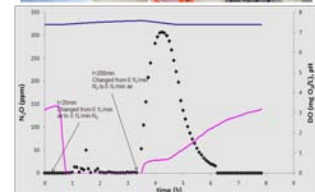
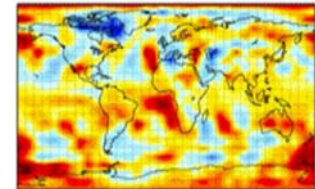


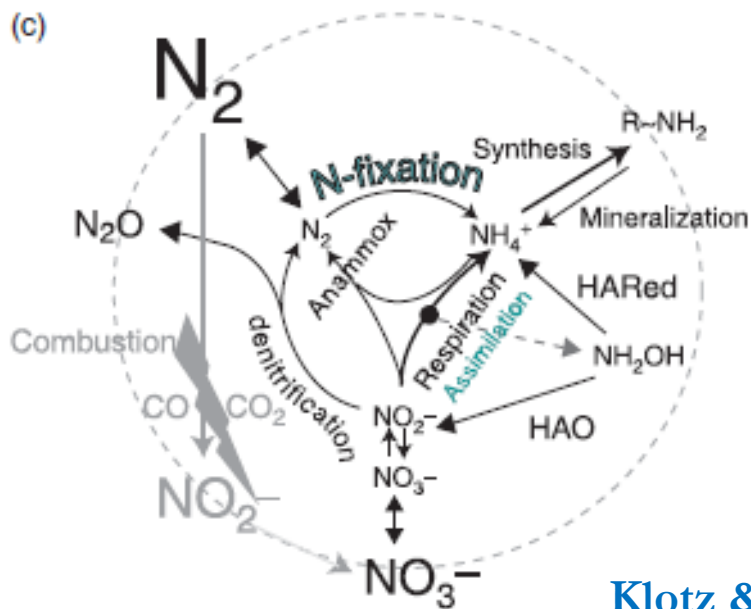
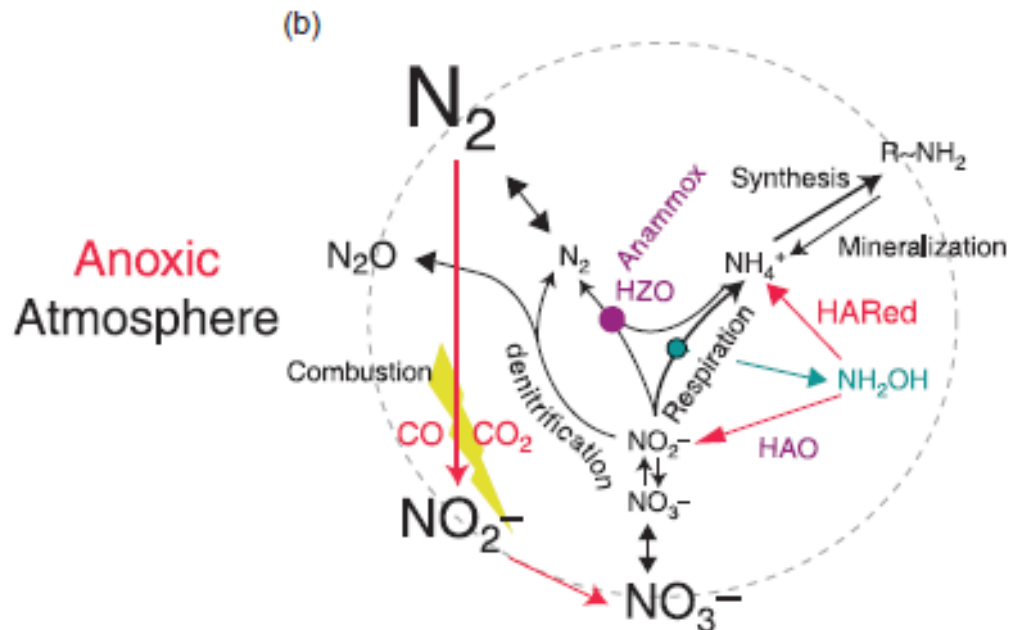
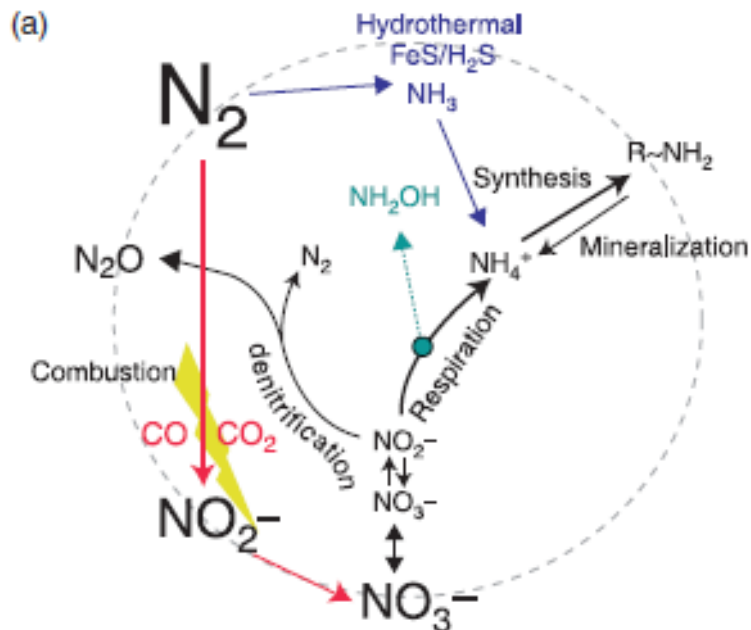
# Microbial Nitrogen Transformations

Kartik Chandran  
Columbia University

Real World Wastewater Technologies Workshop

May 16<sup>th</sup>, 2012

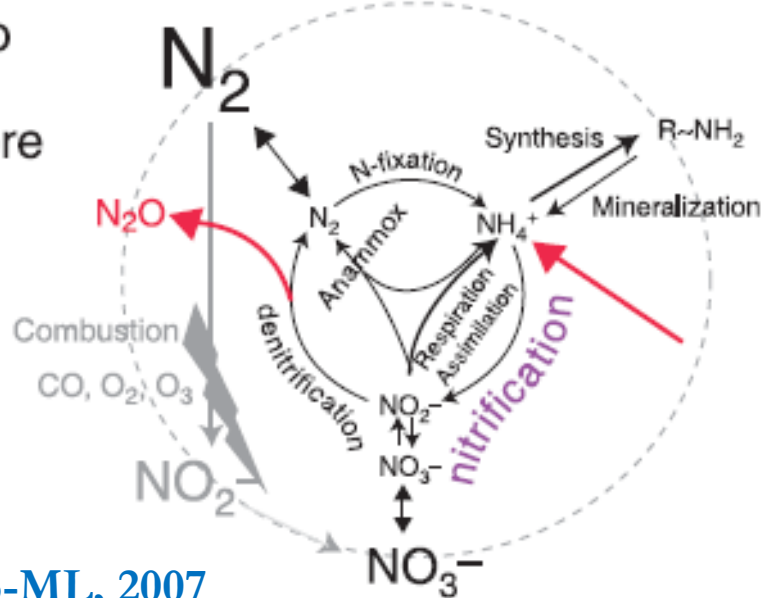




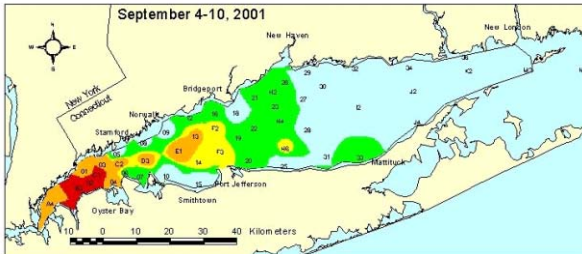
Anoxic to Oxic Atmosphere



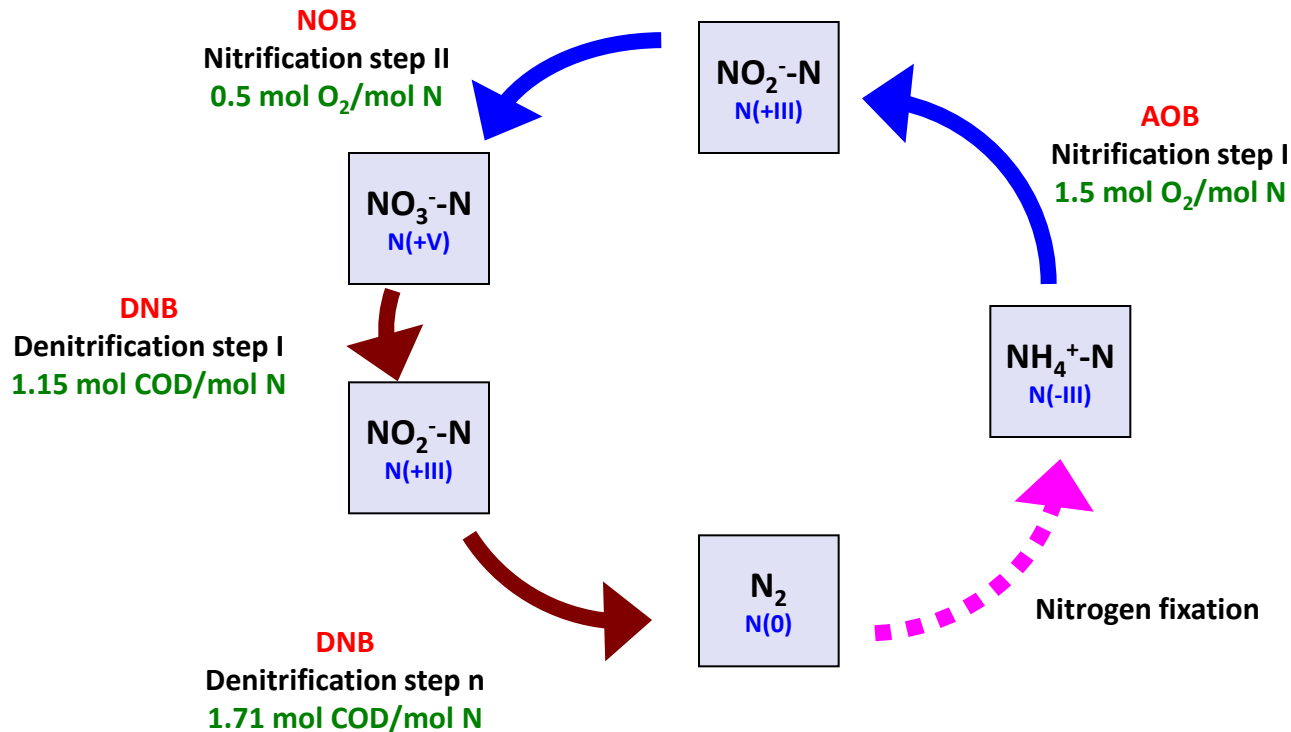
O<sub>2</sub>

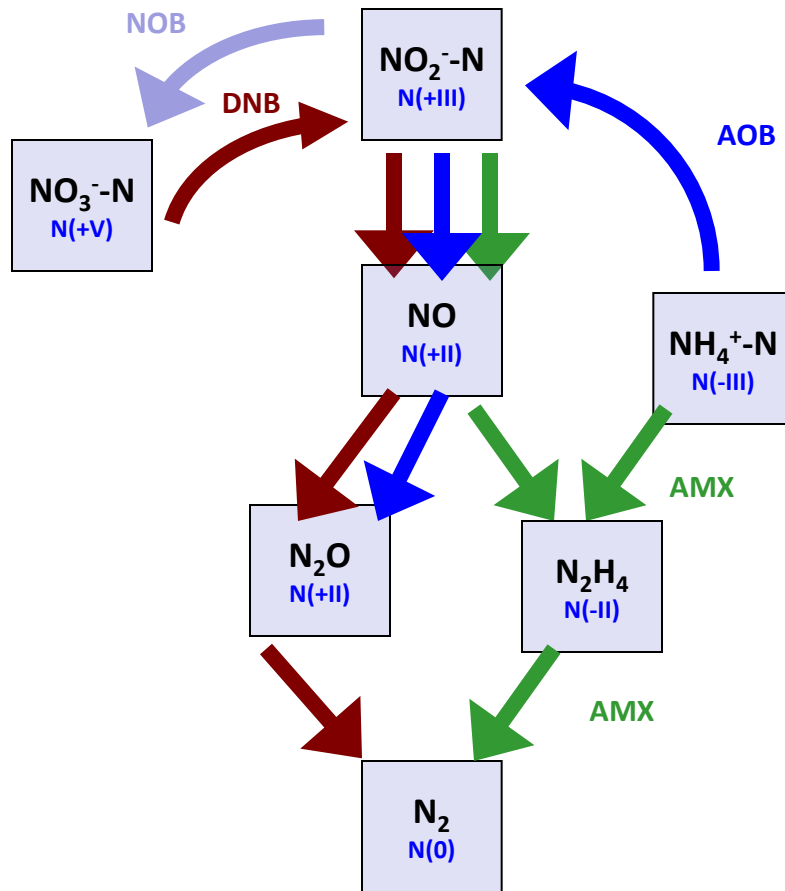


# Engineering the N-cycle



\$\$\$



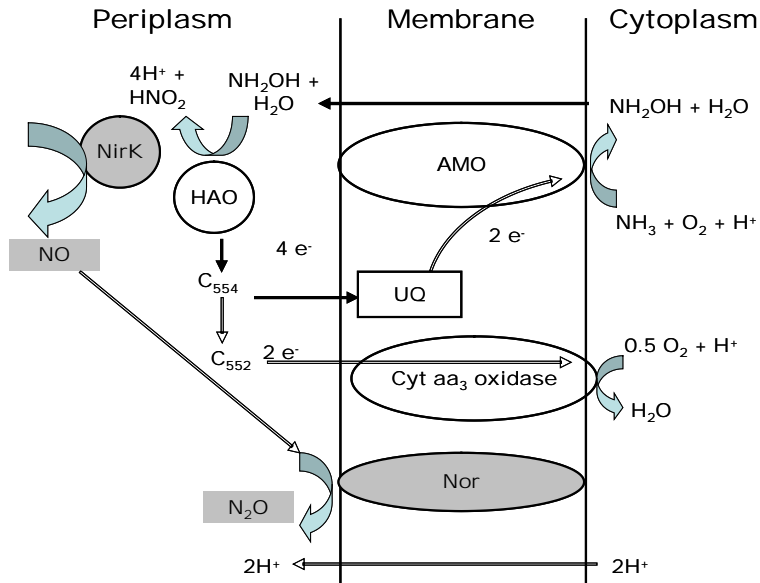


- Several intermediates reactive
  - NO<sub>2</sub><sup>-</sup>-N, NO, NH<sub>2</sub>OH
  - Control expression of pathways
- AOA not even included
- e- donor based interactions not included
- How to resolve activities?
- How to resolve contribution to mass balances?

