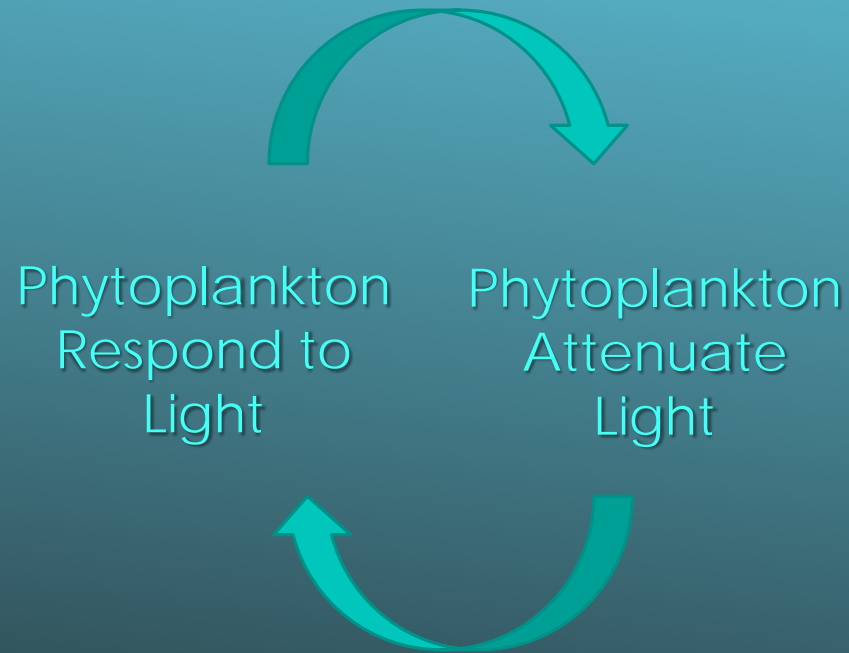


Phytoplankton Indicator of Low-Light Stress



As Secchi depth improves, variability in cellular photopigment levels decreases and stress due to low-light conditions is minimized

Phytoplankton ↔ Water Clarity



Phytoplankton attenuate light in several ways



- Living cells leak DOM – leak faster if stressed by
- Low light levels
 - High cell densities
 - Viruses & parasitic fungi
 - Grazers
 - Salinity



Living cell structure is one component of TSS and mostly scatters light

Phytoplankton Biomass and DOC

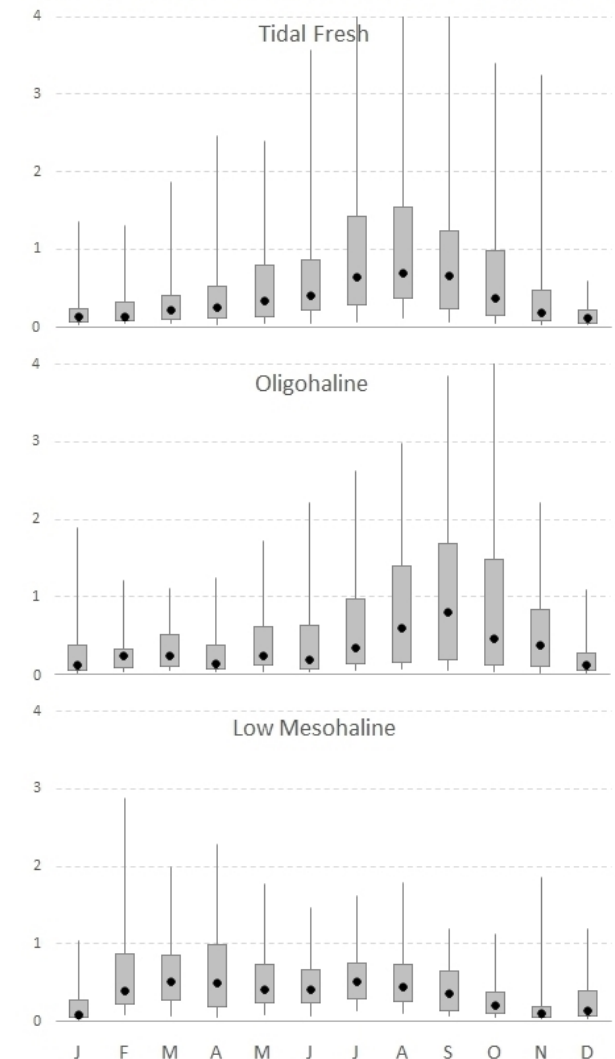
Biomass

DOC

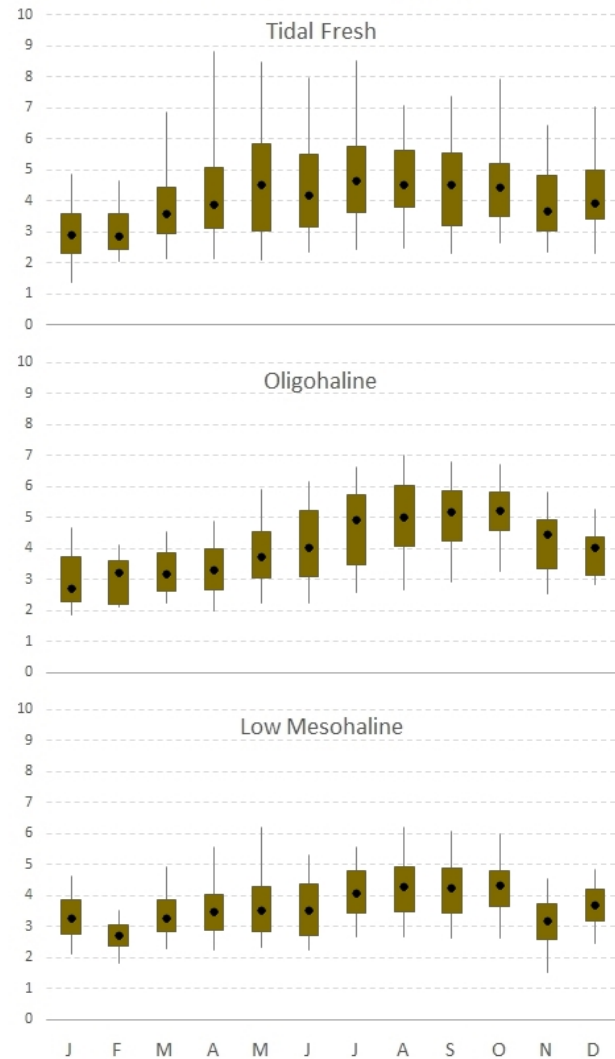
Biomass

DOC

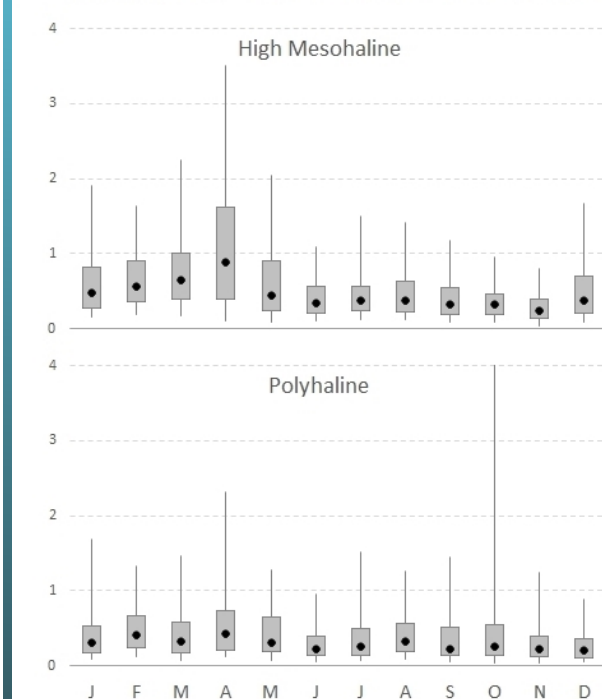
Biomass of Living Nano- Mico- Phytoplankton Cells (mg C/liter)