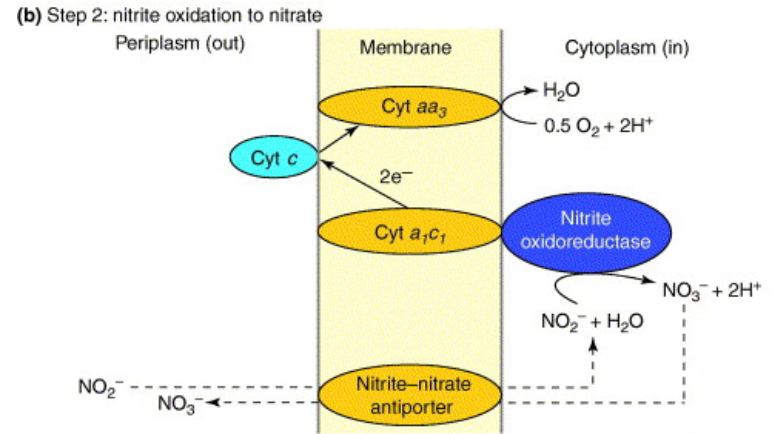
Engineered BNR systems are typified by multiple activities in concert or competition



# Some commonly studied pathways

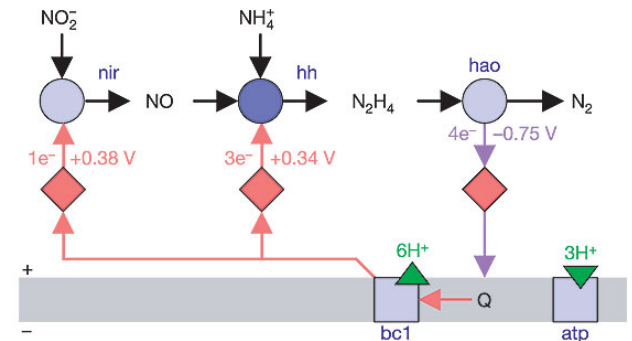
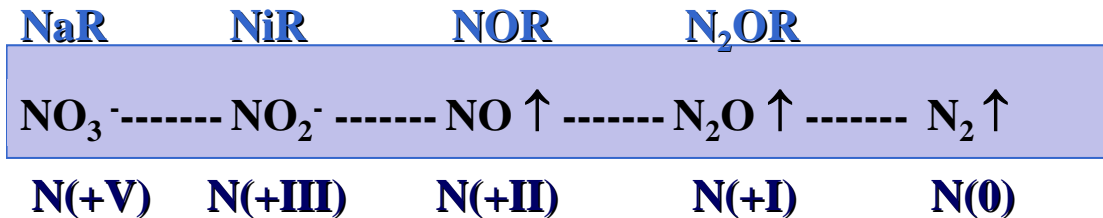


Yu *et al.*, ES&T 2010, 44(4), 1313-1319



TRENDS in Microbiology

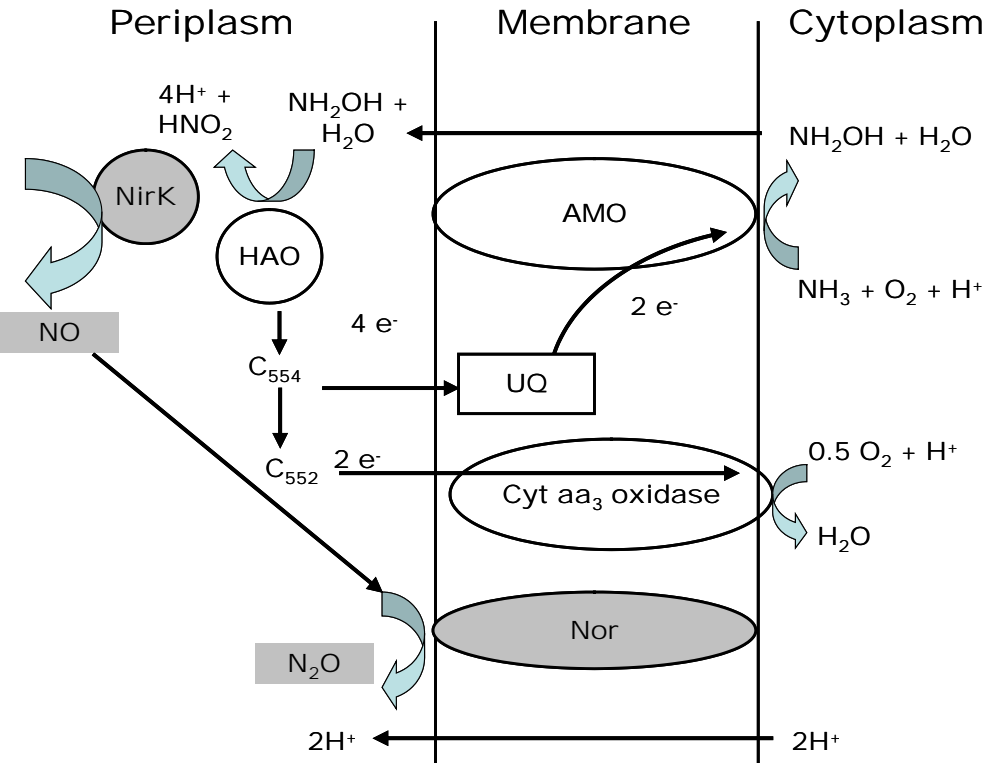
Costa *et al.*, Trends in Microbiology 2006, 14, 213-219



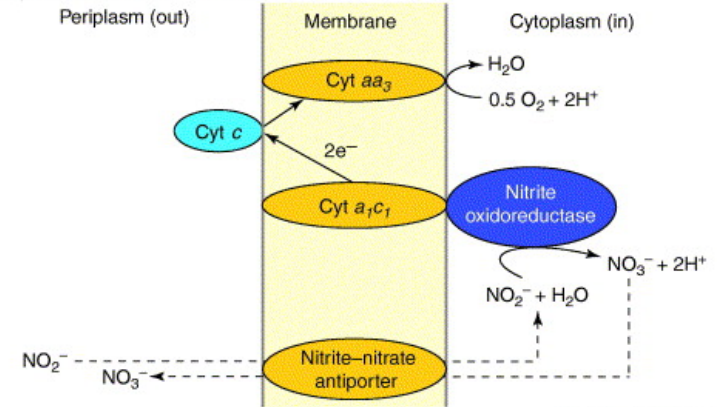
Strous *et al.*, Nature 2006, 440, 790-794



# Nitrification



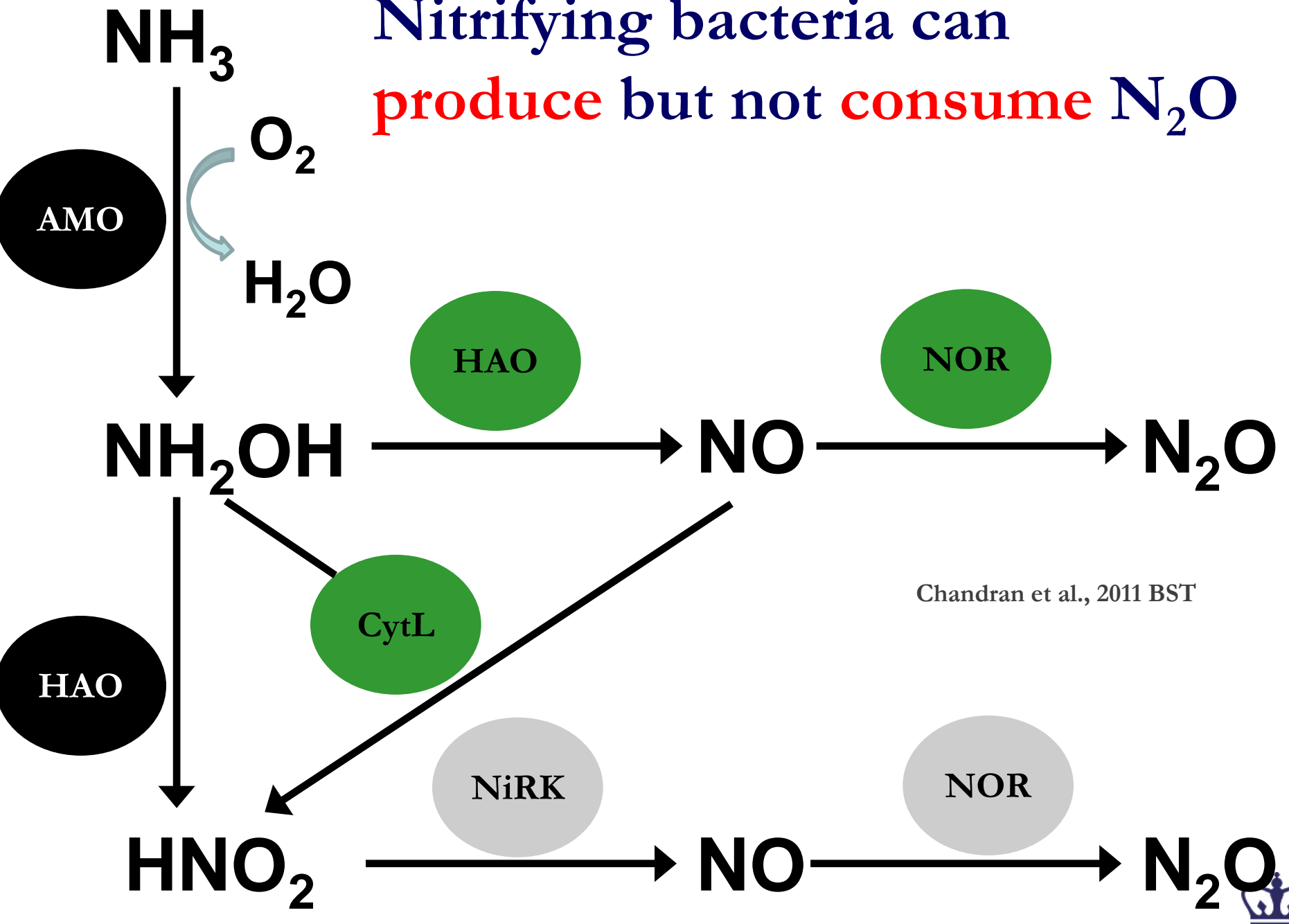
(b) Step 2: nitrite oxidation to nitrate



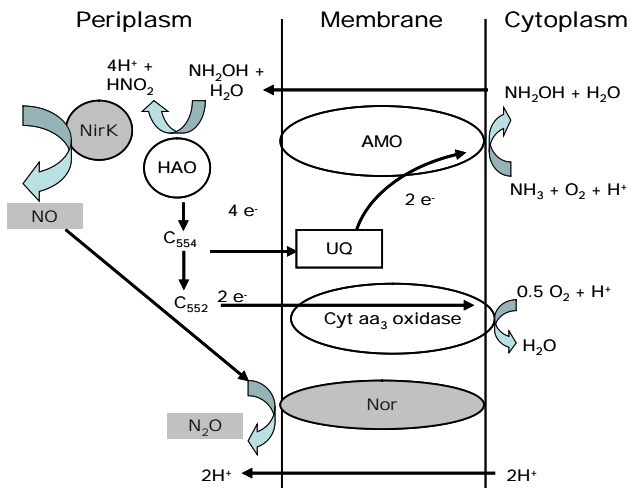
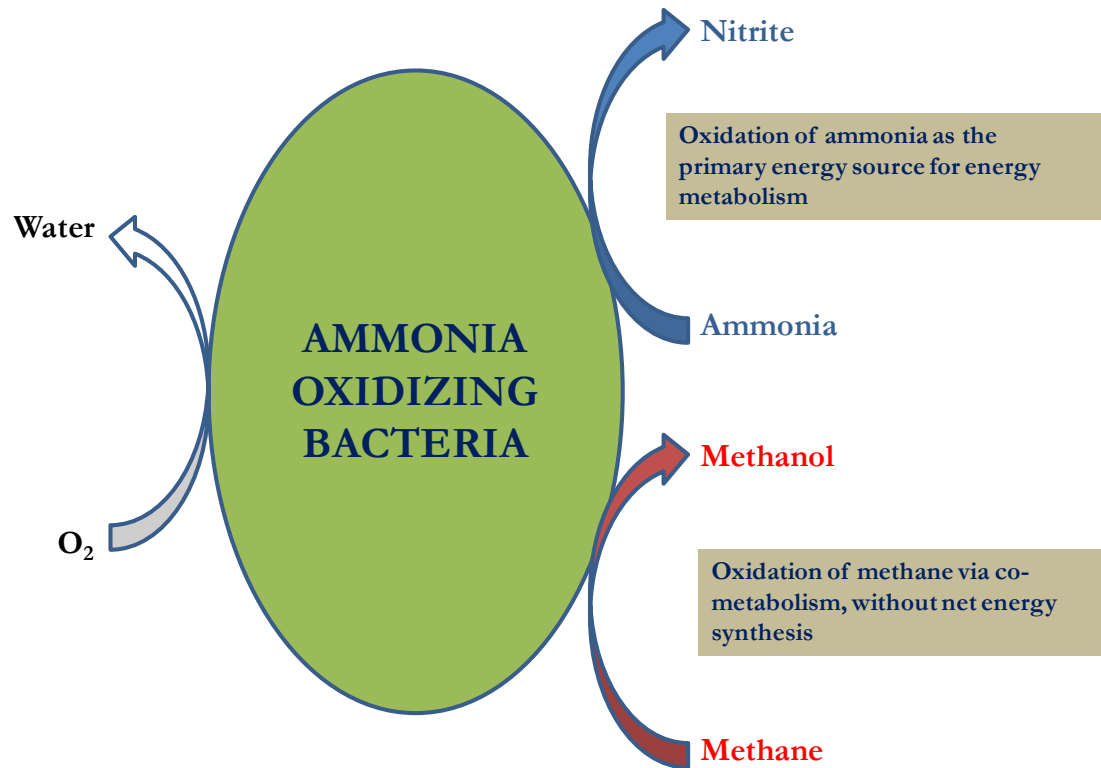
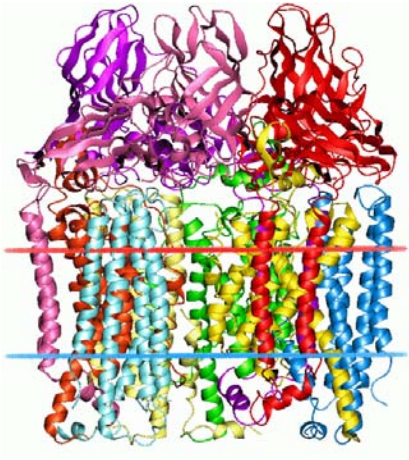
TRENDS in Microbiology



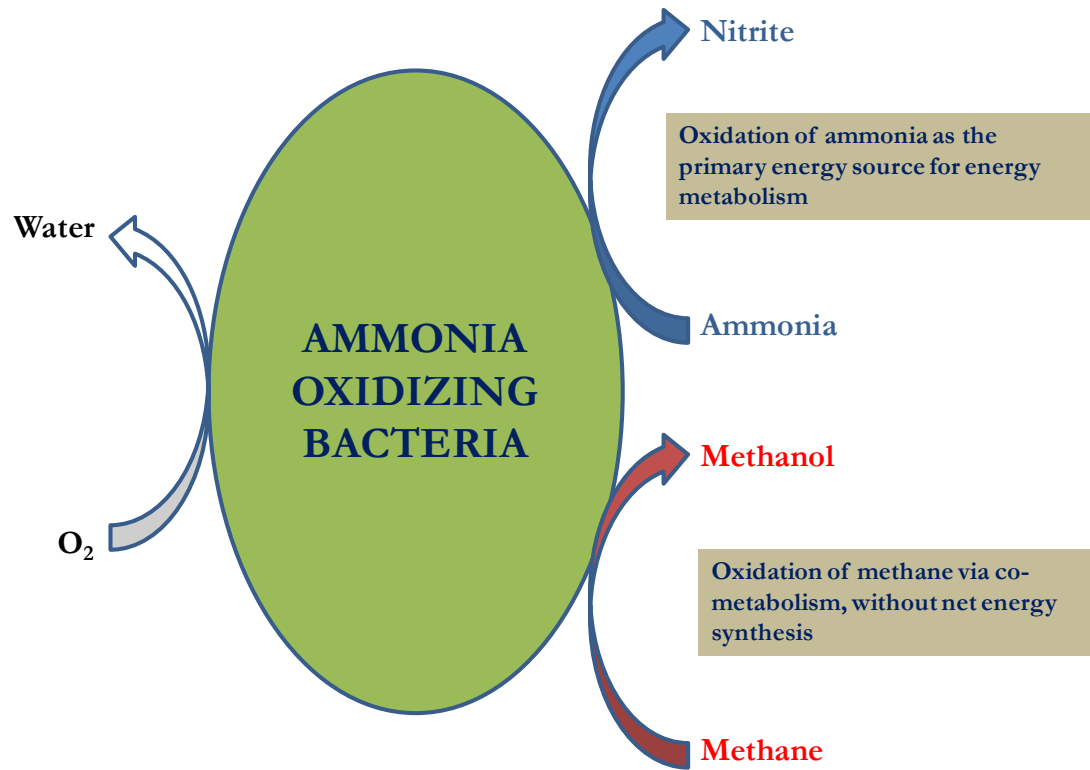
Nitrifying bacteria can  
**produce** but not **consume**  $N_2O$



# Conceptual system in AOB







- Concomitant oxidation of CH<sub>4</sub> and CO<sub>2</sub> fixation
  - Digester gas contains CO<sub>2</sub>
  - Foulant for chemical catalyst; but a food source for AOB
  - Moisture- not really an issue
- Prospect of combining C & N cycles



# Biological production of methanol

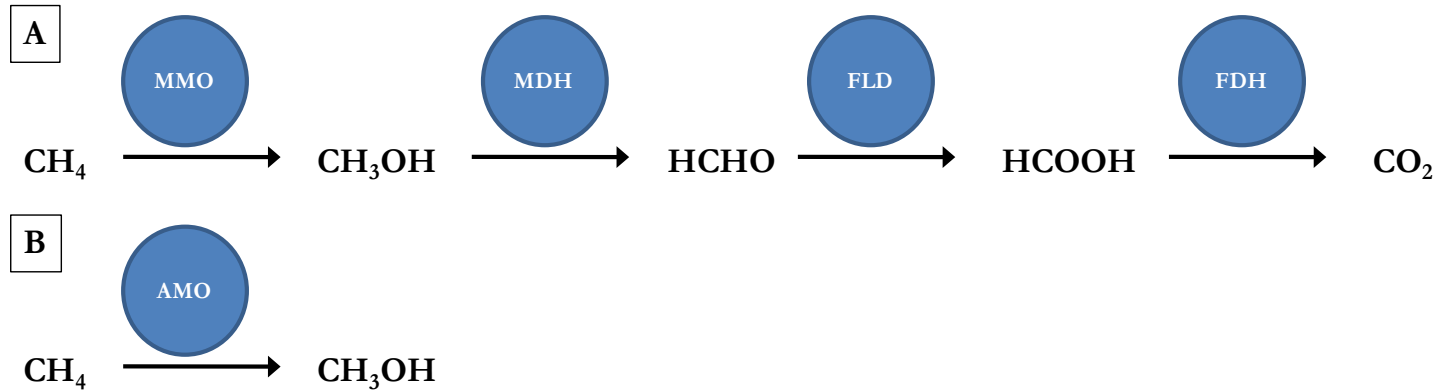
- Difficult to chemically break the C-H bonds
- (Some) bacteria do this all the time



- Ideally, partial oxidation of CH<sub>4</sub> is desired



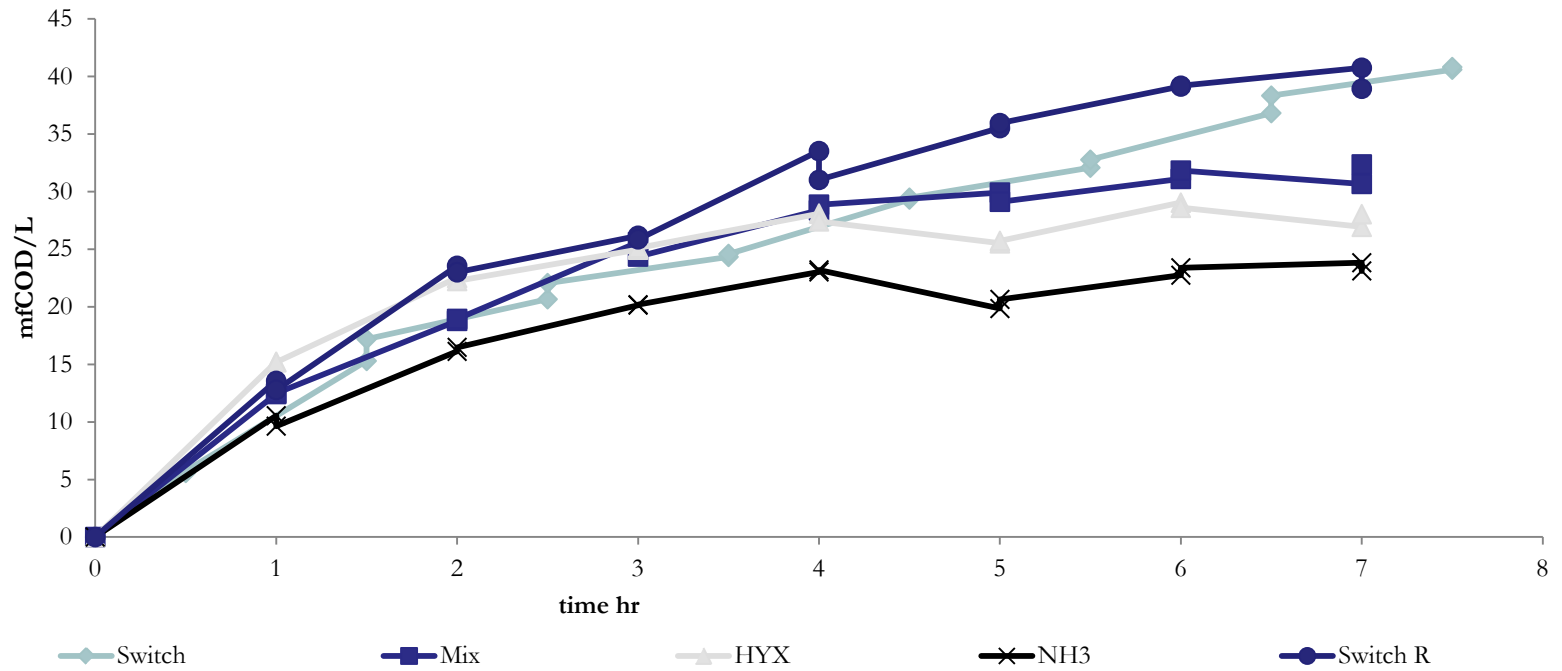
# Exploring the prospect of selective oxidation of $\text{CH}_4$ to $\text{CH}_3\text{OH}$



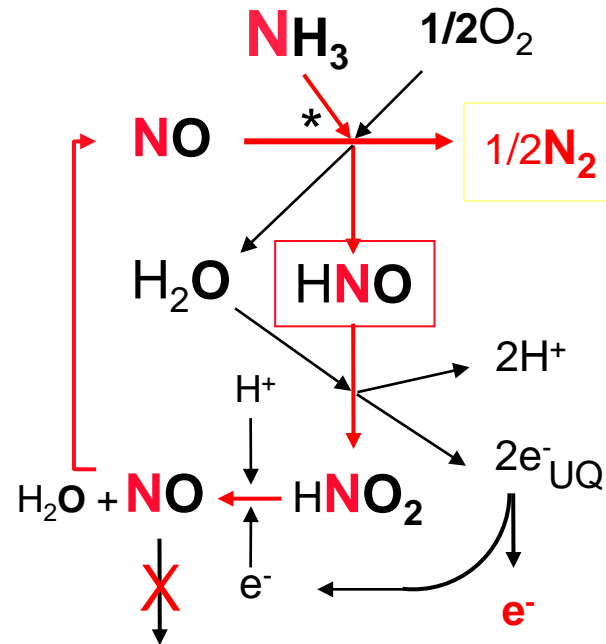
- Ammonia oxidizing bacteria (AOB) mostly oxidize methane to methanol
- They lack the metabolic pathways to produce  $\text{CO}_2$



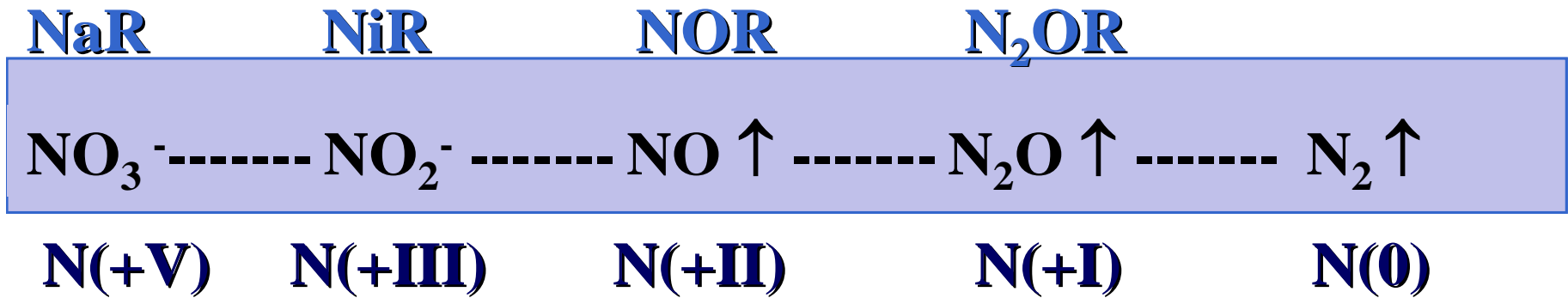
# Results



# Ammonia oxidation by archaea



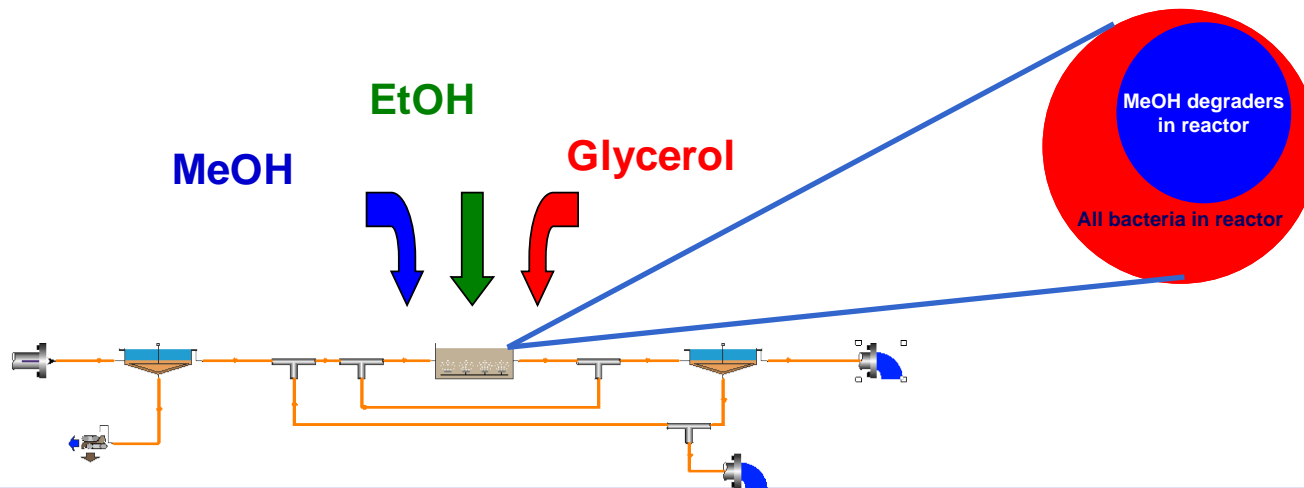
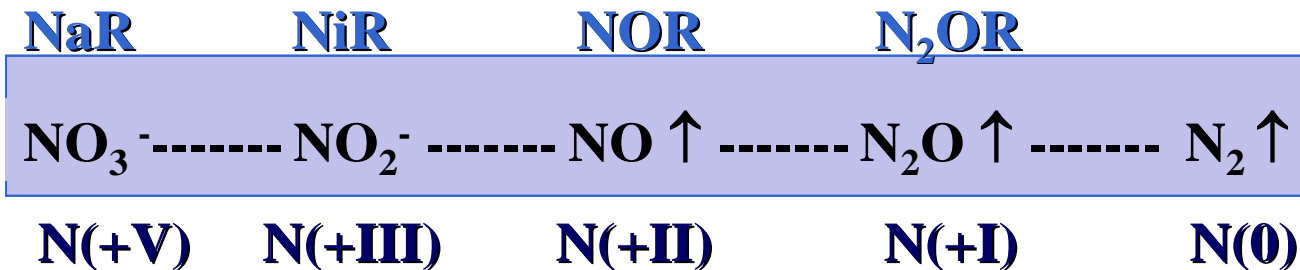
# Denitrification



- Can be driven by organics, (CH<sub>4</sub>), S



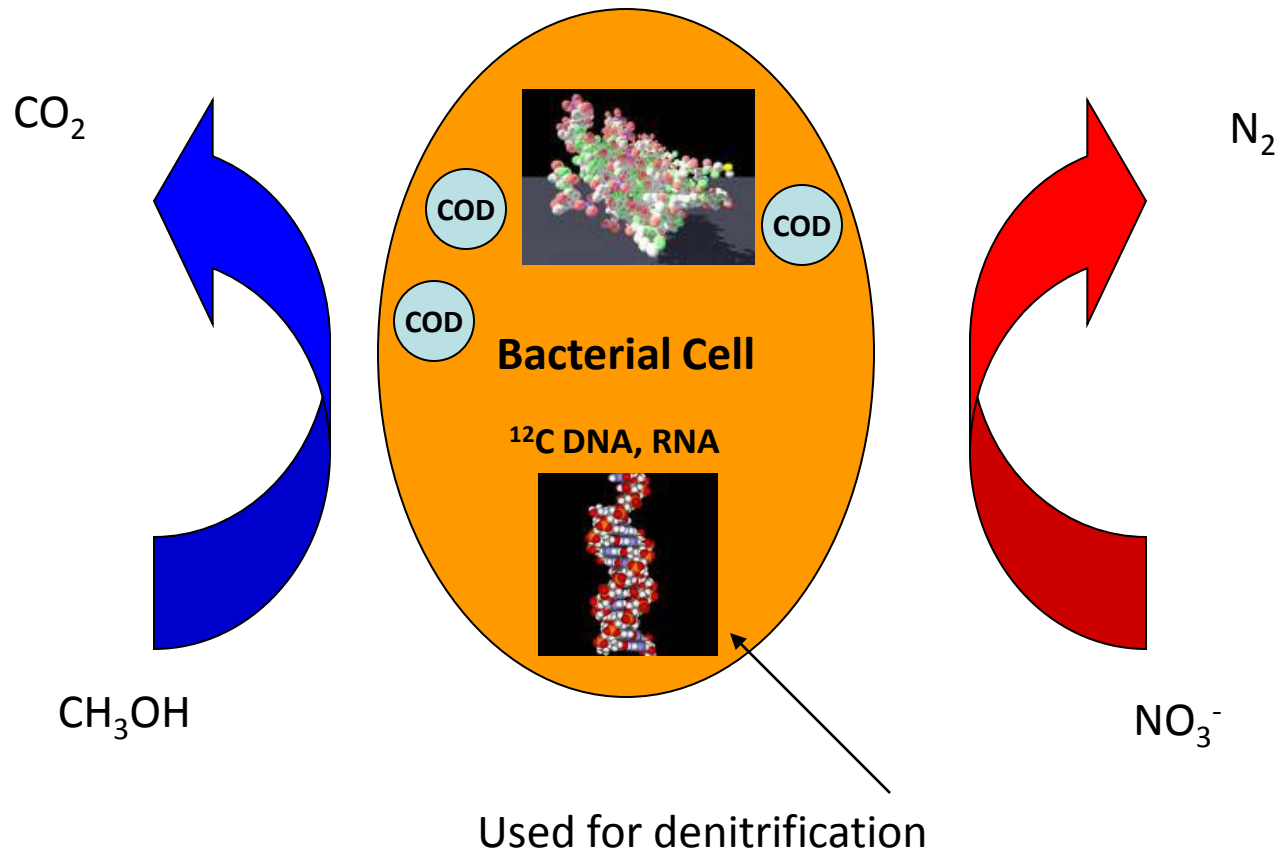
# Who consumes which organic electron donor in engineered denitrification



- Do same bacteria utilize all organic C- sources?
- What happens to community structure upon changing organic C-source?
- Implications for process modeling, design and optimization?