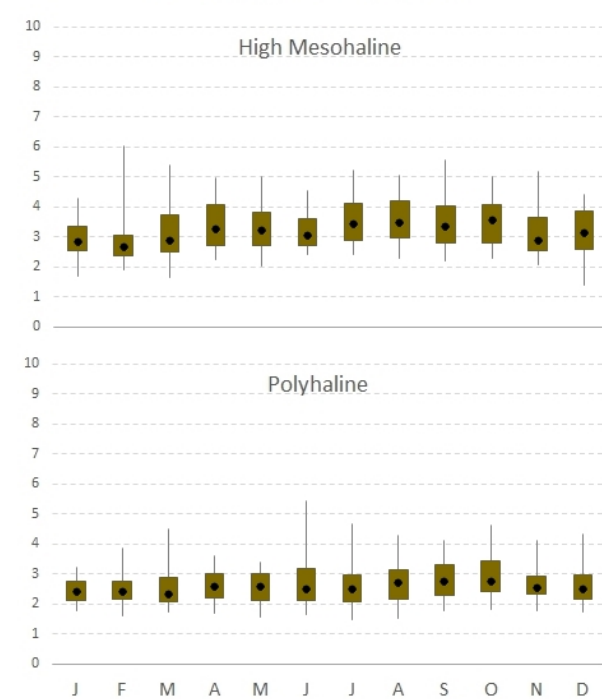
Dissolved Organic Carbon (mg/liter)



Biomass of Living Nano- Mico- Phytoplankton Cells (mg C/liter)



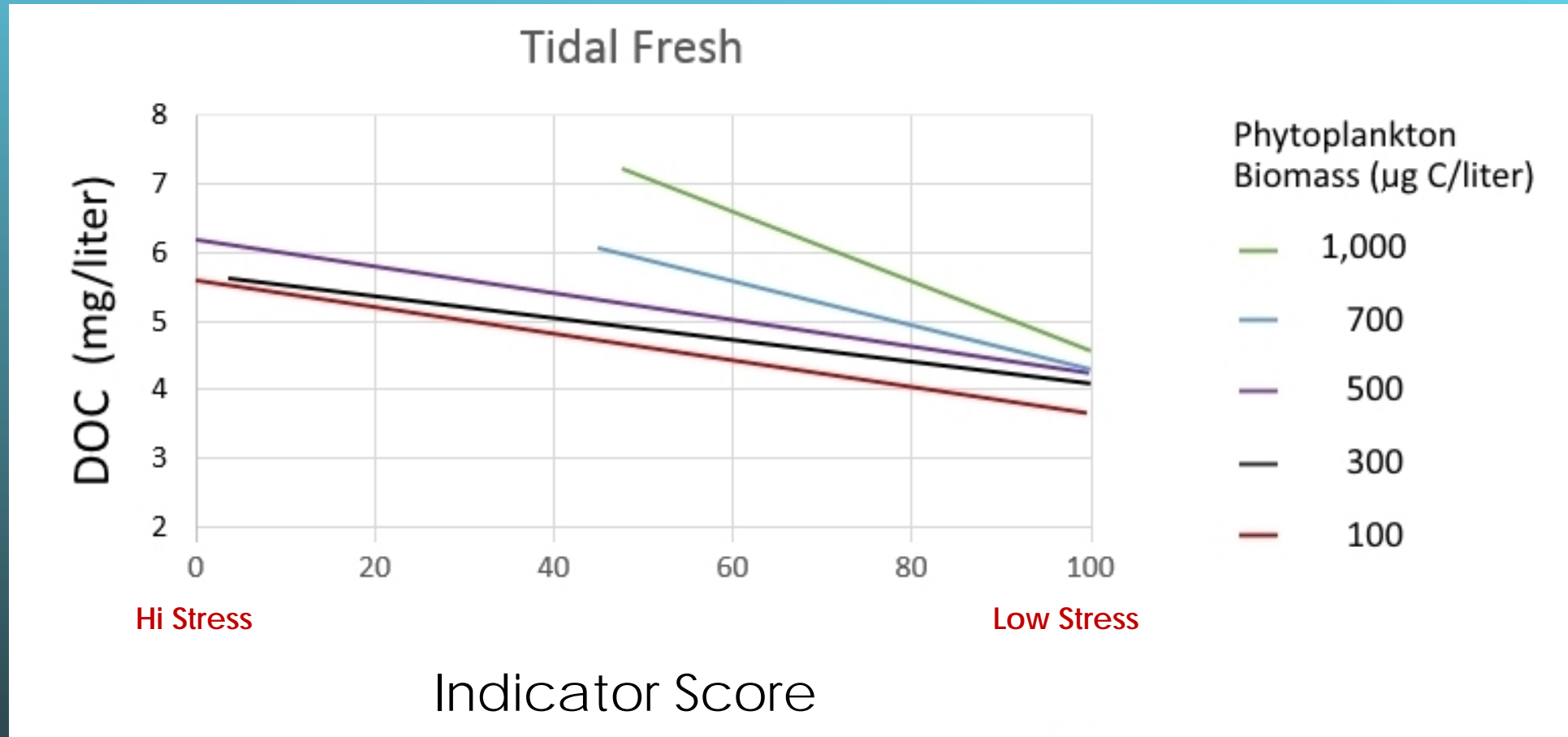
Dissolved Organic Carbon (mg/liter)



CBP data, 1985 – 2010

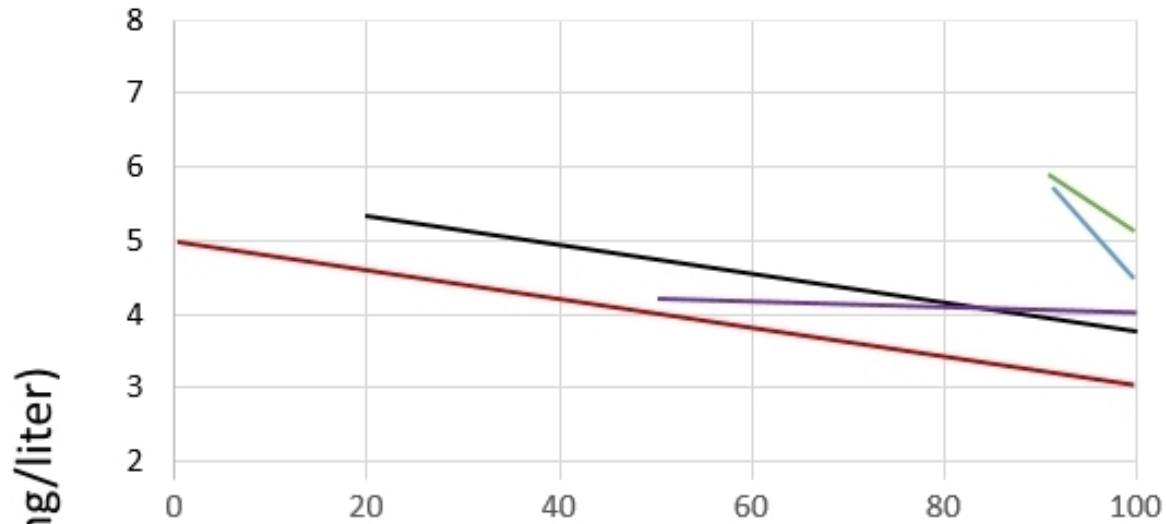
DOC paired with biomass at phytoplankton monitoring stations only