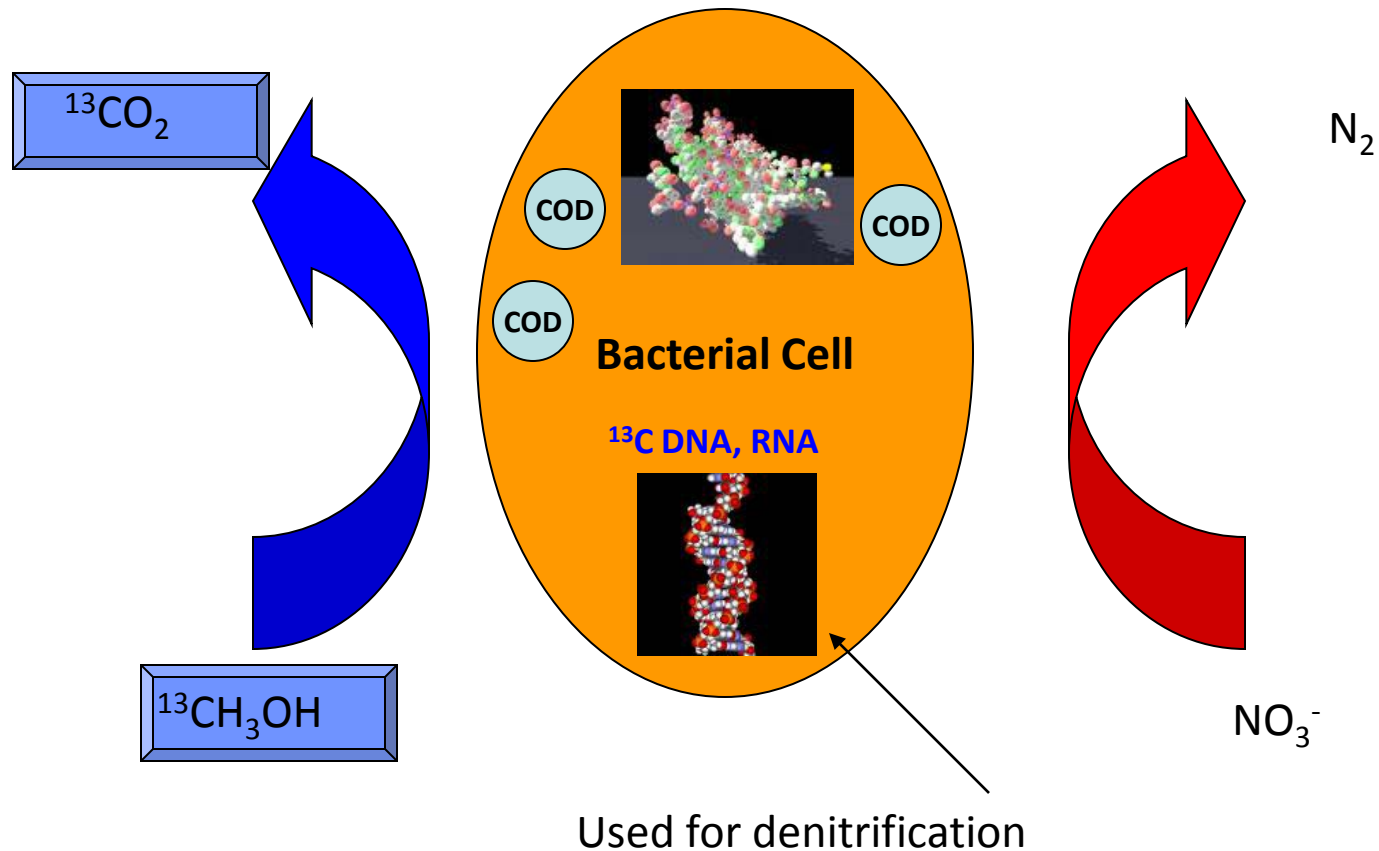


# Organic carbon uptake during denitrification



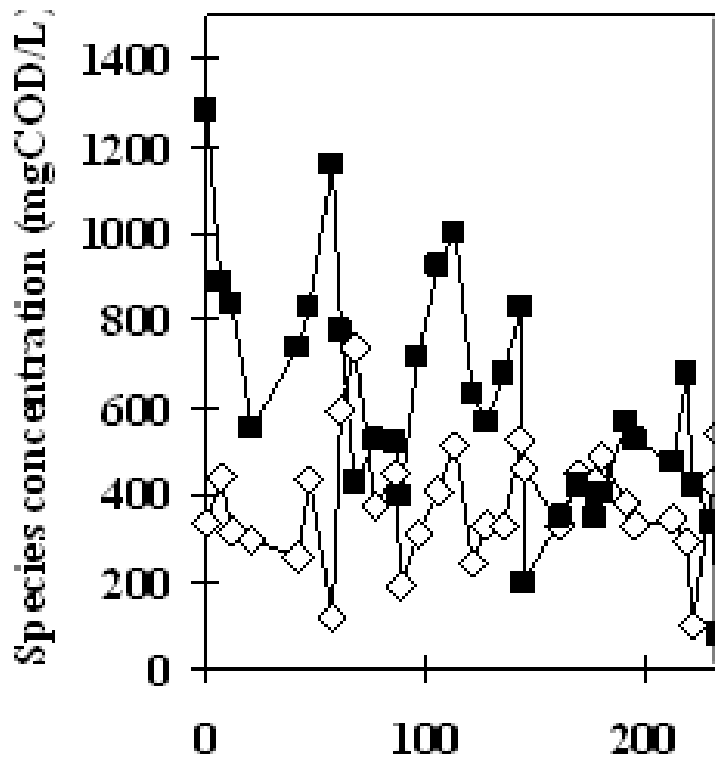


# Organic carbon uptake during denitrification



# Tracking dominant methylotrophic populations in the SBR

Phase I



Day of operation

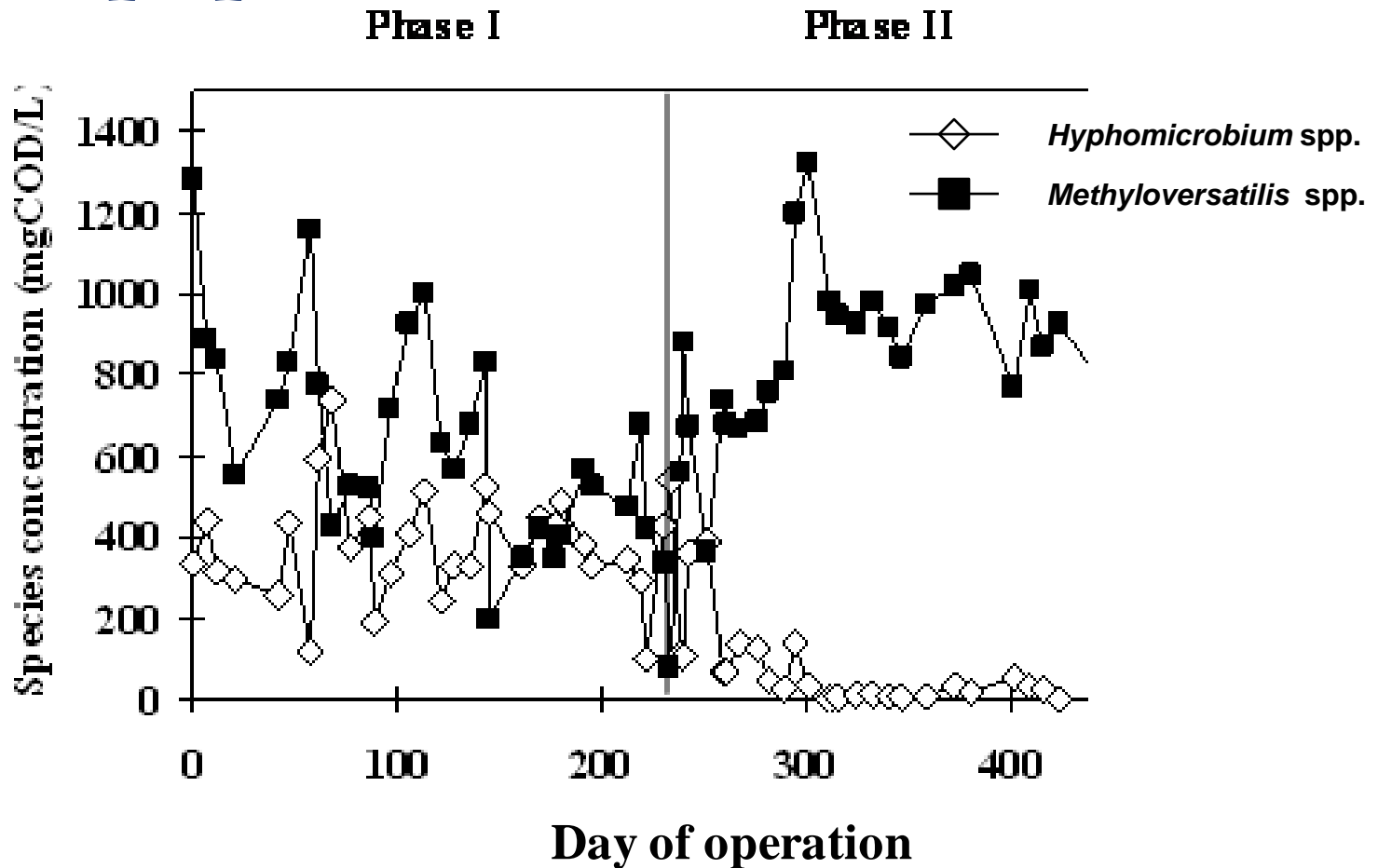
◇ *Hyphomicrobium* spp.

■ *Methyloversatilis* spp.

- *Methyloversatilis* spp. more abundant than *Hyphomicrobium* spp.
- ‘Relatively’ stable during methanol feed phase



# Tracking dominant methylotrophic populations in the SBR



Survival of methylotrophic populations depended upon their nutritional modes

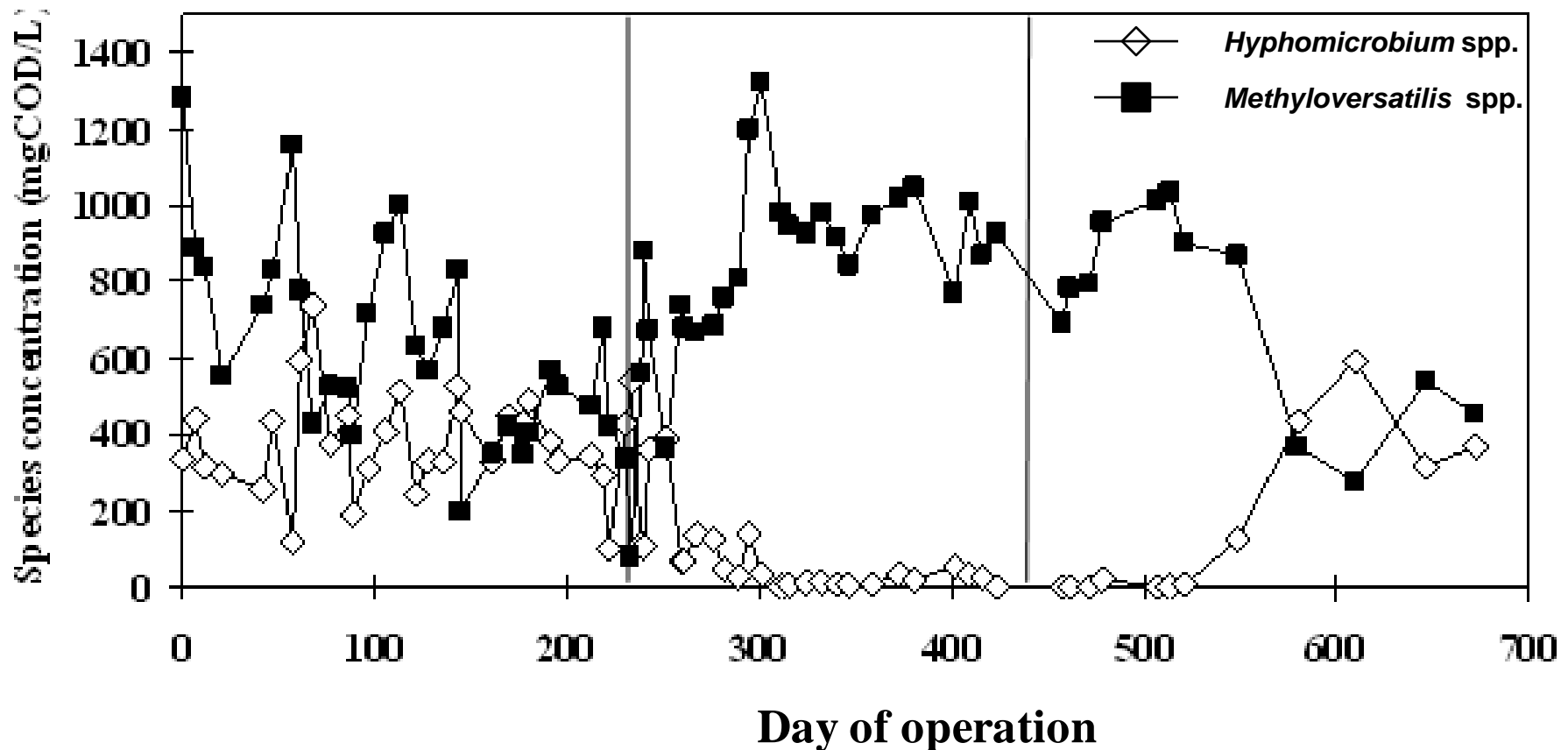


# Tracking dominant methylotrophic populations in the SBR

Phase I

Phase II

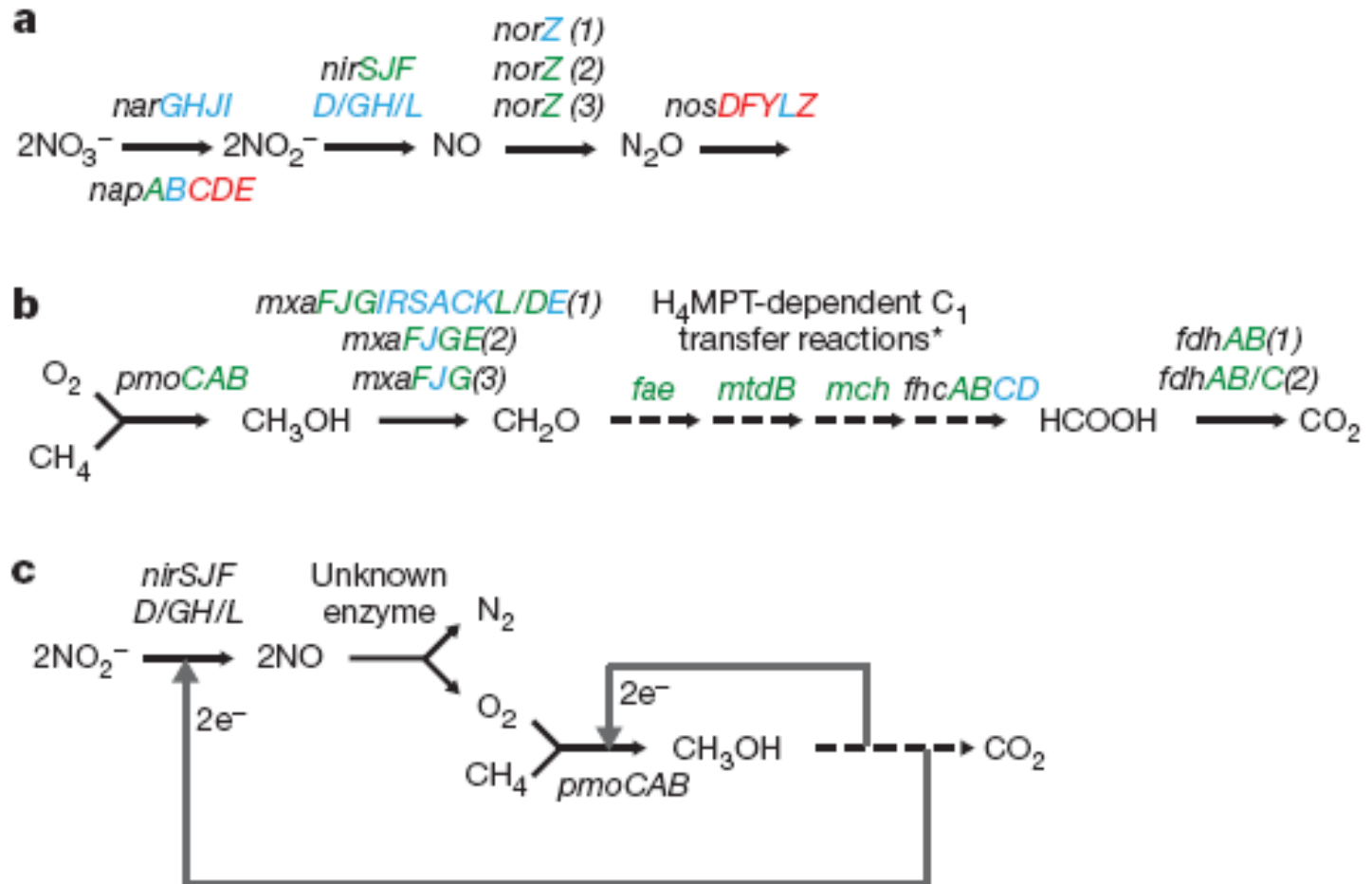
Phase III



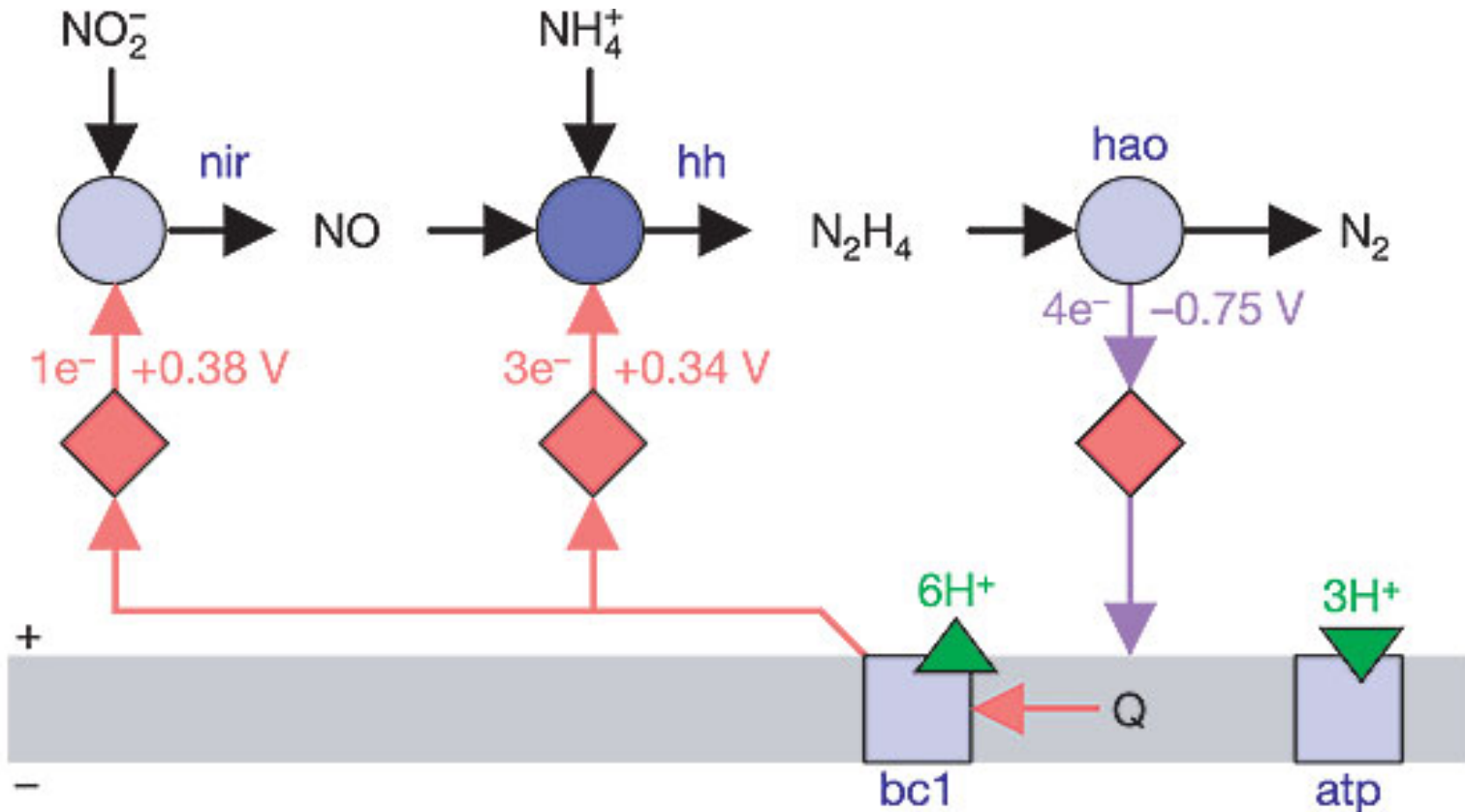
Survival of methylotrophic populations depended upon their nutritional modes



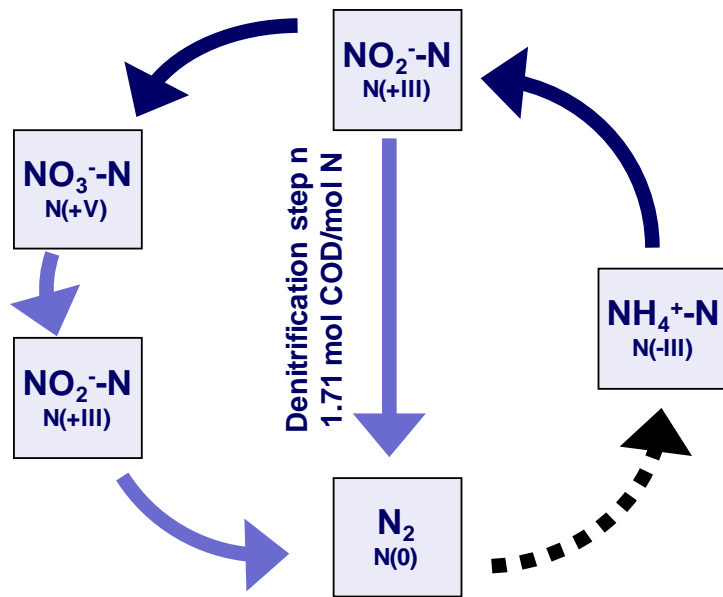
# Nitrite based anaerobic methane oxidation by *C. Methyloirabilis oxyfera*



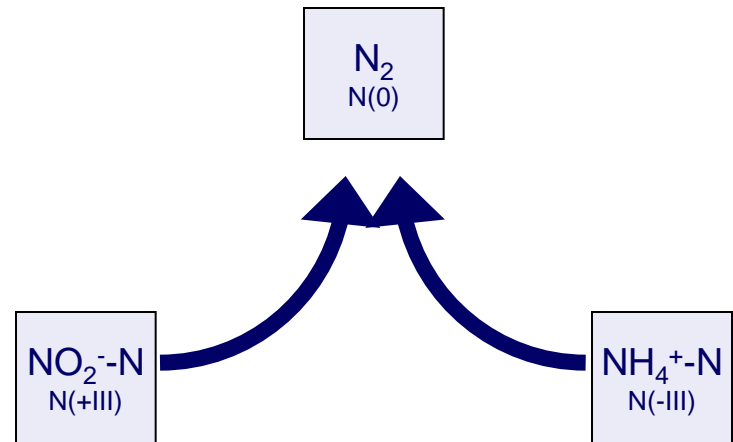
# Anaerobic ammonia oxidation



# Conventional BNR vs ANAMMOX



Conventional BNR

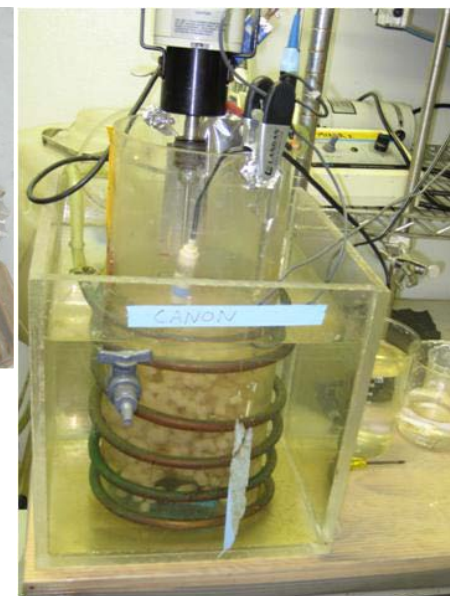
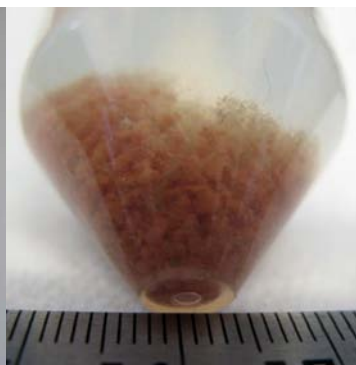


ANAMMOX

Only two pilot-scale applications of ANAMMOX in the U.S. ever



# Two bioreactor configurations



## Granular anammox

## Biofilm CANON

### Configuration

20 L SBR

4L Chemostat, 37.5% Kaldnes K1 media

### Operating conditions

HRT = 4d, SRT = 30d, completely anaerobic

HRT = 0.5d, Intermittent aeration (0.5min on/1min off)

### Seed

NYC anammox pilot plant + Strass, AT

Local activated sludge (Red Hook)

### Influent

$490 \pm 194$  mg  $\text{NH}_4^+\text{-N/L}$ ,  
 $518 \pm 222$ mg  $\text{NO}_2^-\text{-N/L}$

$615 \pm 164$  mg  $\text{NH}_4^+\text{-N/L}$

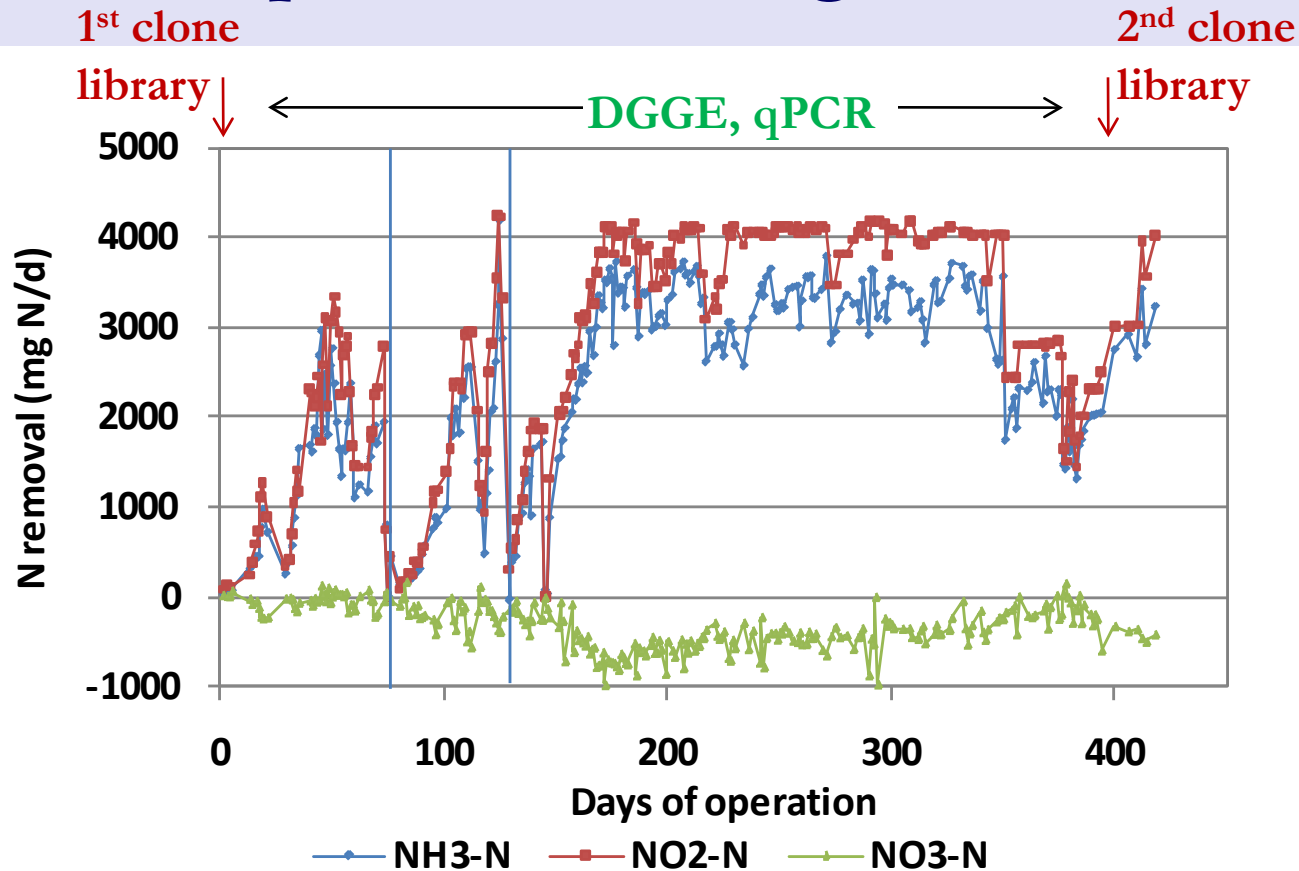
### N-Removal

$0.2$  kg  $\text{N/m}^3\text{/d}$ , 86~91%

$0.6\sim 1.3$  kg  $\text{N/m}^3\text{/d}$ , 55~88%



# N removal performance in granular anammox

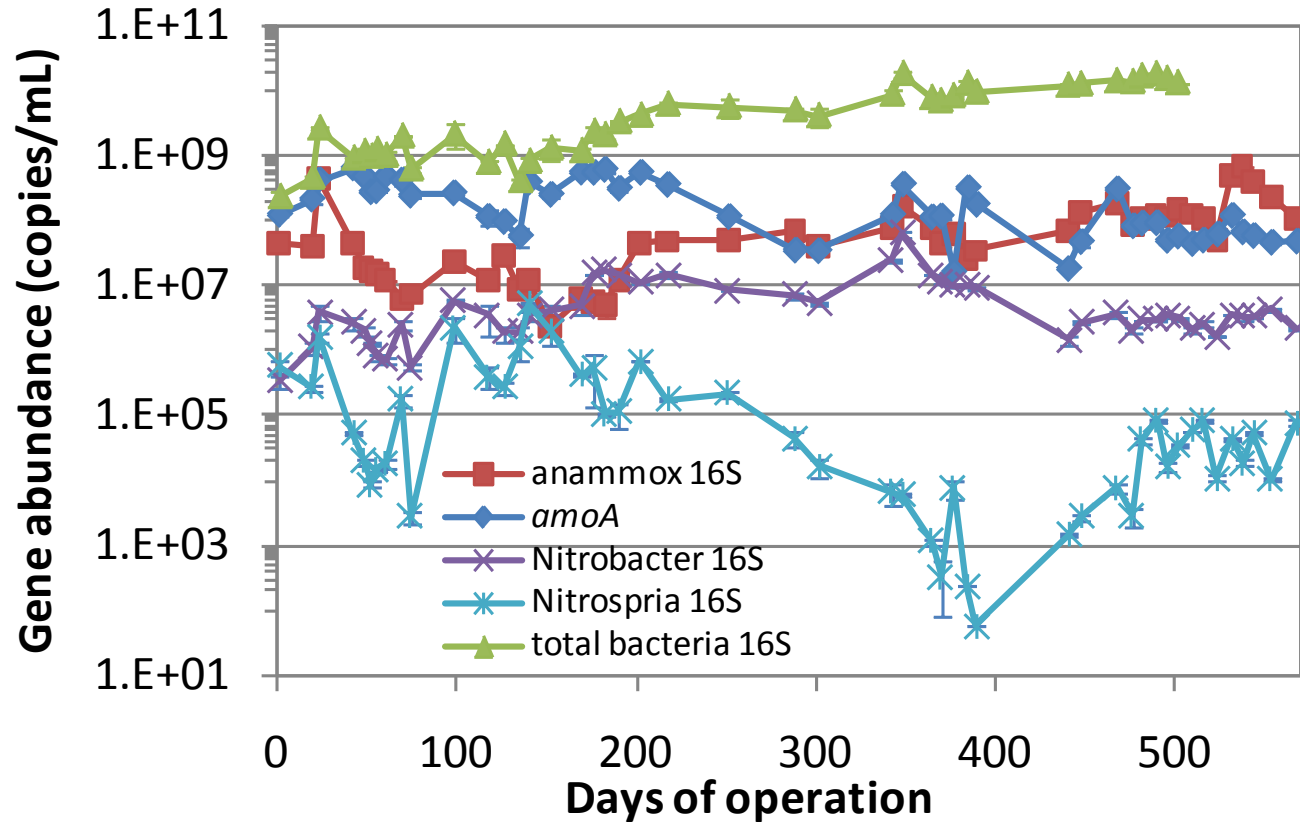


Park *et al.*, 2010, ES&T

- Stable operation accomplished for > 200 days with  $88 \pm 3$  % removal efficiency
  - Some transient upsets
- *Reactor performance gives little information on ecology and community abundance*



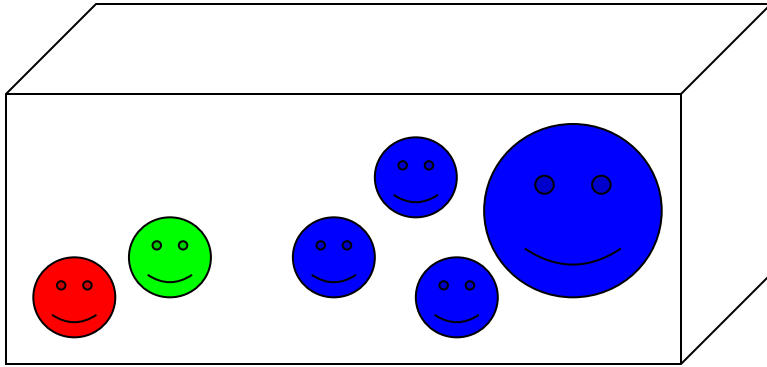
# AMX are not alone in anammox reactors



- Stable performance of anammox reflected in microbial ecology
  - Co-existence of AOB, NOB and AMX
- *Abundance gives little direct information on activity*



# Implications of activity on design

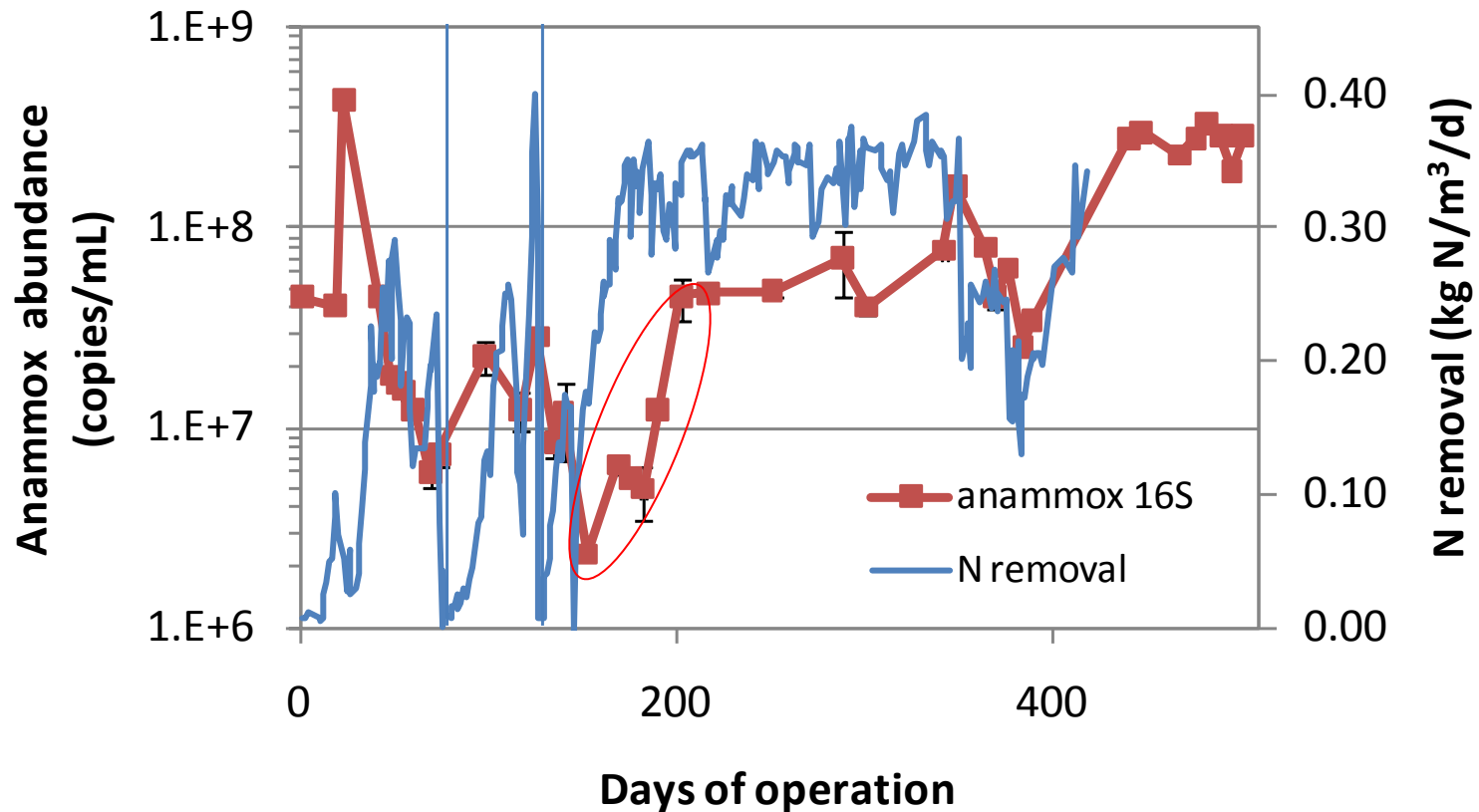


$$\Theta_{C,\min} = \frac{1}{\mu_{\max} - b}$$

- Traditional batch tests not applicable to estimate  $\mu_{\max}$ 
  - AOB, NOB, AMX all use  $\text{NO}_2^-$
  - AOB and AMX use  $\text{NH}_3$
  - Cannot infer anammox activity using  $\text{NH}_3$  or  $\text{NO}_2^-$  depletion profiles