Relationship of Low-Light Stress Indicator to DOC

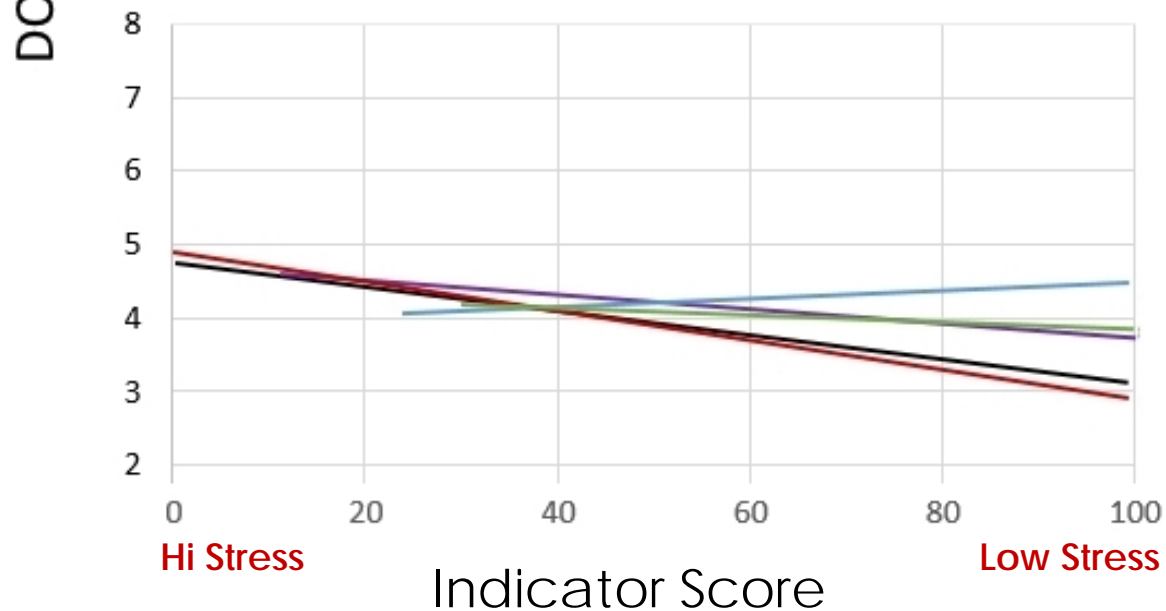


- For a given stress level, higher phytoplankton biomass \Leftrightarrow higher ambient DOC
- For a given biomass level, stressed cells \Leftrightarrow higher ambient DOC

Oligohaline



Low Mesohaline



Hi Stress

Low Stress

Indicator Score

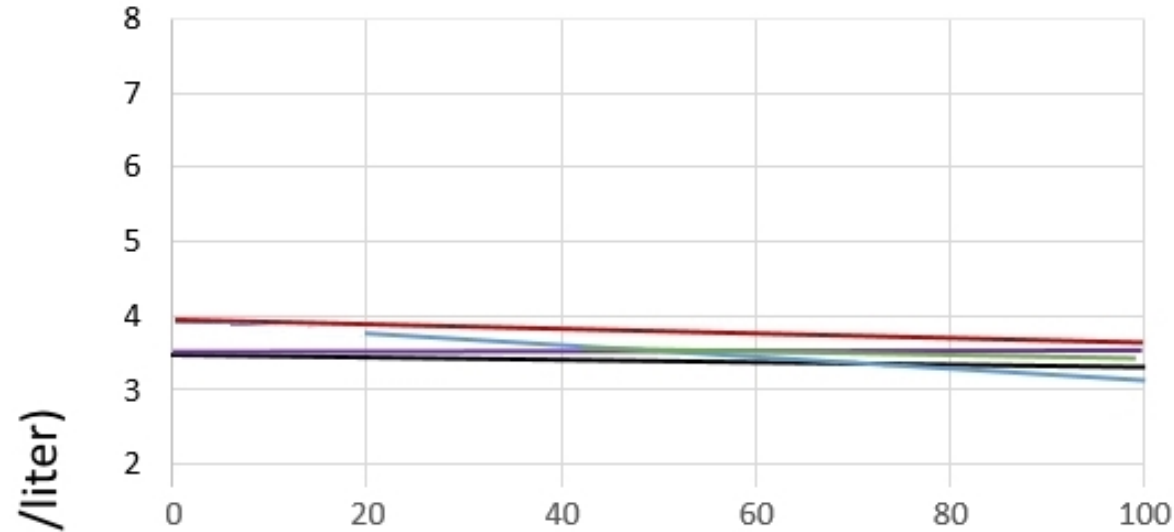
Phytoplankton
Biomass (µg C/liter)

- 1,000
- 700
- 500
- 300
- 100

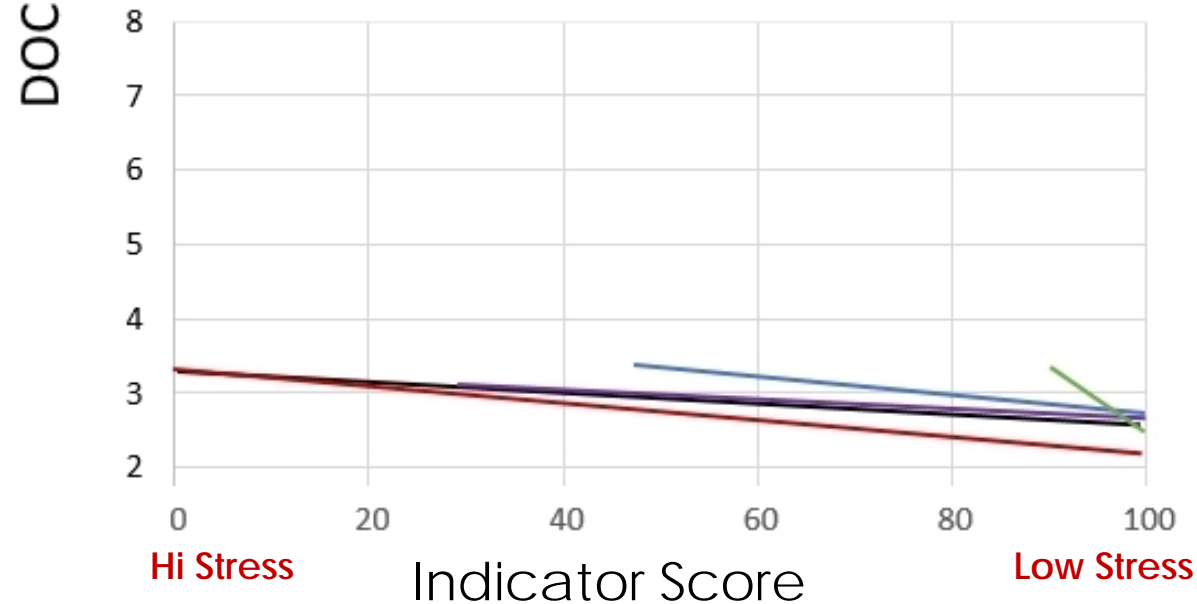
Oligohaline & Low Mesohaline

- For lower biomasses, stressed cells \leftrightarrow higher ambient DOC
- For a given stress level, higher phytoplankton biomass \leftrightarrow higher ambient DOC

High Mesohaline



Polyhaline

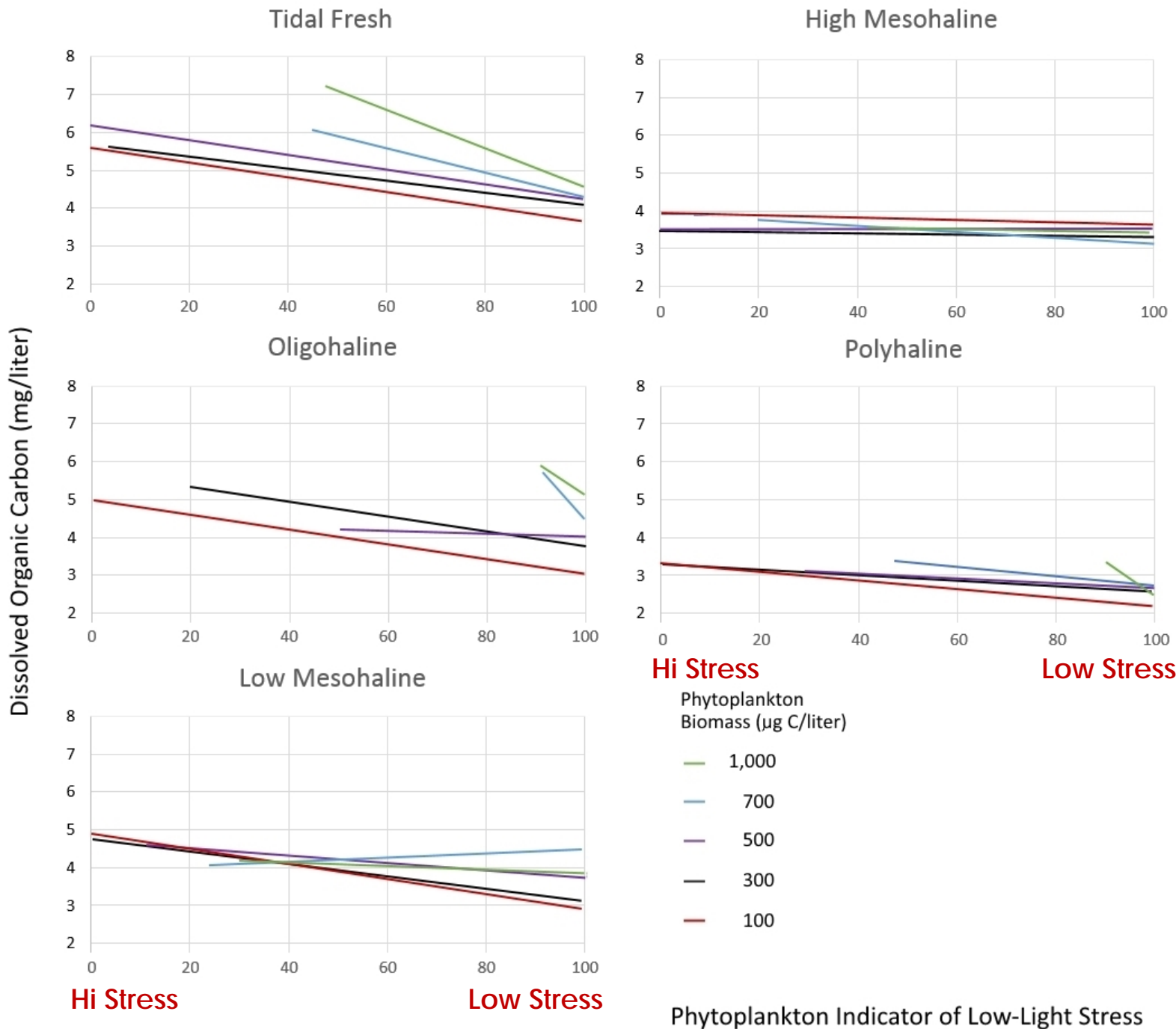


Phytoplankton
Biomass ($\mu\text{g C/liter}$)

- 1,000
- 700
- 500
- 300
- 100

Hi Mesohaline & Polyhaline

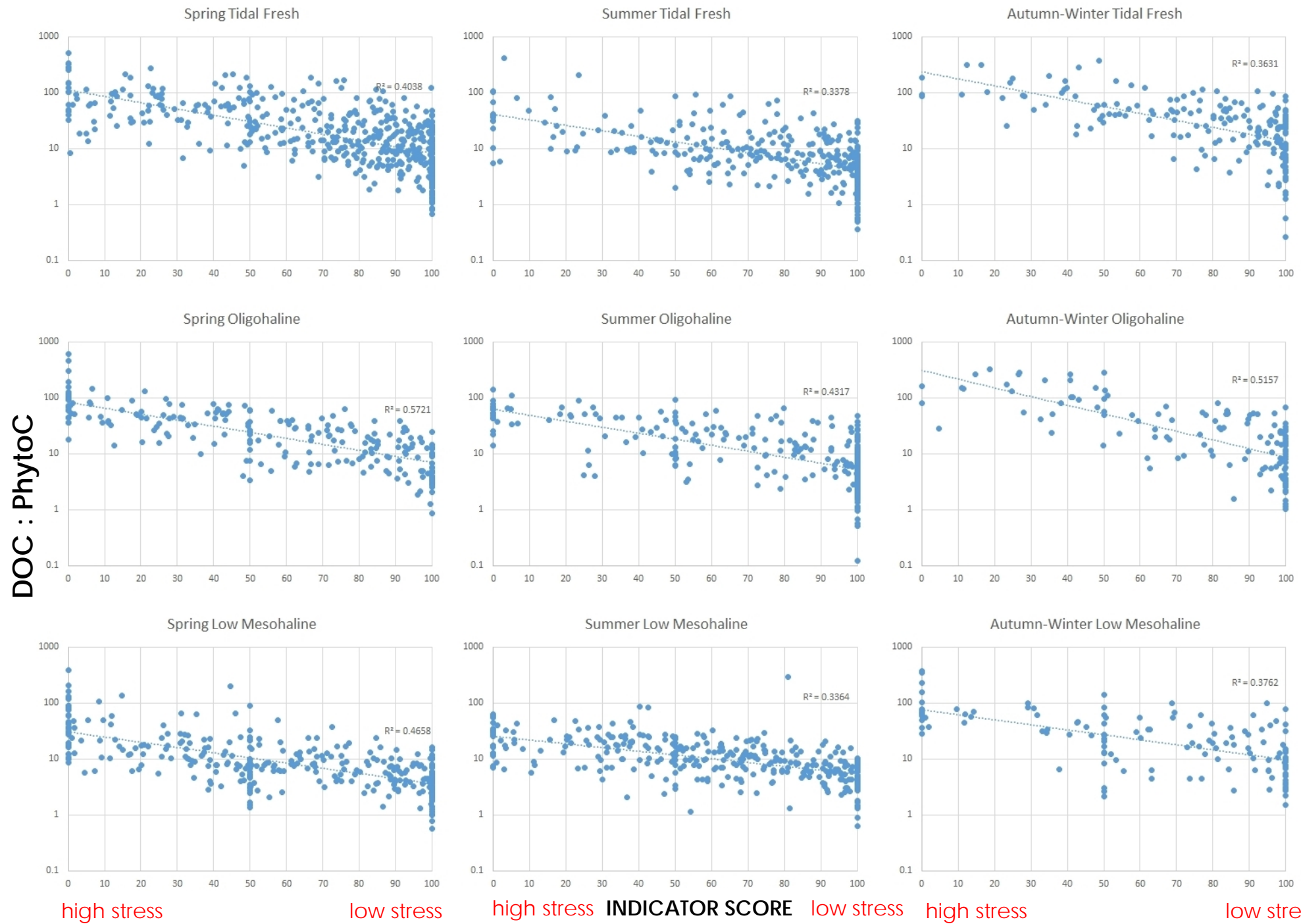
- Decreasing stress is associated with a *smaller drop* in ambient DOC
- Large blooms don't increase DOC as much (but large blooms are rarer here)