# Estimates of activity from $X_{amx}$ conc.



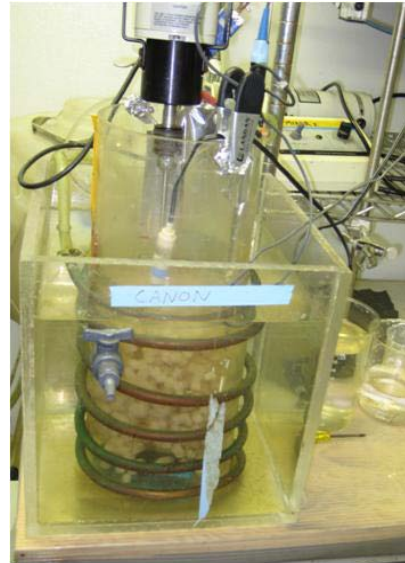
- Combination of  $X_{amx}$  with steady state mass balances to estimate  $\mu_{max}$

$$\ln X_{amx} = \ln X_{amx,0} + \mu \times t \rightarrow \mu_{max} = 0.11-0.15 \text{ d}^{-1}, t_d = 5.3 \text{ days}$$

- We don't want to rely on process upsets to estimate  $\mu_{max}$



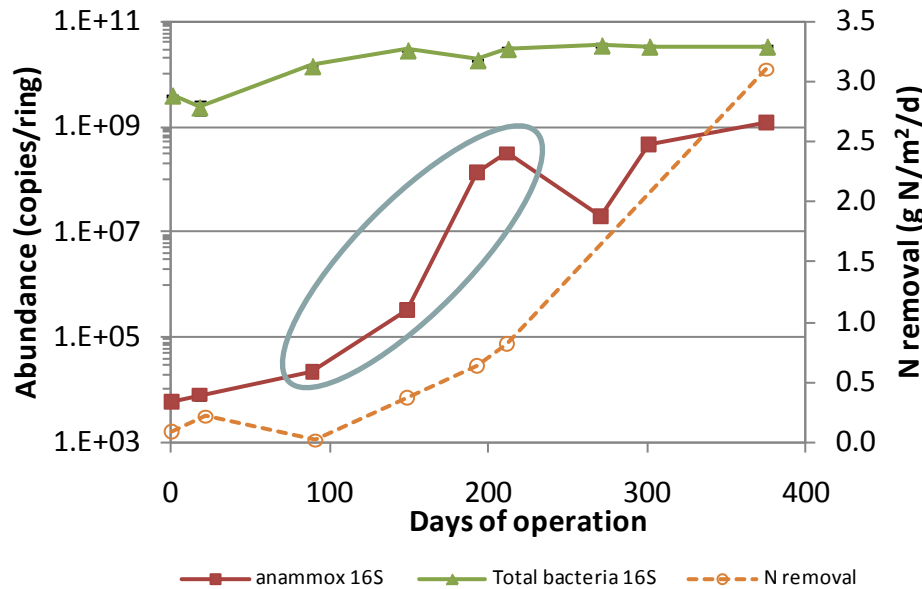
# Biofilm reactor set up



- Inoculum derived from activated sludge basin of Red Hook WPCP in New York City
- Virgin media seeded with activated sludge
- Fed with anaerobic digestion centrate from Ward's Island (no nitrite fed)
- 0.5 min air on, 2.0 min air off



# Measures of anammox activity

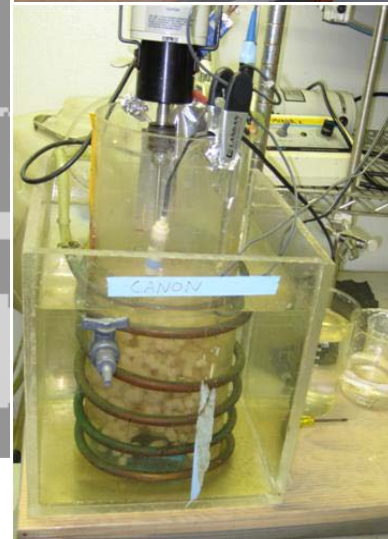
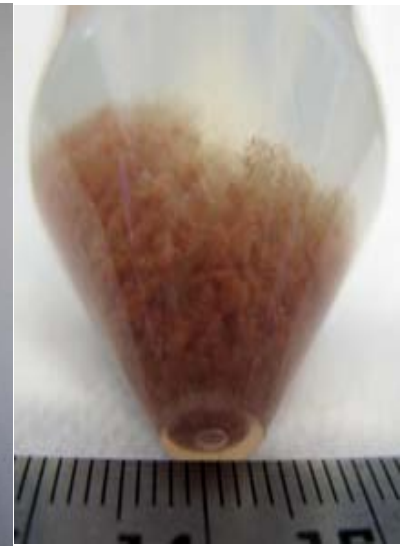
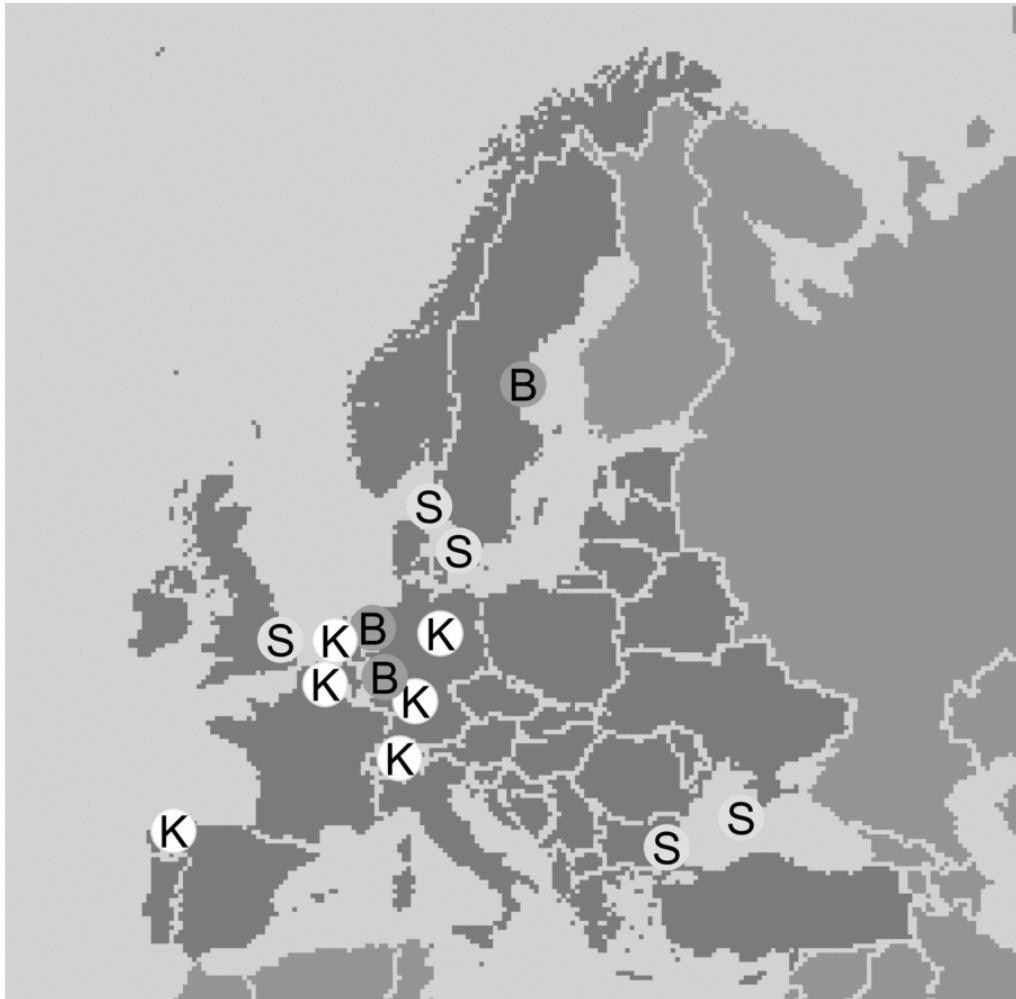


$$t_d = \frac{\ln(2)(t - t_o)}{\ln \frac{C}{C_o}}$$

- $t_d = 8.9$  days
- Another utility of directly measuring  $X_{AMX}$



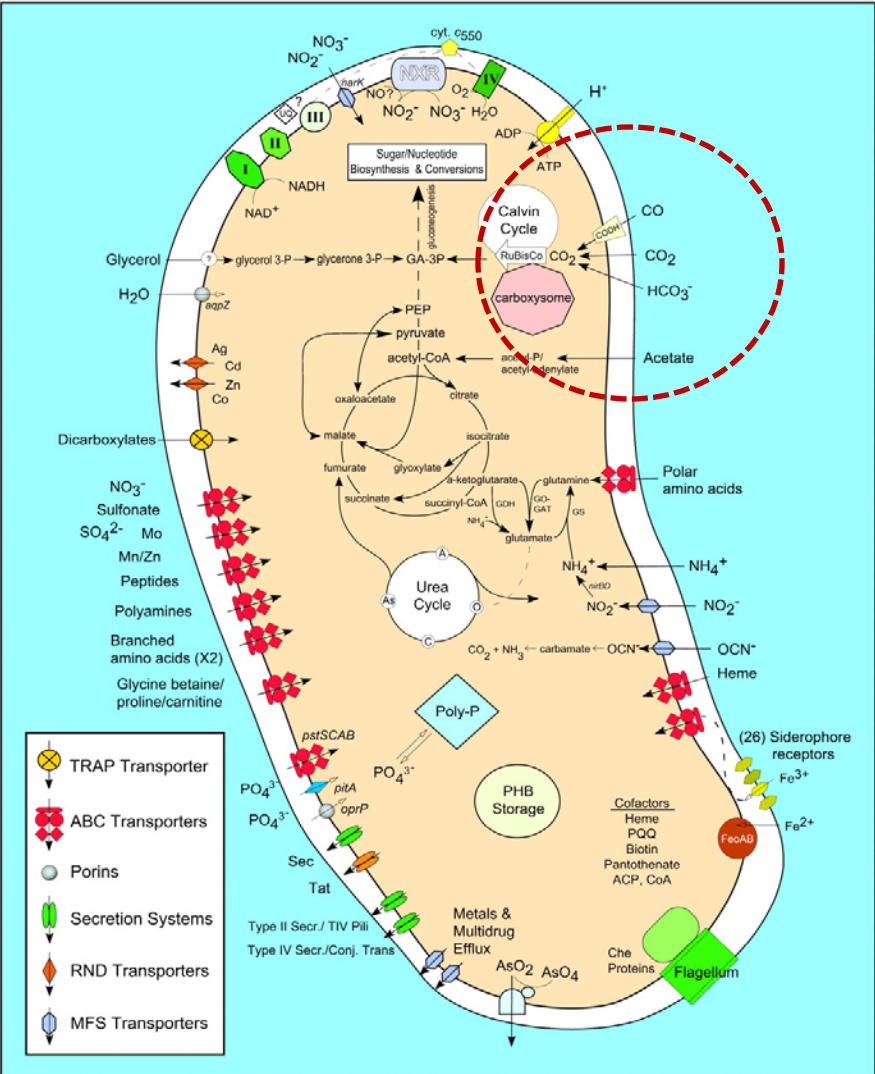
# How do microbial communities compare in different anammox configurations?



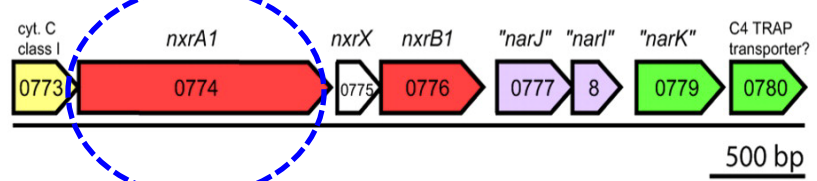
Kuenen et al., Biochemical Society Transactions (2005) 33, 119-123



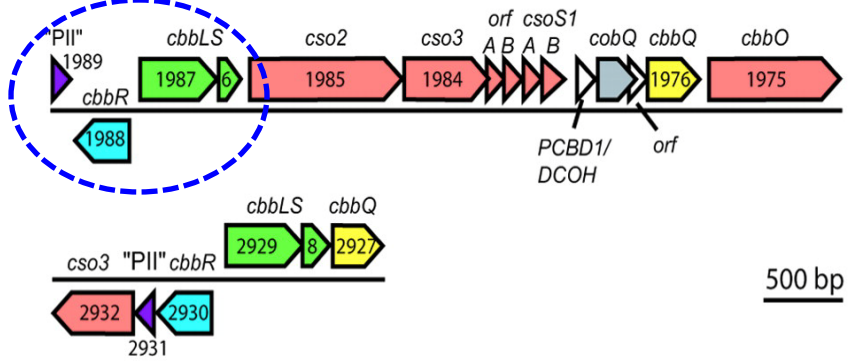
# Nitrobacter winogradskyi



## A. Nitrite oxidoreductase Cluster



## B. RuBisCO and Carboxysome Neighborhoods

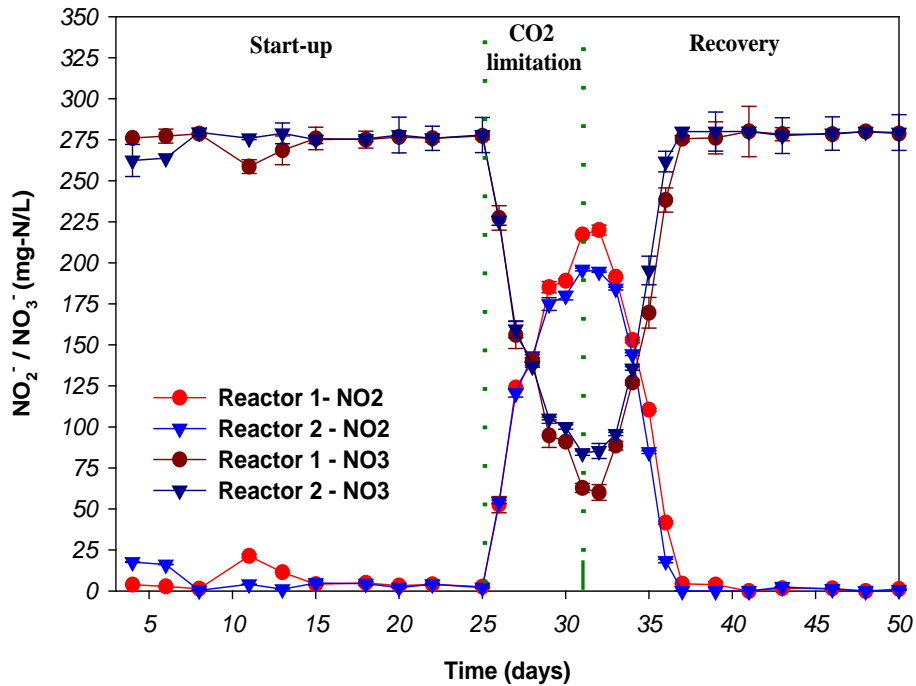




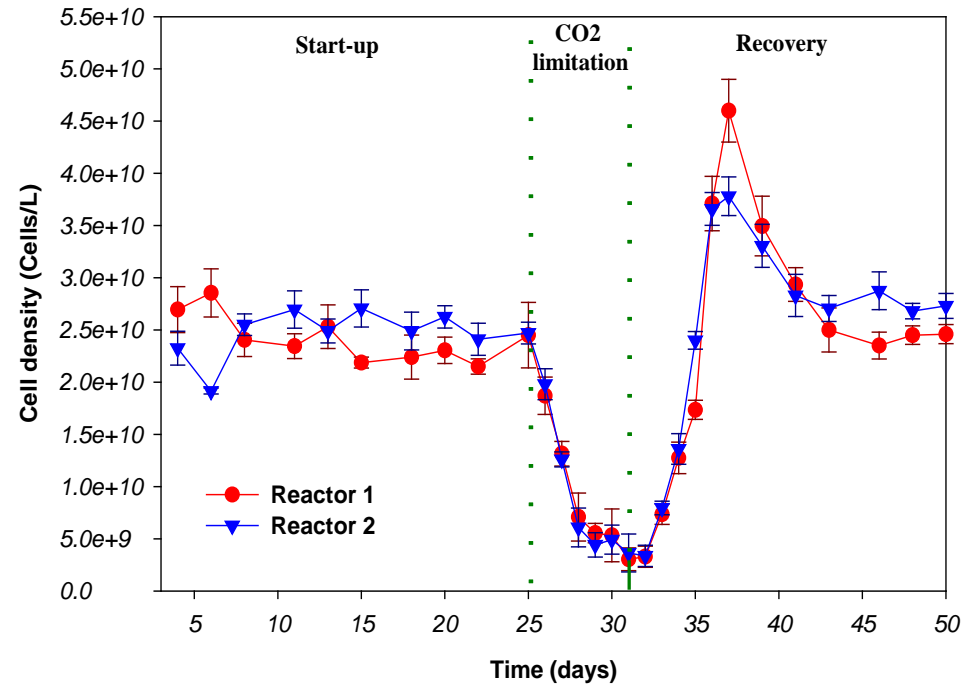
# The impact of CO<sub>2</sub> limitation on NOB