Phytoplankton Indicator of Low-Light Stress

Different slopes in TF-OH-LoMH and HiMH-PH caused by changes in:

- Taxa?
- Non-living factors that attenuate light?
- Rates of vertical mixing?
- Other?
- **All of the above?**



TF

OH

LoMH

high stress

low stress

high stress

INDICATOR SCORE

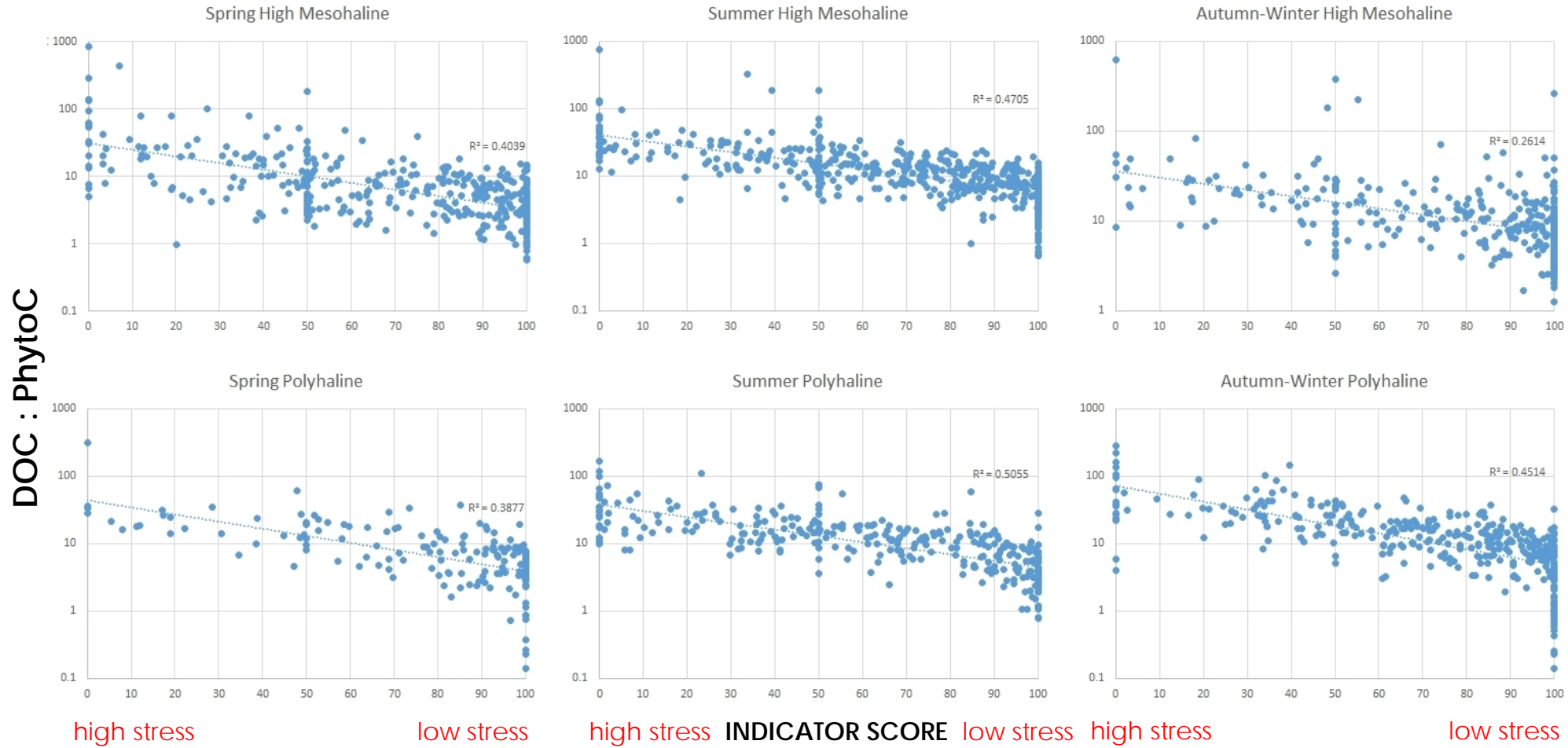
low stress

high stress

low stress

HiMH

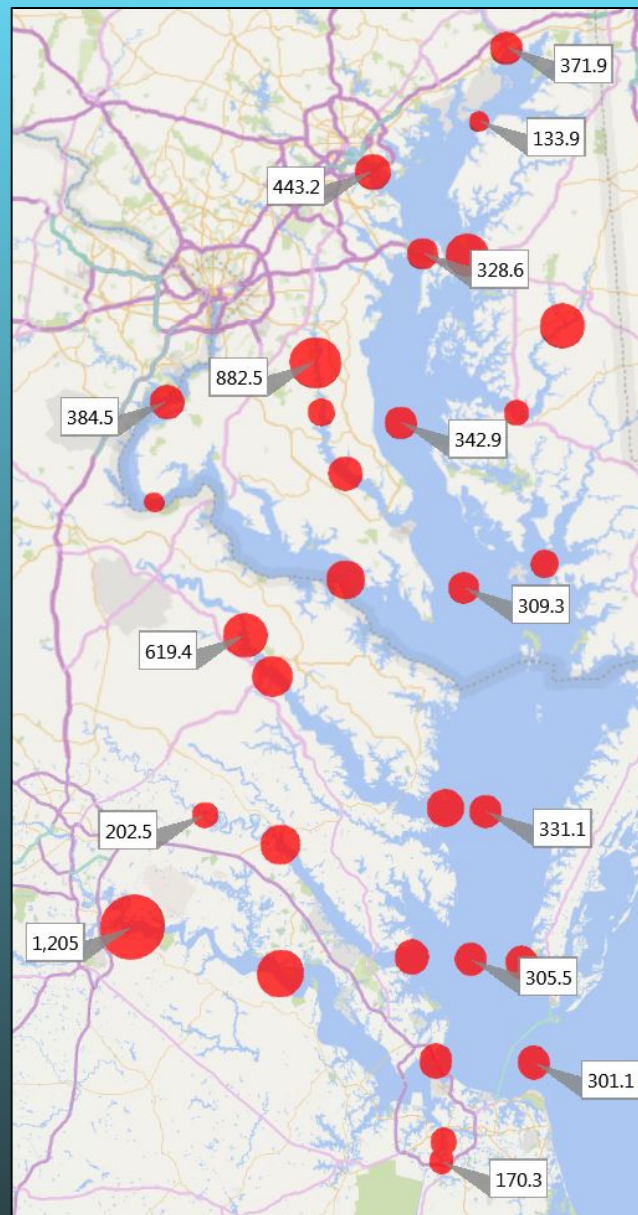
PH