## (A) NOB performance

### (a) Nitrogen conversion



### (b) Cell concentration



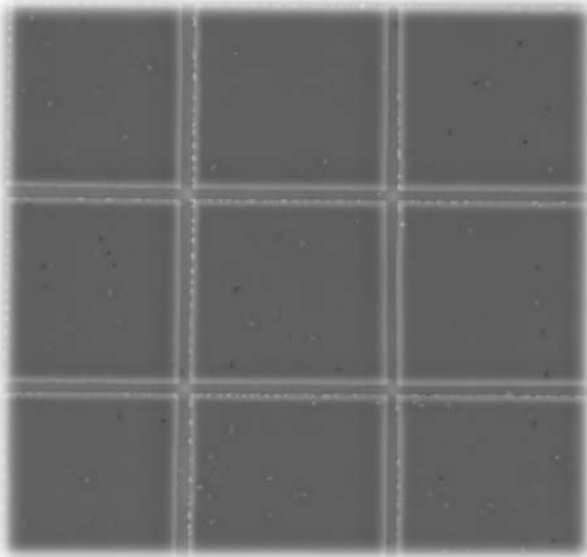
- CO<sub>2</sub> limitation → nitrite accumulation & a decrease in cell density
- A quick recovery occurred within 3 SRTs



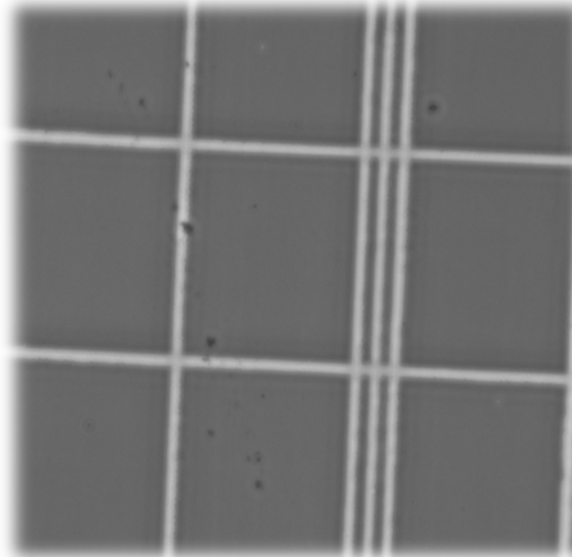
# The impact of CO<sub>2</sub> limitation on NOB

## (B) Cell pictures

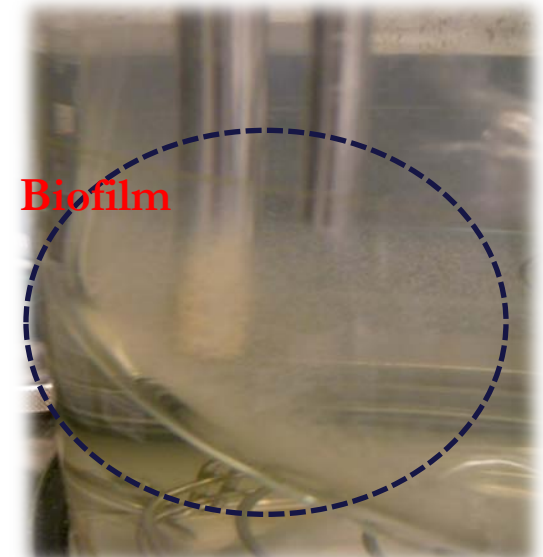
(a) Start-up



(b) CO<sub>2</sub> limitation (at 6 days)



(c) Recovery stage (Reactor)

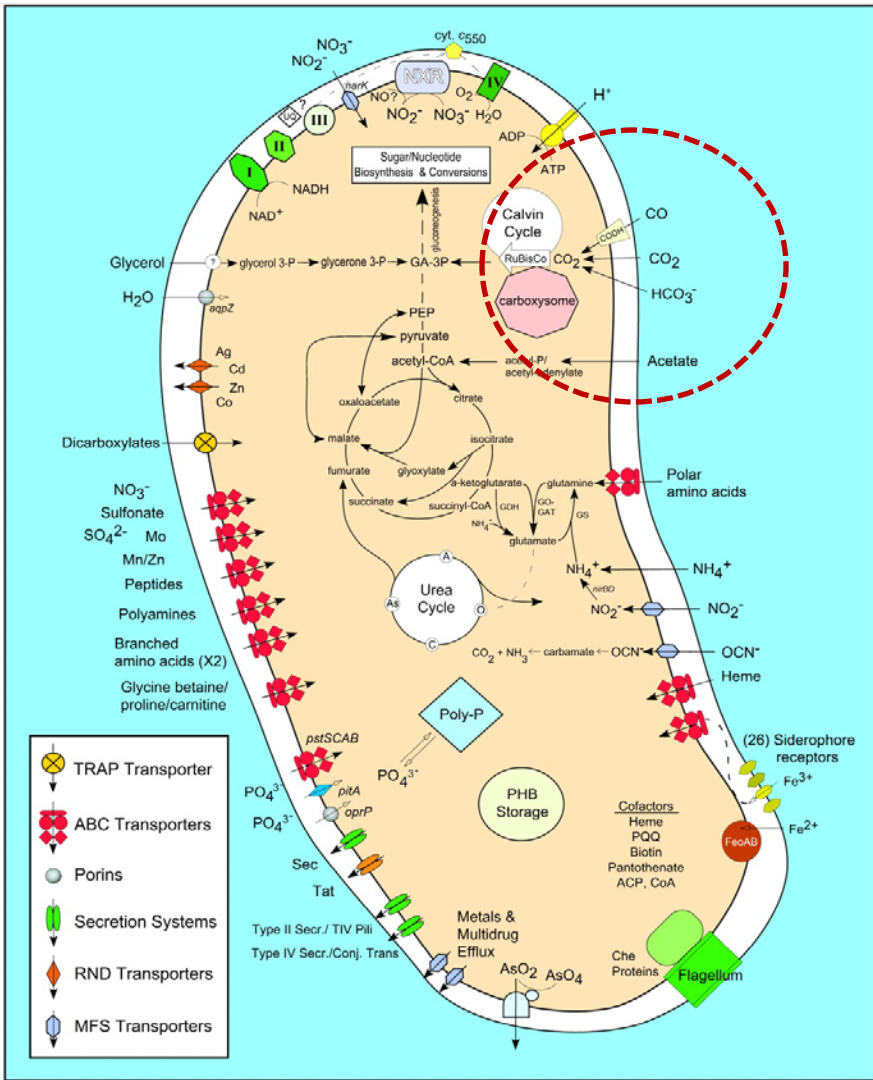


- During CO<sub>2</sub> limitation, cell clumps were observed
- Environmental stress might affect cell physiology & morphology



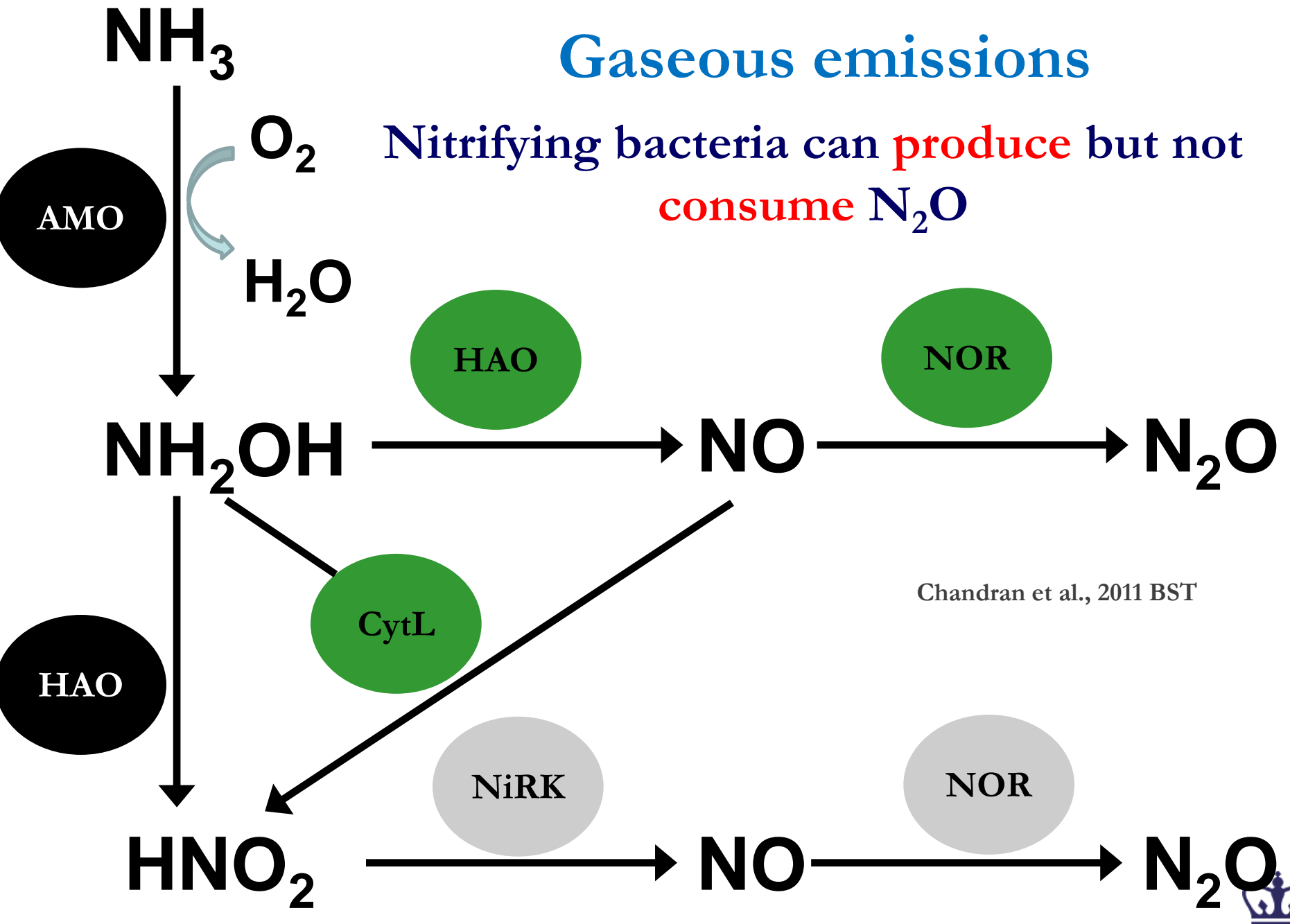
# How do Anammox reactor conditions impact NOB ?

- CO<sub>2</sub> supply
- Hydrazine
  - Anammox intermediate
- O<sub>2</sub> limitation
- Hydroxylamine

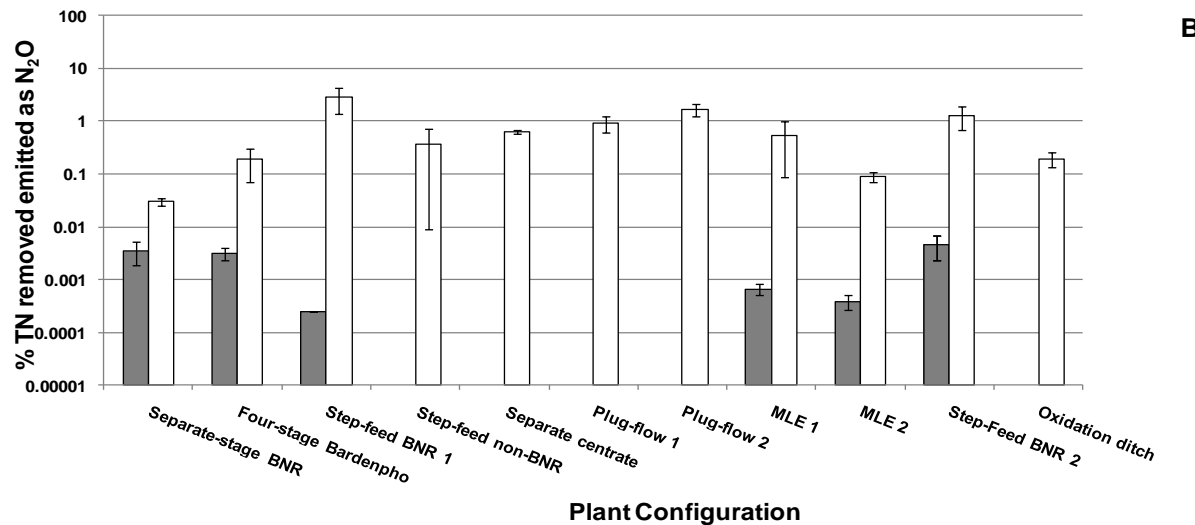
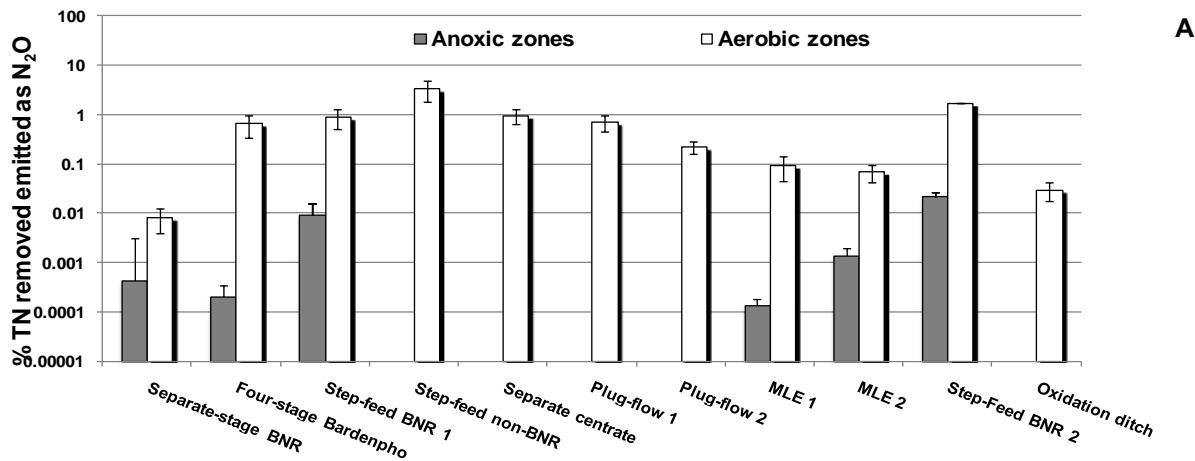