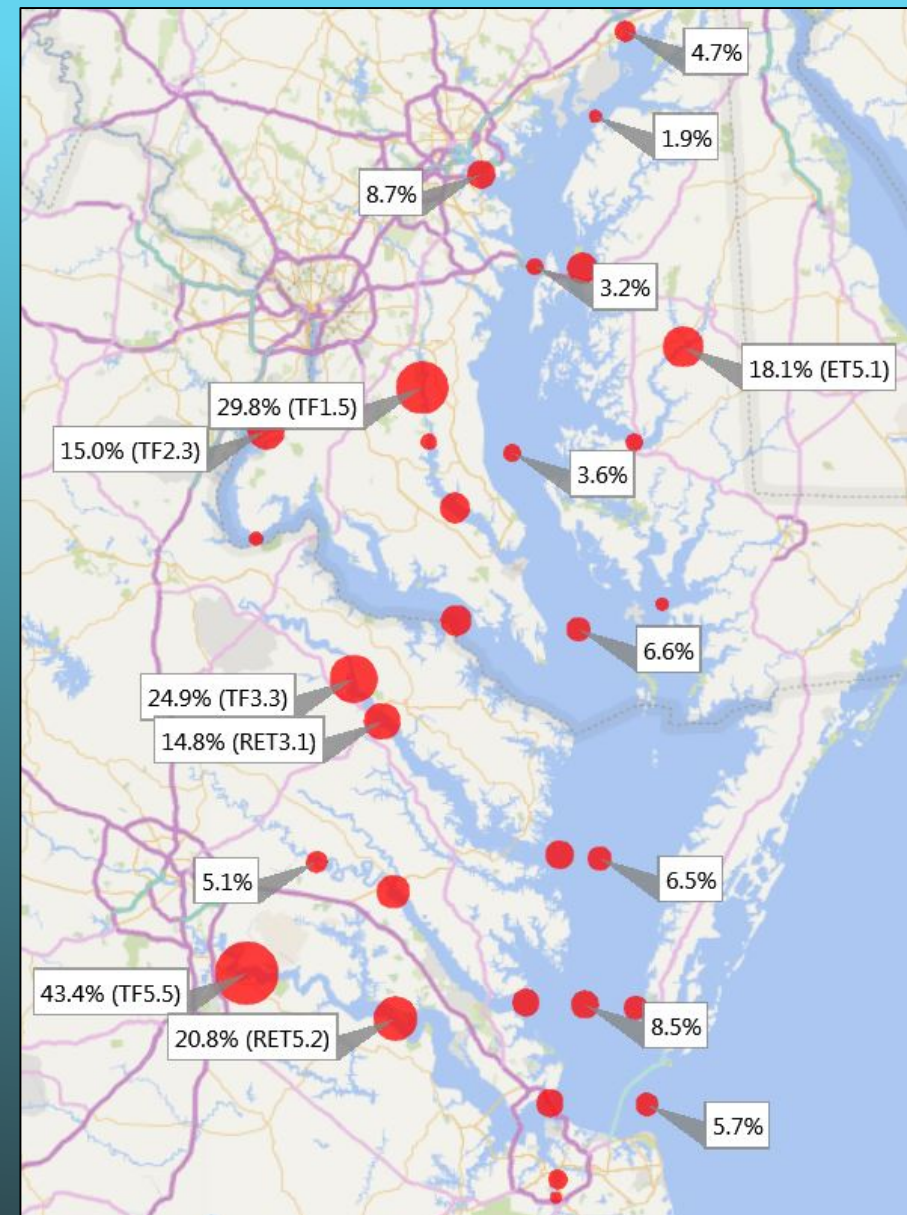
Geometric Mean DOC:PhytoC			
	Hi Stress	Low Stress	% Drop
F	77.19	4.75	-94%
O	74.34	4.87	-93%
LoM	41.01	4.27	-90%
HiM	47.30	4.72	-90%
P	41.67	2.41	-94%

Cells leak (exude)
proportionally more DOC
when stressed by low-light
conditions

Higher phytoplankton
biomass = greater amount
of DOC exuded



Station median phytoplankton
biomass (µg C/liter)



% biomass exceedances of
1,500 µg C/liter (blooms)

Phytoplankton exude:

Glycolic acid

Carbohydrates

Polysaccharides

Amino acids

Peptides

Organic phosphates

Volatile compounds

Enzymes

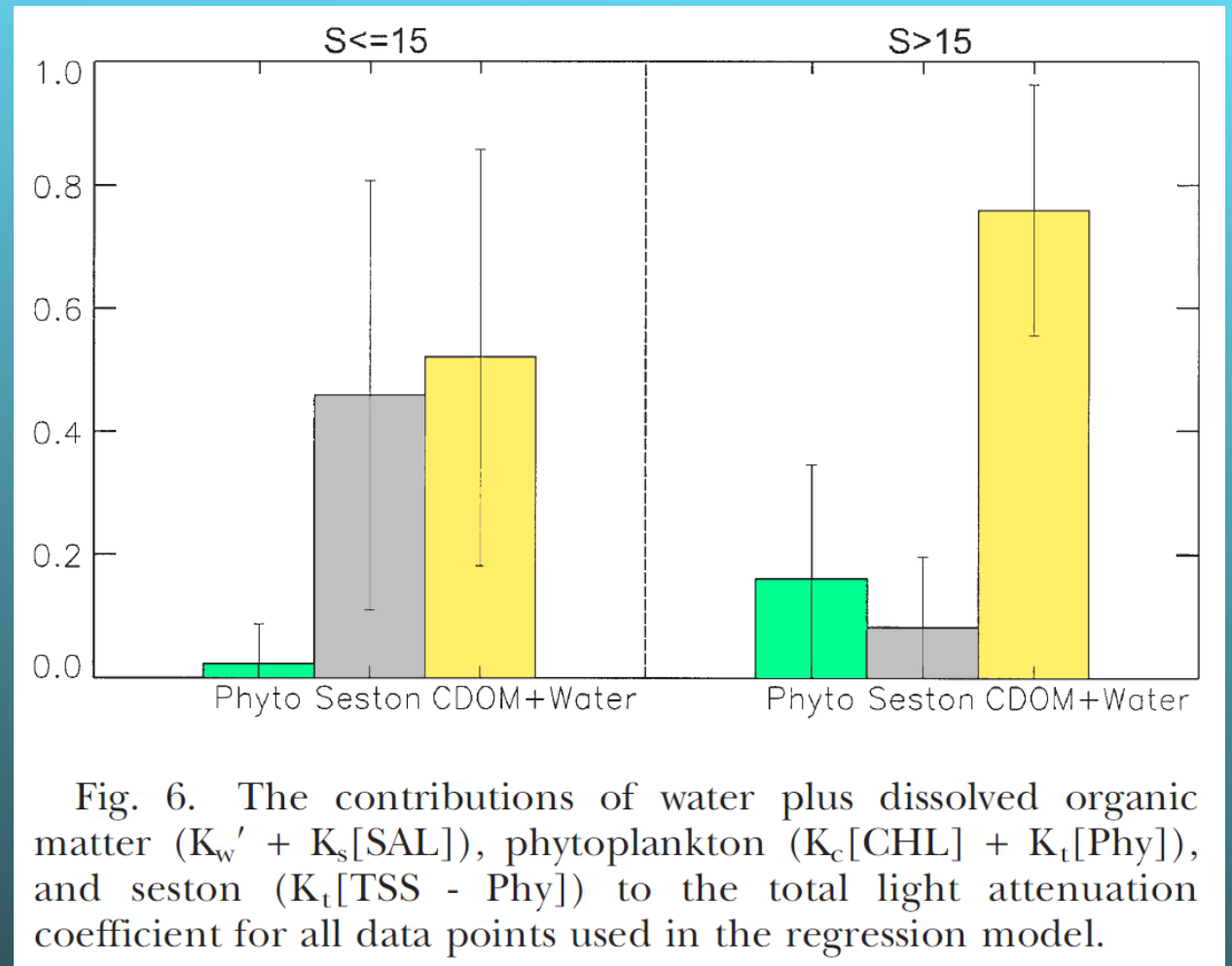
Vitamins

Hormonal substances

Toxins

Allelochemicals that inhibit photosynthesis
in competing taxa

Many are small, labile, low-molecular-weight compounds

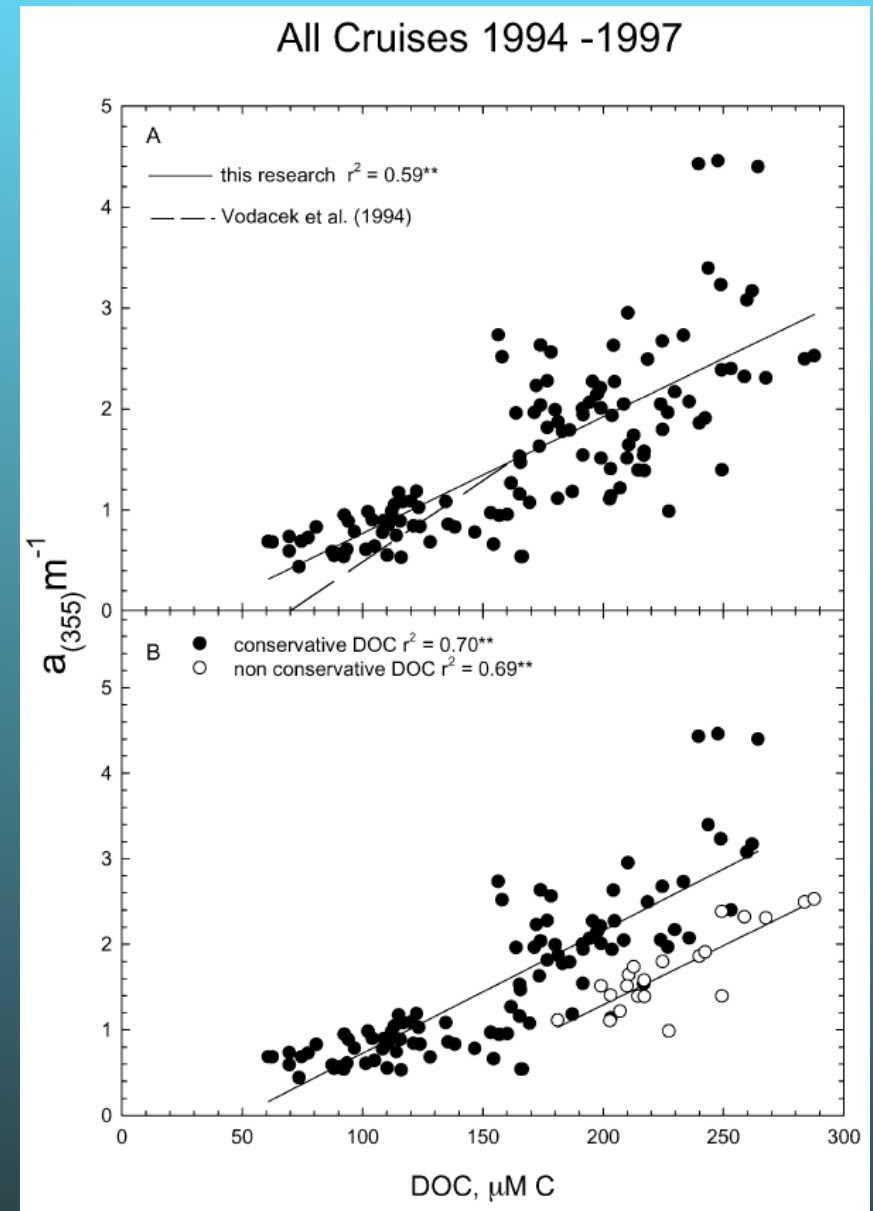


Xu et al. (2005) – non-spectral empirical model of light attenuation in Chesapeake Bay

Rochelle-Newall and Fisher (2002) – measured CDOM, DOC, and Chla in Bay mainstem cruises

- Overall, DOC and CDOM correlate (others have found this relationship too)
- CDOM behaves ‘conservatively’ (like salinity)
→ large, high-molecular-weight, refractory
- Increased production of non-chromophoric DOC occurs in Chla max of Bay
- No clear relationship between Chla and CDOM
- Concluded there were two or more sources of CDOM to Bay

CDOM (absorption at 355 nm)



DOC

Possible Phytoplankton Role in Light Attenuation

- CDOM and suspended sediments are most responsible for light attenuation in Bay mainstem (e.g., Xu et al. 2005)
 - Microbial processing of detritus (multiple sources) generates much of CDOM in Bay mainstem (e.g., Rochelle-Newall & Fisher 2002)
6. Die-offs of these phytoplankton & bacteria are a large source of autochthonous detritus
 5. Largest biomasses of phytoplankton & bacteria presently occur in tributaries (not upper Bay)
 4. Exuded DOC contains relatively little CDOM but inputs copious amounts bacterial substrate; bacterial biomass increases and produces CDOM
 3. Cells leak (exude) proportionally more DOC when stressed by poor light
 2. Phytoplankton increase cellular concentrations of photopigments to compensate; boom-bust growth cycles occur if waters are also nutrient-rich
 1. Non-living suspended solids create poor light conditions; impede photosynthesis in open water

- Phytoplankton are a *controllable* internal source of DOC and can influence CDOM concentrations in open waters



Caption: Always bring lights — that's one lesson Nick Caloyianis learned from decades of filming and photographing underwater life in the Chesapeake Bay. His photographs document the loss of natural light in the estuary. In 2008 (above top) he worked with multiple lights as he tried to film oyster restoration on Dominion Reef in the mainstem Bay. Back in 1979 (above bottom), he had plenty of light to photograph his partner, Clarita Berger, as she took pictures of the grassbeds of the Choptank River. Credits: Michael Eversmier, Aqua Ventures, inc. (left) and Nick Caloyianis (right). Fincham, M. 2008 "Naturalist at Bay: A Winter's Tale" Chesapeake Quarterly vol. 7 no. 4