# Gaseous emissions

Nitrifying bacteria can **produce** but not **consume**  $N_2O$



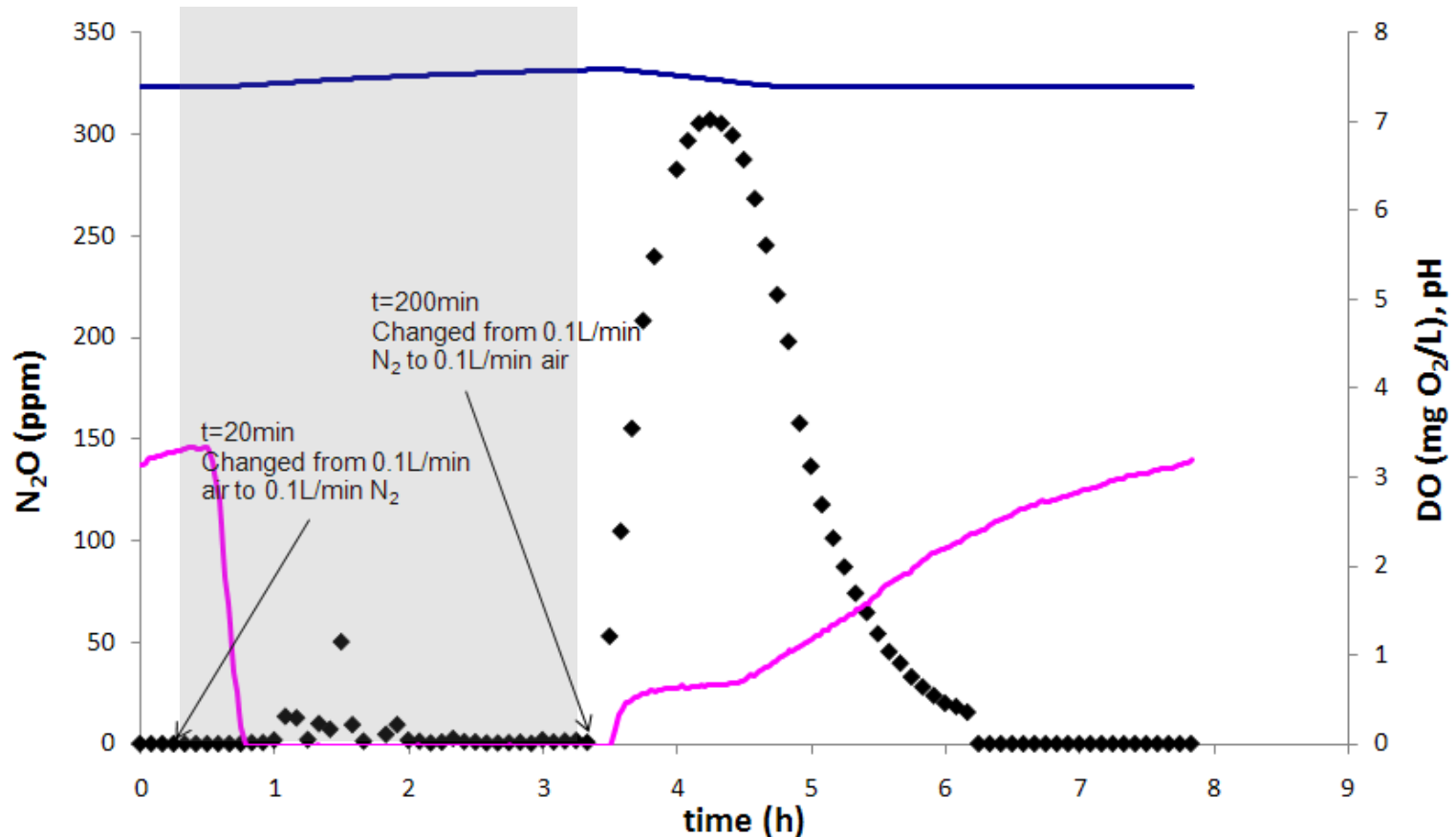
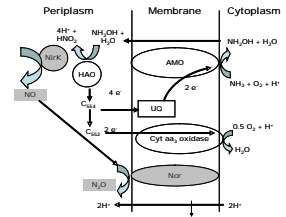
# Relative emissions from aerated and non-aerated zones



- Aerated zones contributed more to emissions than non-aerated zones



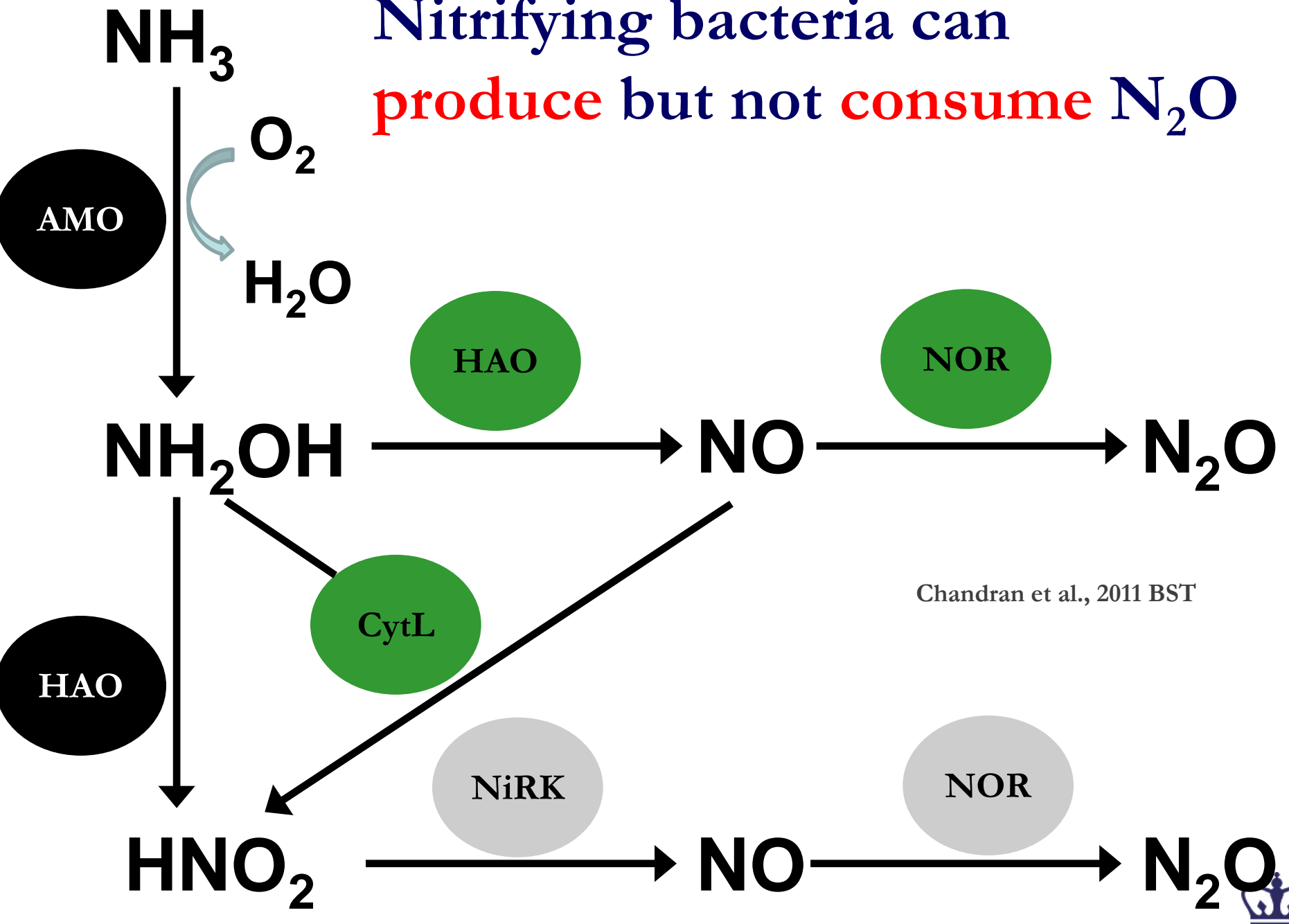
# Short term change in DO- Nitrification



- N<sub>2</sub>O production is directional
  - Manifestation of recovery response

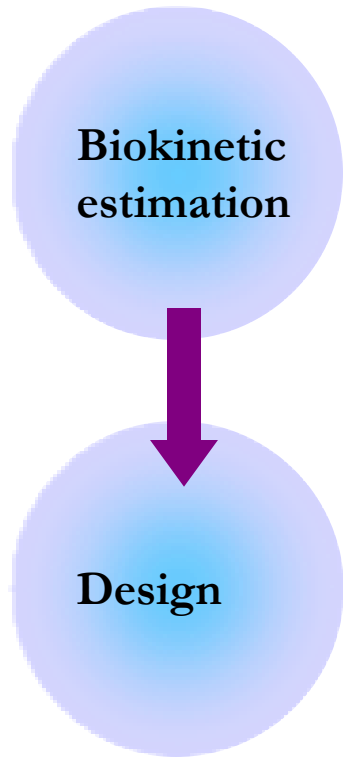


Nitrifying bacteria can  
**produce** but not **consume**  $N_2O$



Chandran et al., 2011 BST

# Why is this significant?



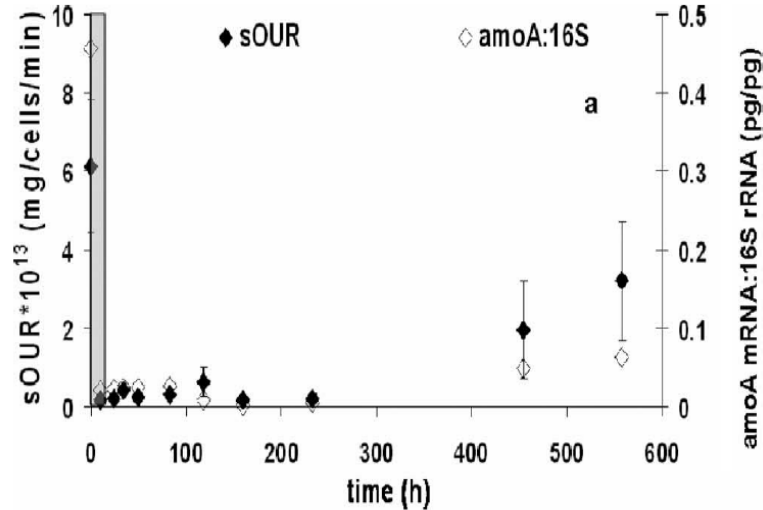
$$\mu_{\max} = \frac{f_s}{(1 - f_s)} * \frac{OUR_{\max}}{X}$$

$$S_{nh,eff} = \frac{K_s \left( \left( \frac{1}{\theta_c} \right) + b_a \right)}{\mu_{\max} - \left( \left( \frac{1}{\theta_c} \right) + b_a \right)}$$

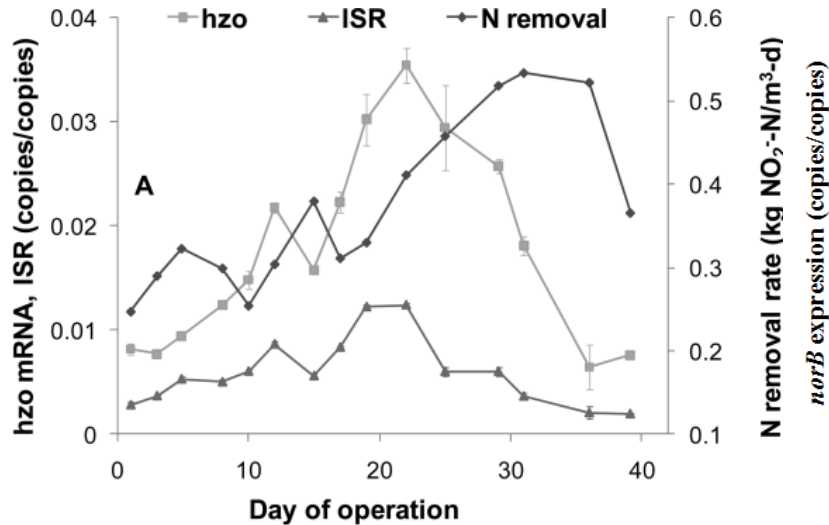
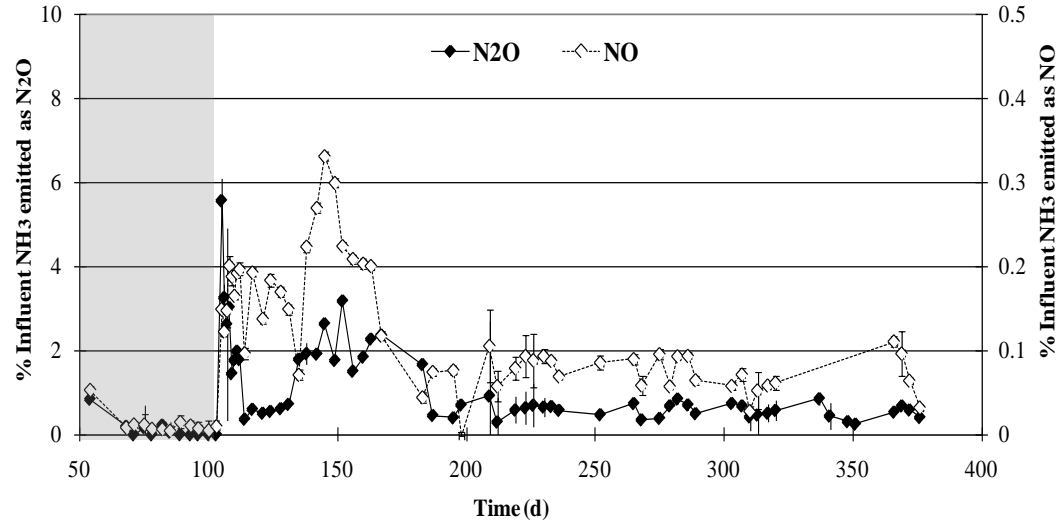




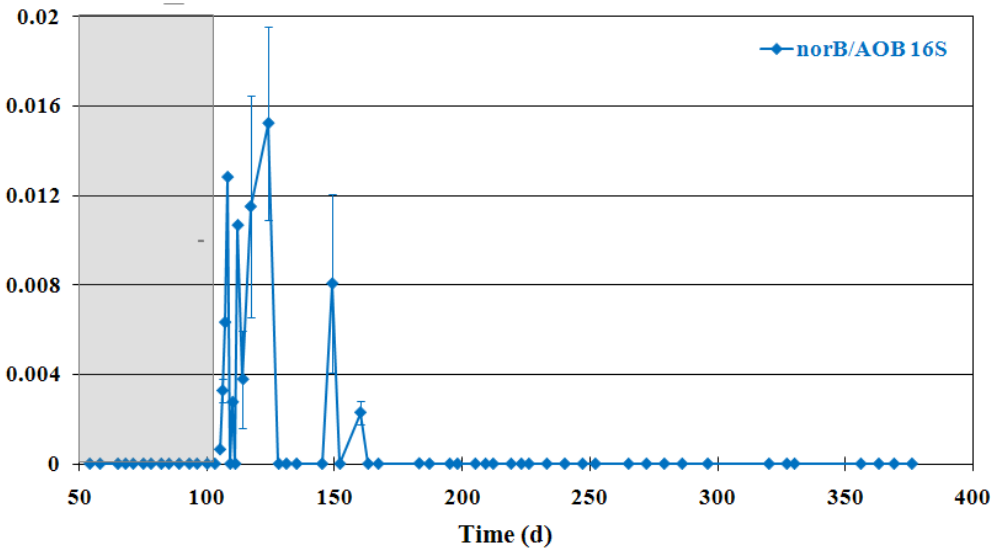
# Links between functional gene expression and activity



(Chandran and Love, 2008)



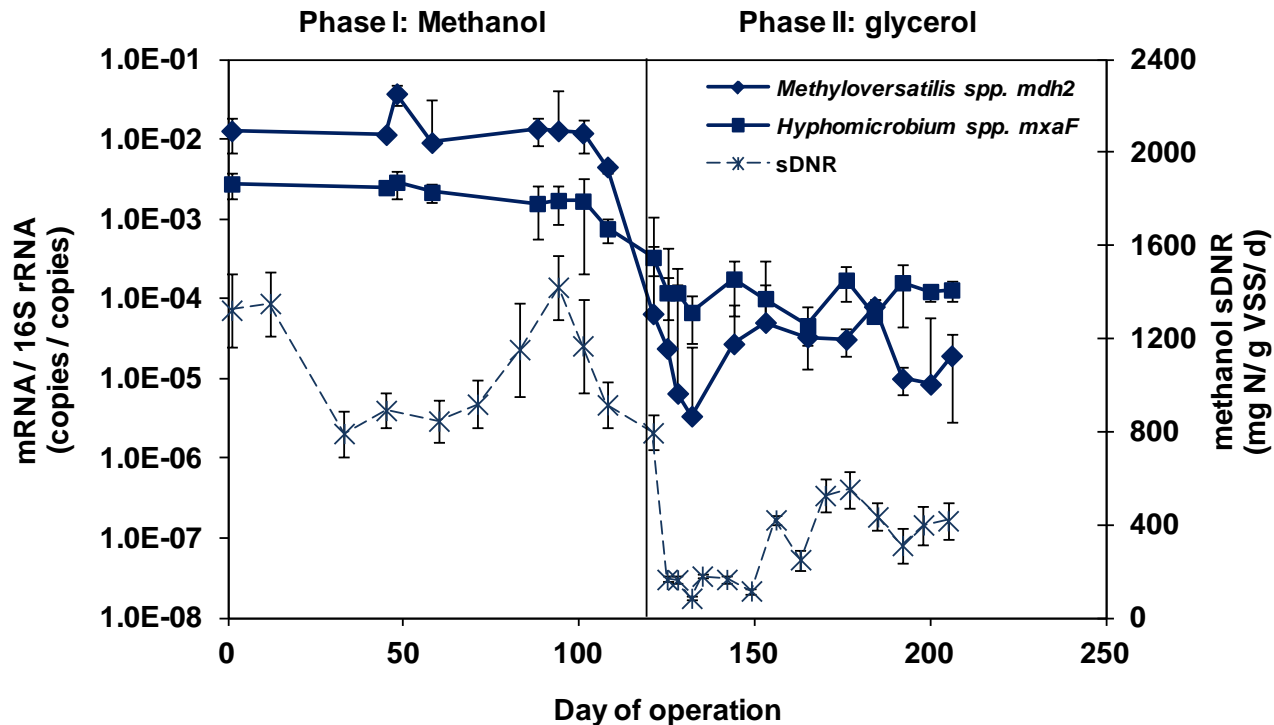
(Park et al., 2010)



(Ahn et al., 2011)



# Methanol dehydrogenase gene expression and methanol denitrification rates



- Significant positive correlations ( $\alpha=0.05$ ) between:
  - Methanol dehydrogenase gene transcription
  - Methanol sDNR values



# Contact information

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Phone: (212) 854 9027  
URL: [www.columbia.edu/~kc2288/](http://www.columbia.edu/~kc2288/)