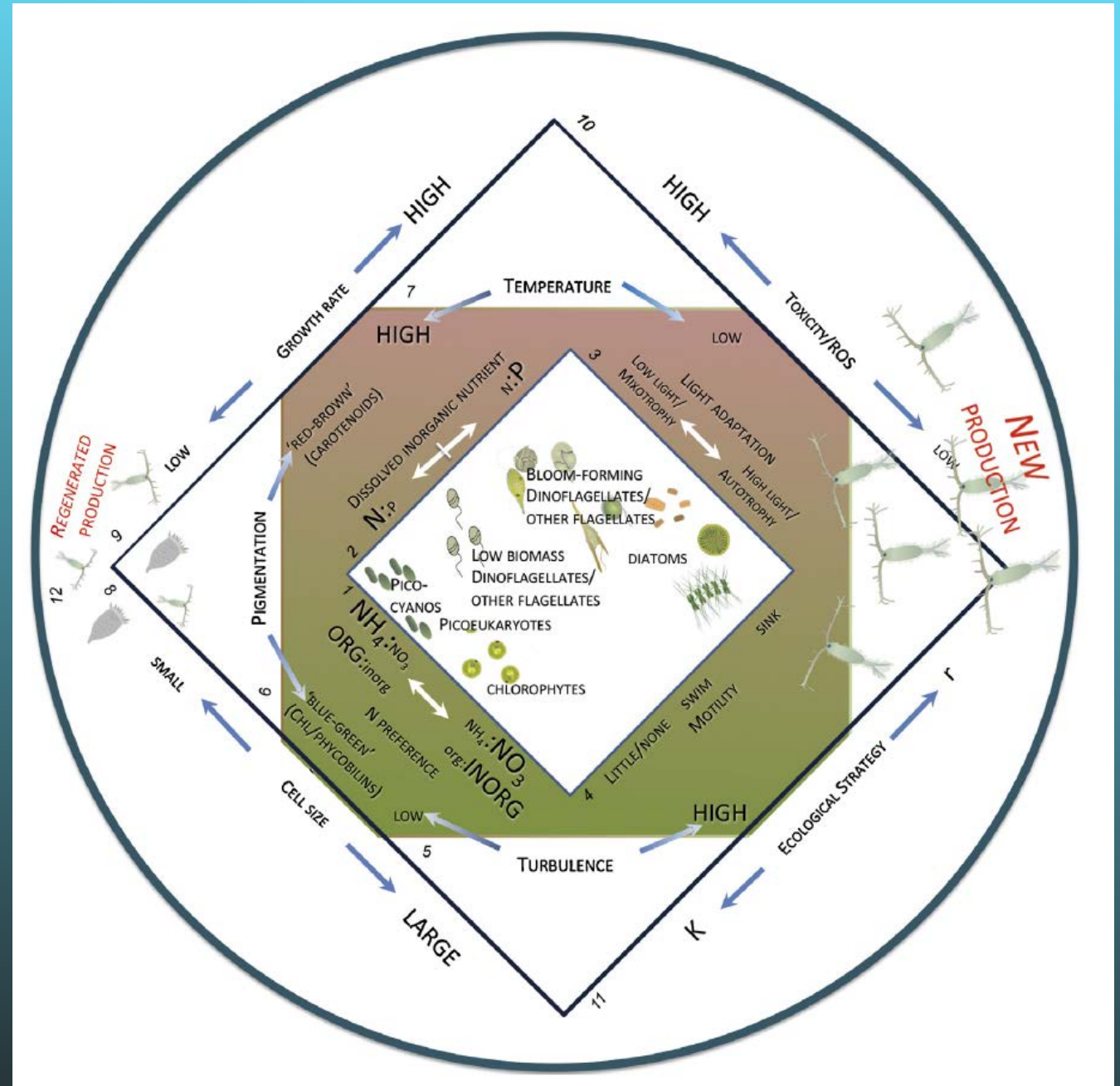
		Buchanan et al. 2005				Revised thresholds (2016)			
Secchi Depth (meters)		worst	poor	better	best	worst	poor	better	best
Season	Salinity	<	≤	>	>	<	≤	>	>
Spring	F	0.7	0.9	1.1	1.1	0.6	0.8	1.1	1.1
	O	0.5	0.7	1.1	1.1	0.7	0.8	1.0	1.0
	LoMH	1.35	1.8	2.25	2.25	1.2	1.4	1.7	1.7
	HiMH	1.35	1.8	2.25	2.25	1.5	1.8	2.0	2.0
	P	1.6	2.15	2.55	2.55	1.7	2.1	2.6	2.6
June	F					0.6	0.7	0.8	0.8
	O					0.6	0.7	0.8	0.8
	LoMH					1.0	1.2	1.5	1.5
	HiMH					1.1	1.4	1.7	1.7
	P					1.3	1.6	1.8	1.8
Summer	F	0.6	0.8	1.0	1.0	0.6	0.8	1.1	1.1
	O	0.55	0.6	0.7	0.7	0.6	0.8	1.0	1.0
	LoMH	1.2	1.45	1.7	1.7	0.9	1.2	1.4	1.4
	HiMH	1.2	1.45	1.7	1.7	1.4	1.6	1.8	1.8
	P	1.55	1.85	2.35	2.35	1.5	1.8	2.1	2.1
Autumn	F	0.7	0.9	1.2	1.2	0.7	0.9	1.2	1.2
	O	0.4	0.5	0.7	0.7	0.6	0.8	1	1
	LoMH	1.7	2.0	2.2	2.2	1.1	1.2	1.5	1.5
	HiMH	1.7	2.0	2.2	2.2	1.6	2.0	2.3	2.3
	P	1.7	2.1	3.5	3.5	1.7	2.0	2.5	2.5
Winter	F	0.5	0.6	0.9	0.9	0.6	0.8	1.1	1.1
	O	0.5	0.6	0.9	0.9	0.6	0.9	1.0	1.0
	LoMH	1.4	1.8	2.2	2.2	1.2	1.4	1.7	1.7
	HiMH	1.4	1.8	2.2	2.2	1.5	2.0	2.2	2.2
	P	1.8	2.3	2.7	2.7	1.7	2.1	2.5	2.5
DIN (mg/liter)		worst	poor	better	best	worst	poor	better	best
All Seasons	All Salinities	>	>	≤	<	>	>	≤	≤
		various	0.07	0.03	0.03	0.35	0.07	0.03	0.03
PO ₄ (mg/liter)		worst	poor	better	best	worst	poor	better	best
All Seasons	All Salinities	>	>	≤	<	>	>	≤	≤
		various	0.007	0.003*	0.003	0.035	0.007	0.003	0.003

Thresholds used to separate Secchi depth, dissolved inorganic nitrogen (DIN), and ortho-phosphate (PO₄) into four classes: worst, poor, better, and best. Buchanan *et al.* (2005) did not include June thresholds and did not have different thresholds for low and high mesohaline salinities. Seasons: Spring, March – May; Summer, July – September; Autumn, October – November; Winter, December – February. Salinity zones: tidal fresh (F), <0.5‰; oligohaline (O), >0.5 – 5.0‰; low mesohaline (LoMH), >5.0 – 10‰; high mesohaline, (HiMH), >10 – 18‰; polyhaline (P), >18.0‰. *The mesohaline PO₄ threshold in Buchanan *et al.* (2005) was 0.002 mg/liter. See text for further details.

Patricia Glibert. (2016) in Harmful Algae. Phytoplankton mandala

12 dimensions

Each axis depicts a phytoplankton functional type



Keller & Hood (2011) model of DOC sources at CB3.3C (Low Mesohaline)

“...phytoplankton exudation was the dominant source of DOC, averaging 63% of total DOC production throughout the year [at CB3.3C]...”

Flowing in
to CB3.3C

19%

30%

51%



CB3.3C

phyto exudation, mortality

Labile 40%

Semi-labile 59%

Refractory 1%

sloppy feeding

Labile 53%

Semi-labile 45%

Refractory 2%

detritus decay

Labile 50%

Semi-labile 49%

Refractory 1%

